Detection of Corynespora cassiicola in Hevea rubber tree from China

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Abstract. *Corynespora cassiicola*, which causes Corynespora leaf fall disease, is one of the most destructive and economically important fungal pathogens of Hevea rubber trees in Asian and African countries. This fungus is present in some Hevea rubber plantations and nurseries in Hainan and Yunnan provinces of South China. We describe the development of a PCR assay for *C. cassiicola*. With this test, of the 32 leaf samples obtained from 28 Hevea rubber trees in 15 rubber plantations and nurseries of Hainan and Yunnan provinces, 21 tested positive for the fungus. This assay is useful for the screening and certification of young *C. cassiicola*-free Hevea rubber plants (for distribution to commercial growers) due to its high sensitivity, reliability and efficacy.

Corynespora cassiicola, the causal agent of Corynespora leaf fall disease, is one of the most economically important fungi of Hevea rubber trees in Asian and African countries (Ismail and Jeyanayagi 2003). It was first isolated from Hevea rubber in Malaysia (Newsam 1960) and afterwards in India (Ramakirishnan and Pillai 1961). It can cause circular or irregular amphigenous spots on both young and old leaves and 'fishbone' or 'railway-track' appearances on the main vein or small veinlets of the leaves, which results in leaves falling all year round, yield reduction of mature rubber trees and even plant death on susceptible clones. Therefore, the International Rubber Research and Development Board (IRRDB) has repeatedly warned rubber producers of this disease and of the risk of outbreaks (IRRDB 2000). Corynespora cassiicola was isolated from Hevea rubber trees in rubber nurseries and plantations of Hainan and Yunnan provinces, China (Pu et al. 2007). Since the rubber tree is vegetatively propagated, the use of the C. cassiicola-free planting materials is the primary method of disease control. To ensure the production of C. cassiicolafree young rubber plants, it is necessary to have a sensitive disease detection method. In this communication, we describe a PCR detection method for C. cassiicola, which we have used to detect diseased leaves of Hevea rubber trees from rubber plantations and nurseries in Hainan and Yunnan provinces of China.

Thirty-two Hevea rubber leaf samples obtained from 28 Hevea rubber trees in 15 rubber plantations and nurseries of Yunnan and Hainnan Provinces, China (Table 1) were screened for *C. cassiicola* by PCR detection. Primers CCF and CCR (CCF: 5'-CCC TTC GAG ATA GCA CCC-3'; CCR: 5'-ATG CCC TAA GGA ATA CCA AA-3') were designed by the alignment software Omega 2.0, Primer Express and Primer 5.0, on the basis of the ITS sequence of the rRNA gene

of *C. cassiicola* [GenBank accessions EF490450, EF471932, EF198115, EF198116, EF198117, EF545008, DQ780453, DQ780421, AY238606, AY237605, AF163087 and U95173]. Primers were designed to amplify a fragment of 272 bp. The specificities of the primers were computer-tested, as was the theoretical PCR product. No significant alignments were found with sequences deposited in the public database of the US National Center for Biotechnology Information (NCBI).

DNA was extracted from Hevea rubber tree leaves according to the method described by Ausubel et al. (1998). Negative controls (no template and healthy rubber leaves) and a positive control (a recombinant plasmid CATAS001) were used. PCR reactions were performed in a total volume of $20\,\mu\text{L}$ with $10\times$ concentrated PCR reaction buffer containing 1.5 mM MgCl₂ and a final concentration of 200 µM of each dNTP, 1 µM of each primer and 2.5 units of Taq DNA polymerase per reaction. Amplification was performed in thin-walled PCR tubes in a PTC200 DNA thermal cycler (TM Research) with heated lid, which was programmed as follows: 1 cycle of 2 min at 94°C, followed by 30 cycles of 1 min at 94°C, 1 min at 62°C. One cycle of 5 min at 72°C was conducted after the 30 cycles. After amplification, 8 µL of the reaction mixture was loaded onto a 0.8% agarose gel in $1 \times TAE$ buffer, separated by electrophoresis, stained with ethidium bromide and viewed and photographed under UV light. A 2000-bp DNA ladder standard (TaKaRa Biotech) was used as the marker.

Of the 32 samples tested for *C. cassiicola* by PCR detection, 21 were found to be infected (Fig. 1). Negative controls (no DNA target and healthy rubber leaves) were included in every experiment to test for contamination, as well as a positive control (a recombinant plasmid CATAS001). The positive control, but not the negative controls, was to yield an amplicon of 272 bp

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Detection no.	Source	Result	Detection no.	Source	Result
1	Danzhou Hainan	+	17	Hekou Yunnan	_
2	Danzhou Hainan	+	18	Hekou Yunnan	+
3	Danzhou Hainan	+	19	Qionghai Hainan	+
4	Dingan Hainan	+	20	Qionghai Hainan	+
5	Dingan Hainan	+	21	Qionghai Hainan	+
6	Dingan Hainan	_	22	Qionghai Hainan	_
7	Dingan Hainan	_	23	Qiongzhong Hainan	+
8	Changjiang Hainan	+	24	Qiongzhong Hainan	_
9	Changjiang Hainan	+	25	Qiongzhong Hainan	_
10	Changjiang Hainan	_	26	Qiongzhong Hainan	+
11	Lingao Hainan	+	27	Ledong Hainan	+
12	Lingao Hainan	+	28	Ledong Hainan	+
13	Lingao Hainan	_	29	Ledong Hainan	_
14	Sanya Hainan	+	30	Qiongshan Hainan	_
15	Sanya Hainan	+	31	Qiongshan Hainan	_
16	Sanya Hainan	+	32	Qiongshan Hainan	+

 Table 1. PCR assay of Hevea rubber tree leaves for C. cassiicola

 +, infected rubber leaves; -, non-infected rubber leaves



Fig. 1. Detection of *C. cassiicola* in Hevea rubber tree leaves. M, molecular weight marker; P, positive control (a recombinant plasmid CATAS001); 1–32, Hevea rubber leaf samples (Table 1); N, negative controls (no template and healthy rubber leaves).

with the primer pair CCF–CCR. Twenty-one samples yielding amplicons were identical to the positive control in size and shared 99–100% nucleotide sequence homology with the positive control by cloning and sequence analysis.

Using this PCR technique, it is feasible to process a large number of Hevea rubber leaf samples at a single time. This will be useful for plant diagnostic clinics, for *C. cassiicola*free certification of tissue culture plants, quarantine verification, germplasm screening and selection of disease-resistant plants. The symptomatic expression of *C. cassiicola* is variable and cannot be relied on as a means of diagnosis and indeed most of samples tested were symptomless. Infected plants may, therefore, be inadvertently propagated if diagnosis is only by visual screening.

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