

First record of powdery mildew on carrots in Australia

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Abstract. Powdery mildew is reported on carrots for the first time in Australia. The affected plants were on a property located in the Murrumbidgee Irrigation Area of New South Wales. Based on morphological data and an rDNA ITS sequence, the fungus was identified as *Erysiphe heraclei*. Inoculation tests showed that the fungus was aggressive on carrot, but weakly pathogenic on parsnip and parsley. *Erysiphe heraclei* is common on parsnip but this is the first record on carrot in Australia.

Carrots (*Daucus carota*) are grown in the Murrumbidgee Irrigation Area of New South Wales either for juicing or the fresh market. The crops are grown all year and watered by furrow irrigation. Parsnips (*Pastinaca sativa*) growing on the same

properties are commonly infected by powdery mildew. Early in March 2007, powdery mildew was found affecting large areas of carrots on one farm in the region. When powdery mildew of carrots was examined on the farm that had the original outbreak,

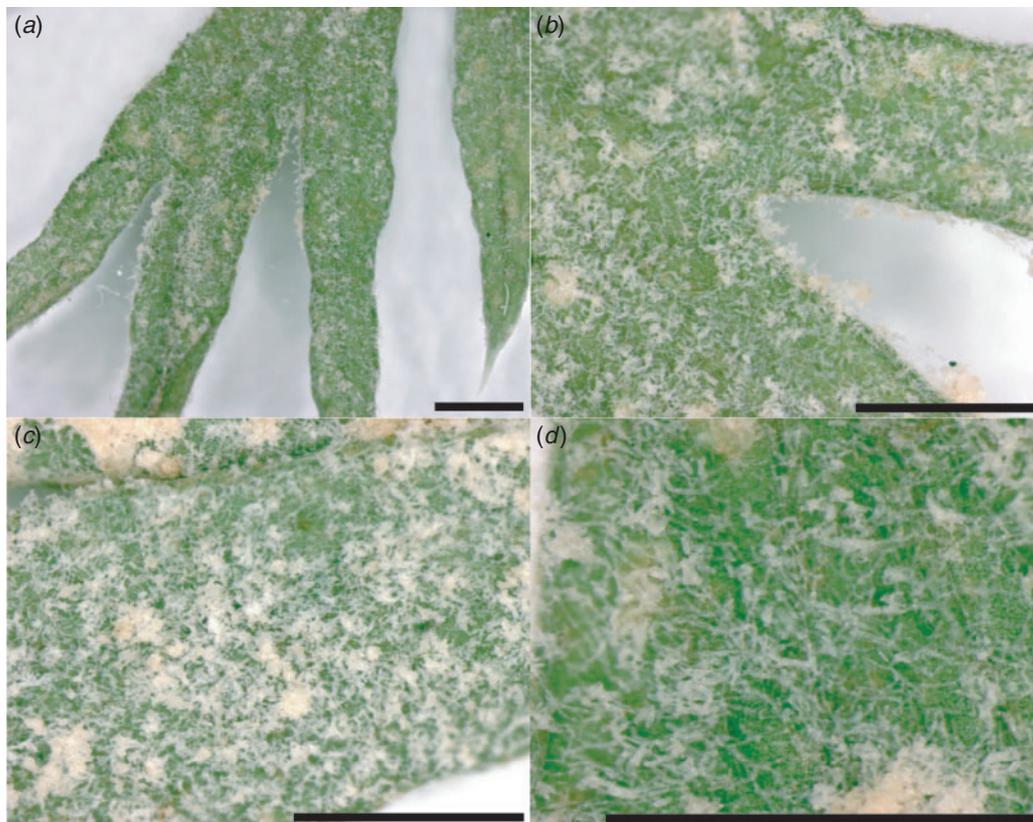


Fig. 1. Anamorphic *Erysiphe heraclei* on *Daucus carota* seen under stereomicroscope (VPRI 41227) (bars = 1 mm).

all blocks were infected and infection within the blocks was severe. The infected blocks ranged in maturity from 6 weeks after sowing to blocks that were ready for harvesting. The crops closer to maturity with the heavier canopies were more seriously infected. Spraying with sulfur was immediately carried out. The fungus showed conspicuous growth of white mycelium and conidia that covered the whole plant (Fig. 1). Morphology of the fungus was examined on microscopic mounts in lactic acid. Biometric data was obtained only from the examination of turgid

structures and only mature conidia (those not attached to conidiophores) were measured. The fungus was identified as *Erysiphe heraclei* and its description is given below.

