# THE EFFECT OF METABOLIC INHIBITORS ON THE LOCAL ACTION OF OESTRONE AND OESTRADIOL-3,17 $\beta$ ON THE VAGINA OF OVARIECTOMIZED MICE 

By P. J. Claringbold*

[Manuscript received June 24, 1953]


#### Abstract

Summary In the intravaginal Allen-Doisy test for oestrogenic activity potassium cyanide produced a significant increase in response to oestrone, but not to oestradiol-3,17 $\beta$.

Sodium azide, 2,4-dinitrophenol, and sodium monoiodoacetate caused a reduction in response to both oestrogens studied.

Inhibitory dose response lines obtained with sodium monoiodoacetate in mice receiving maximal doses of oestrone or oestradiol- $3,17 \beta$ indicate greater variability of response to oestradiol- $3,17 \beta$ than to oestrone.


## I. Introduction

While it has been known for some years that vaginal cornification in the rodent may be caused by local administration of oestrogens (see Emmens 1950, for review of the literature), the actual site and mode of action of the oestrogen in the epithelium have been the subject of speculation. It was thought that the effect of metabolic inhibitors, administered intravaginally, might help to elucidate this problem.

## II. Materials and Methods

A colony of 350 albino mice bred in this department were ovariectomized. Their management and use in tests were as described by Biggers (1951), tests being carried out fortnightly, with two injections in 24 hr . The oestrogens and inhibitors were administered together by the intravaginal route in phosphate buffer solution ( $\mathrm{pH} 7.0,0.1 \mathrm{M}$ ), so that the total volume of two injections was 0.02 ml . The oestrone and oestradiol $-3,17 \beta$ were obtained from Organon Laboratories.

Standard procedures for probit analysis, as described by Finney (1952) have been employed throughout. At the start of this work it was thought probable that significant slope differences would be found, particularly when different oestrogens were being compared (Biggers and Claringbold 1953; Biggers 1953a). The estimation of relative potency for the comparison of activity is only justified if the dose response lines are parallel and if this is not so the median effective dose (M.E.D.) ratio test must be used (Biggers 1951). If the final

[^0]Table 1
EFFEGT OF POTASSIUM GYANIDE ON THE DOSE RESPONSE LINES OBTAINED BY THE LOGAL ADMINISTRATION OF OESTRONE OR OESTRADIOL-3,17 $\beta$

partitioning of $\chi^{2}$ indicated no significant slope differences the relative potency estimate was computed, otherwise the more conservative M.E.D. ratio test was employed.

Response is recorded in the tables as a ratio of the number of animals positive to the number of animals per group. Standard errors of mean values follow a $\pm$ sign; fiducial limits of error ( $P=0.05$, except where otherwise indicated by subscript) have been enclosed in braces following the appropriate estimate; and M.E.D. ratios and relative potencies are given in their arithmetic form to avoid constant reference to the logarithmic base adopted in a particular test.

## III. Results

## (a) Potassium Cyanide (Table 1)

In the five tests reported $24 \mu \mathrm{~g}$ of potassium cyanide were injected with the oestrogen in buffer solution, three tests being carried out with oestrone and two with oestradiol $-3,17 \beta$. Where oestrone was the oestrogen under test, a significant increase in response occurred on all occasions. The partitioning of $\chi^{2}$ gives no evidence of any significant slope differences, and while the data obtained with oestrone show evidence of heterogeneity, this has been ignored since the total $\chi^{2}$ for heterogeneity over all the tests reported in this paper is well within the range of random sampling variation ( $\left.\chi^{2}{ }_{(54)}=50.67, P=0.59\right)$.

## (b) Sodium Monoiodoacetate (Tables 2 and 3)

The tests shown in Table 2 were based on the results of preliminary investigations and were designed for the purpose of comparing the inhibition produced by monoiodoacetate in mice receiving a maximal dose of (1) oestrone and (2) oestradiol $-3,17 \beta$. The dose response lines have negative slopes and may be used to estimate the median inhibitory dose (M.I.D.), i.e. the dose required to prevent cornification in 50 per cent. of animals. Although equal amounts of oestrone and oestradiol-3,17 $\beta$ were given to both groups ( $20 \times 10^{-4}$ $\mu \mathrm{g}$ ), the expected response in the absence of inhibition is in each approximately 98 per cent. Under all conditions of administration oestradiol- $3,17 \beta$ has been found more variable in action than oestrone (Biggers and Claringbold 1953), with the result that the expected responses to doses of approximately $20 \times 10^{-4}$ $\mu \mathrm{g}$ are equal (cf. Biggers and Claringbold 1953, Fig. 3).

Analysis of the results of Table 2 indicates that in order to produce 50 per cent. inhibition significantly more monoiodoacetate is required with oestra-diol- $3,17 \beta$ than with oestrone. Partitioning of $\chi^{2}$ indicates that the slope of the inhibitory dose response line for the latter is significantly steeper than for the former.

A simple factorial experiment (Table 3) was used to test the hypothesis that the activities of oestrone and monoiodoacetate are additive, i.e. to test whether the variability of response to oestrone is independent of the level of monoiodoacetate used. The results of the experiment have been fitted by means of a probit plane (Finney 1952), the regression equation involving the
probit of response, the log dose of oestrone, the log dose of monoiodoacetate, and a constant. The goodness of fit of this equation was found to be satisfactory $\left(\chi^{2}{ }_{(7)}=2.68,0.95>P>0.9\right)$. There is thus no evidence of departure from additivity and it may be concluded that the variability of response to oestrone is independent of the level of monoiodoacetate.

Table 2
EFFECT OF FOUR CONGENTRATIONS OF MONOIODOACETATE ON MAXIMAL CORNIFICATION PRODUCED BY $20 \times 10^{-4} \mu \mathrm{~g}$ OF OESTRONE OR OESTRADIOL- $3,17 \beta$

Twenty animals per group


* Median inhibitory dose.

The data of this test have been analysed by alternative methods for purposes of comparison; the results of this investigation are published elsewhere (Claringbold, Biggers, and Emmens 1953). With these alternative procedures the same conclusions have been reached.
(c) Sodium Azide (Table 4) and 2,4-Dinitrophenol (Table 5)

In the tests reported $60 \mu \mathrm{~g}$ of sodium azide or $600 \mu \mathrm{~g}$ 2,4-dinitrophenol were injected with the oestrogen in buffer solution. The inhibitors always
caused a significant lowering of response to the oestrogens. The partitionings of $\chi^{2}$ show no significant slope differences.

Table 3
EFFEGT OF GOMBINATIONS OF LOGALLY ADMINISTERED DOSES OF MONOIODOACETATE AND OESTRONE

Twenty animals per group is constant; number positive tabulated

|  | Monoiodoacetate Dose <br> $(\mu \mathrm{g})$ |  |  |  |
| :---: | :---: | :---: | :---: | :---: |
| Oestrone <br> Dose <br> $\left(10^{-4} \mu \mathrm{~g}\right)$ | $12 \cdot 5$ | 25 | 50 | 100 |

Mean slopes:
Oestrone: $1 \cdot 17 \pm 0 \cdot 24$
Monoiodoacetate: $-1 \cdot 19 \pm 0 \cdot 24$

## IV. Discussion

Following the intravaginal administration of a dose of oestrogen, the changes which take place may be divided into two stages. In the first stage the oestrogen is absorbed by the epithelium and transferred to the final site of action, perhaps undergoing transformation on the way. If the dose is sufficient the second stage begins, involving morphological changes associated with greatly increased mitotic rate in the cells of the stratum germinativum. Metabolic inhibitors or activators may act on either of these stages; if acting at the first stage their effect may depend on the oestrogen used, whereas if acting at the second, their effect should be independent of the oestrogen used.

Evidence for the local conversion of oestrone to oestradiol- $3,17 \beta$ has been presented in an earlier paper by Biggers and Claringbold (1953), who have also pointed out the fundamental difference in variability of response to the local administration of these oestrogens. The differential action of cyanide with regard to oestrone and oestradiol $-3,17 \beta$ cannot be explained simply in terms of a local metabolic action taking place in the second stage, but must indicate that oestrone and oestradiol- $3,17 \beta$ undergo different changes in reaching their site of action.