Anamorphic *Erysiphe heraclei* DC., Fl. Fr. VI: 107 (1815) on *Daucus carota* (Fig. 2)

Mycelium amphigenous, mainly on the upper leaf surface. Hyphae 5–7.5 µm wide. Mycelial appressoria lobed.

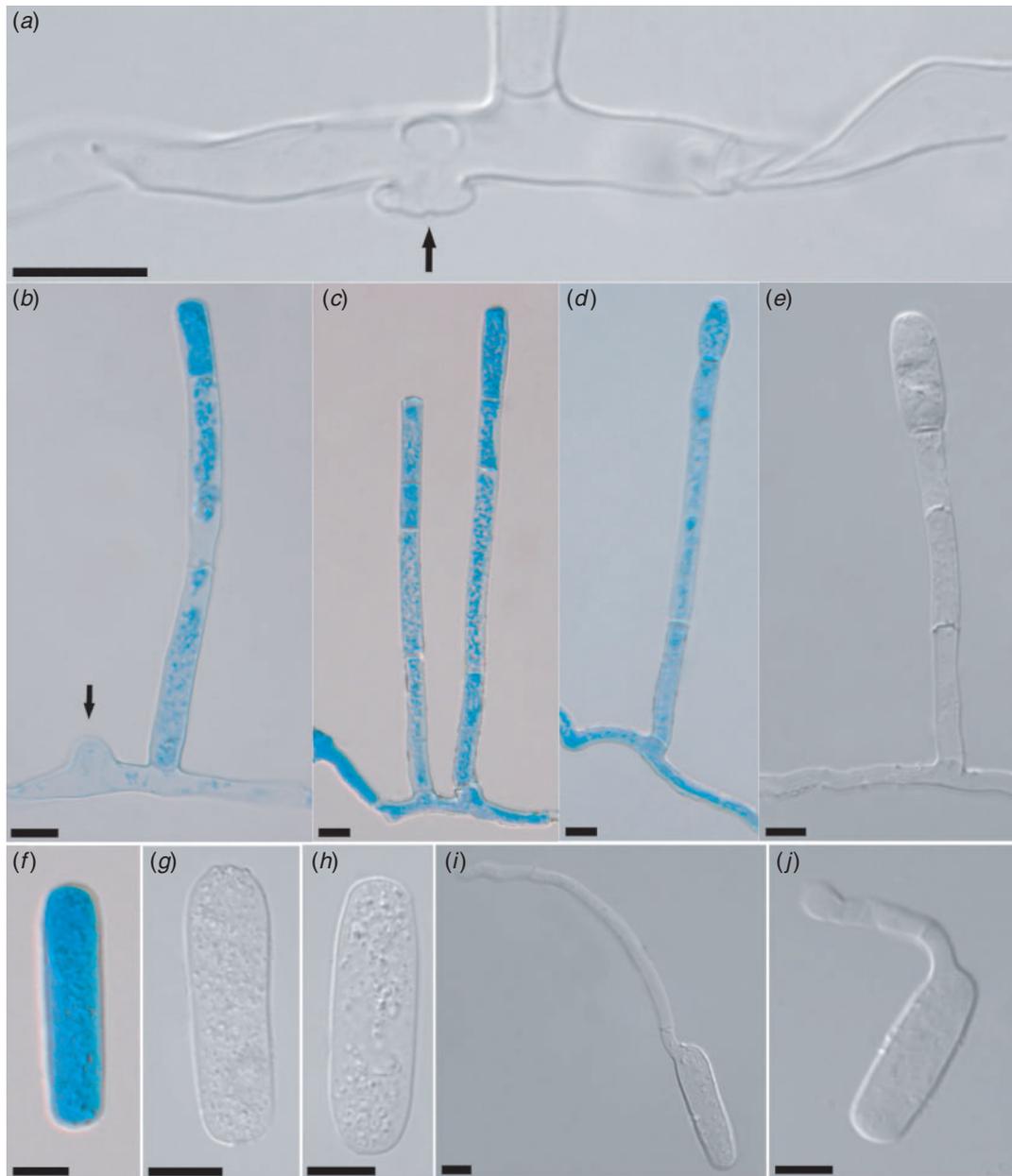


Fig. 2. Anamorphic *Erysiphe heraclei* on *Daucus carota* (bars = 10 µm). (a) Mycelial lobed appressorium (arrowed). (b) Stained conidiophore. A second conidiophore is arising from the same mycelial mother-cell (arrowed). (c, d) Stained conidiophore. (e) Conidiophore. (f) Stained conidium. (g, h) Conidia. (i, j) Germinated conidia. (a–j) VPRI 41227. (e) BRIP 49115.