Bullough and Johnson (1951) have studied the effect of various metabolic inhibitors on mitotic activity in adult mouse epidermis and found that cyanide, azide, monoiodoacetate, and 2, 4-dinitrophenol are powerful mitotic inhibitors acting in the antephase. The histological and cytological aspects of the vaginal
Table 4
effect of sodium azide on the dose response lines obtained by the local administration of oestrone or oestradiol-3,17 $\beta$


* A common M.E.D. for the two treated groups was obtained and tested against the control M.E.D.
Table 5
EFFEGT OF 2, 4-DINITROPHENOL ON THE DOSE RESPONSE LINES OBTAINED BY THE LOGAL ADMINISTRATION OF OESTRONE OR OESTRADIOL-3,17 $\beta$

| Oestrone |  |  |  |  |  | Oestradiol-3,17 $\beta$ |  |  |  |  |  |
| :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: |
| Test | Control |  | Treated |  | M.E.D. <br> Ratio | Test | Control |  | Treated |  | M.E.D. Ratio |
|  | $\begin{gathered} \text { Dose } \\ \left(10^{-4} \mu \mathrm{~g}\right) \end{gathered}$ | Response | $\begin{gathered} \text { Dose } \\ \left(10^{-4} \mu \mathrm{~g}\right) \end{gathered}$ | Response |  |  | Dose $\left(10^{-4} \mu \mathrm{~g}\right)$ | Response | $\begin{gathered} \text { Dose } \\ \left(10^{-4} \mu \mathrm{~g}\right) \end{gathered}$ | Response |  |
| 1 | $1 \cdot 0$ $2 \cdot 3$ $5 \cdot 3$ $12 \cdot 2$ | $\begin{aligned} 3 & : 20 \\ 7 & : 20 \\ 12 & : 20 \\ 17 & : 20\end{aligned}$ | $1 \cdot 0$ $2 \cdot 3$ $5 \cdot 3$ $12 \cdot 2$ | $0: 20$ $3: 20$ $8: 20$ $11: 20$ | $\begin{gathered} 0 \cdot 41 \\ \{0 \cdot 23-0 \cdot 69\} \end{gathered}$ | 1 | $1 \cdot 0$ $2 \cdot 3$ $5 \cdot 3$ $12 \cdot 2$ | $6: 20$ $6: 20$ $12: 20$ $15: 20$ | $\begin{array}{r} 1 \cdot 0 \\ 2 \cdot 3 \\ 5 \cdot 3 \\ 12 \cdot 2 \end{array}$ | $\begin{aligned} 5: & : 20 \\ 2: & 20 \\ 9: & 20 \\ 13: & 20 \end{aligned}$ | $\begin{gathered} 0 \cdot 50 \\ \{0 \cdot 21-1 \cdot 03\} \end{gathered}$ |
| 2 | $1 \cdot 0$ $2 \cdot 3$ $5 \cdot 3$ $12 \cdot 2$ | $2: 15$ $4: 19$ $9: 20$ $16: 20$ | $1 \cdot 0$ $2 \cdot 3$ $5 \cdot 3$ $12 \cdot 2$ | $0: 15$ $2: 20$ $6: 20$ $10: 20$ | $\begin{gathered} 0 \cdot 48 \\ \{0 \cdot 24-0 \cdot 94\} \end{gathered}$ | 2 | $1 \cdot 0$ $2 \cdot 3$ $5 \cdot 3$ $12 \cdot 2$ | $7: 20$ $9: 20$ $13: 20$ $16: 20$ | $\begin{array}{r} 1 \cdot 0 \\ 2 \cdot 3 \\ 5 \cdot 3 \\ 12 \cdot 2 \end{array}$ | $\begin{aligned} & 2: 20 \\ & 5: 20 \\ & 10: 20 \\ & 11: 20 \end{aligned}$ | $\begin{gathered} 0 \cdot 32 \\ \{0 \cdot 13-0 \cdot 78\} \end{gathered}$ |
| Mean slopes $1.91 \pm 0.31$ |  |  | $1 \cdot 98 \pm 0 \cdot 36$ |  |  |  | $1 \cdot 18 \pm 0 \cdot 26$ |  | $1 \cdot 25 \pm 0 \cdot 27$ |  |  |
| Common mean slope 1.94 $\pm 0 \cdot 23$ |  |  |  |  |  | $1 \cdot 22 \pm 0 \cdot 19$ |  |  |  |  |  |
| $\overline{\text { Partitioning of }} \overline{\chi^{2}}$    <br> Source of variation <br> Parallelism D.f. $\chi^{2}$ $P$ |  |  |  |  |  | Source of variation Parallelism |  |  | D.f. | $\chi^{2}$ | $P$ |
|  |  |  |  |  |  |  |  |  |  |
| Between mean slope |  |  | 1 | 0.01 | 0.95-0.90 |  |  |  | Between mean slopes |  |  | 1 | $0 \cdot 03$ | 0-9-0•8 |
| Within mean slopes |  |  | 2 | $0 \cdot 01$ | 0.98-0.95 | Within mean slopes |  |  | 2 | $0 \cdot 05$ | 0.98-0.95 |
| Heterogeneity |  |  | 6 | $3 \cdot 41$ | 0.8-0.7 | Heterogeneity |  |  | 8 | $7 \cdot 08$ | 0.7-0.5 |
| $\{0 \cdot 28-0 \cdot 69\}$ |  |  |  |  |  | $0 \cdot 40$ |  |  |  |  |  |
| Relative potency $0 \cdot 44$ (0.29-0.66\} |  |  |  |  |  | $\begin{aligned} & 0.41 \\ & \{0 \cdot 22-0 \cdot 74\} \end{aligned}$ |  |  |  |  |  |