Conidiophores up to 170 µm long, sometimes two conidiophores arising from the same hyphal cell, foot-cells 25–50 × 6–7.5 µm, followed by (1–)2 cells (which can be shorter or one longer and one shorter or one shorter and one longer than the foot-cell). Conidia formed singly, most subcylindric to cylindric, 32.5–62.5 × (10–)12.5–16.5(–19) µm. Germ tube at the shoulder of conidium, up to 1.5 times the conidium length. Teleomorph not seen.

Material examined: *Daucus carota*, Murrumbidgee Irrigation Area, NSW, Australia, March 2007, A. Watson (BRIP 49115), 6 Aug 2007 (VPRI 41227).

To help confirm the identification, an rDNA internal transcribed spacer (ITS) sequence was obtained for the specimen VPRI 41227 according to Cunnington *et al.* (2003), except that total DNA was extracted from the infected plant material using a DNeasy™ Plant Mini Kit (Qiagen). The sequence was identical, or differed by up to three bases, to six *E. heraclei* sequences on GenBank (AB000942, AB104511, AB104514, AB104513, AB104464 and EU010381). There were two *E. heraclei* sequences that differed by five to six bases (AB104510 and AB104512). The next most similar sequences were from the beet pathogen *Erysiphe betae*. These differed by five to seven bases. Intraspecific rDNA ITS sequence similarity in powdery mildew fungi is usually ≥99% (Cunnington *et al.* 2003), which in this case corresponds to ≤5 bases. The sequence obtained here confirms the identity as *E. heraclei* and has been lodged on GenBank as accession EU371725.

To prove the pathogenicity of the fungus, diseased infected carrot plants were collected in the field and brought

to a glasshouse maintained at 20–25°C where conidia were shaken over several potted healthy carrots of ~6 weeks old. As *E. heraclei* has been recorded from 80 genera in the Apiaceae (Braun 1987), young parsnips and parsley (*Coriandrum sativum*) were also inoculated. After 28 days there were only small, restricted colonies of powdery mildew on the parsnips and parsley plants, but the carrots were heavily infected (Fig. 3). This low level of infection on parsnip was surprising and may indicate that this is a new carrot specific strain of *E. heraclei*, rather than infection from parsnips grown in the area. Further controlled experiments using a wide range of species in the Apiaceae would be required to predict the host range of the fungus.

The only powdery mildew belonging to the subfamily *Erysiphoideae* (whose anamorphs belong to the genus *Oidium*) thus far reported on carrot is *E. heraclei*, which is distributed worldwide and can also infect numerous species of various host genera of the Apiaceae (Braun 1987). *Erysiphe heraclei* has been recorded on parsnips in Victoria (Cunnington 2003). Powdery mildew on parsnip has also been recorded in South Australia, Tasmania, New South Wales, Western Australia and Queensland, but these specimens were only identified as *Oidium* sp. (herb. DAR, VPRI and BRIP records). Many of these specimens have been examined by staff at VPRI and are all believed to be *E. heraclei* (I. G. Pascoe, pers. comm.).

The fungus may have been introduced on contaminated seed. But, as a range of carrot varieties from different sources were affected, we could not distinguish if a particular variety was the source. This is the first record of powdery mildew on carrots in Australia.



Fig. 3. Inoculation experiment where carrots infected with powdery mildew were shaken over healthy carrots, parsnip and parsley. After 4 weeks, the carrots were heavily infected, but there was only minor infection on the parsnip and parsley.

Acknowledgements

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References

Braun U (1987) A monograph of the *Erysiphales* (powdery mildews). *Beiheft zur Nova Hedwigia* **89**, 1–700.

Cunnington JH (2003) 'Pathogenic fungi on introduced plants in Victoria. A host list and literature guide for their identification.' (Department of Primary Industries, Victoria: Melbourne)

Cunnington JH, Takamatsu S, Lawrie AC, Pascoe IG (2003) Molecular identification of anamorphic powdery mildew fungi. *Australasian Plant Pathology* **32**, 421–428. doi: 10.1071/AP03045

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