response to oestrogens in mice have been discussed by Allen (1922) and Biggers (1953b), the first sign of oestrogen activity being division of the cells of the stratum germinativum. In the present work it would appear that azide, monoiodoacetate, and 2, 4-dinitrophenol are exerting their characteristic metabolic effect (inhibition of respiration in the first case, inhibition of glycolysis in the last two cases), resulting in inhibition of the morphological response. Bullough and Johnson (1951), however, found that cyanide acted similarly to azide whereas in the work described in this paper no inhibition occurred with the level of dosage used. In the present work the action of cyanide may be a specific enzymic effect. Heller (1940) showed that incubation of oestrone with liver mince resulted in inactivation of the oestrone unless cyanide was added, when oestrone was converted to oestradiol- $3,17 \beta$ with resultant increase in activity.

Further evidence has been presented as to the fundamental differences in activity and variability of oestrone and oestradiol-,$- 17 \beta$ when administered by the intravaginal route. The slopes of the inhibitory dose response lines reflect the variability in response to the oestrogens, and indicate that oestradiol-3,17 $\beta$ elicits the more variable response. The fact that more inhibitor is required to produce 50 per cent. inhibition with oestradiol-3,17 $\beta$ than with oestrone demonstrates the greater relative activity of oestradiol-3,17 $\beta$.

The present work, taken in conjunction with that of Bullough and Johnson (1951), indicates that oestrogens stimulate mitosis in the cells of the stratum germinativum, thus initiating the morphological process. Much remains to be done before a full understanding of the events leading up to this is possible.

## V. Acknowledgments

I wish to express thanks both to Professor C. W. Emmens and Dr. J. D. Biggers for advice and criticism during the course of this work, which was assisted by grants from the Wool Industry Fund and the Commonwealth Bank of Australia.

## VI. References

Allen, E. (1922).—Amer. J. Anat. 30: 297.
Biggers, J. D. (1951).-J. Endocrinol. 7: 163.
Biggers, J. D. (1953a).-J. Endocrinol. 9: 136.
Biggers, J. D. (1953b).-J. Anat. (Lond.) 87 (in press).
Biggers, J. D., and Claringbold, P. J. (1953).-Aust. J. Biol. Sci. 6: 305.
Bullough, W. S., and Johnson, M. (1951).-Proc. Roy. Soc. B 138: 562.
Claringbold, P. J., Biggers, J. D., and Emmens, C. W. (1953).-Biometrics 4 (in press).
Emmens, C. W. (1950).-"Hormone Assay." Ch. XVI. (Academic Press: New York.)
Finney, D. J. (1952).—"Probit Analysis. A Statistical Treatment of the Sigmoid Response Curve." 2nd Ed. (Cambridge University Press.)
Heller, C. G. (1940).-Endocrinology 26: 619.


[^0]:    - Department of Veterinary Physiology, University of Sydney.

