

Supplementary material

**Distribution of arsenic species in an open seagrass ecosystem: relationship to trophic groups, habitats and feeding zones**

A. Price,<sup>A,B</sup> W. Maher,<sup>A,E</sup> J. Kirby,<sup>A,C</sup> F. Krikowa,<sup>A</sup> E. Duncan,<sup>A</sup> A. Taylor<sup>A</sup> and J. Potts<sup>A,D</sup>

<sup>A</sup>Ecochemistry Laboratory, Institute for Applied Ecology, Faculty of Applied Science, University of Canberra, ACT 2601, Australia.

<sup>B</sup>Murray–Darling Freshwater Centre, PO Box 991, Wodonga, VIC 3685, Australia.

<sup>C</sup>CSIRO Land and Water, PMB2 Glen Osmond, SA 5064, Australia.

<sup>D</sup>Coastal Water Science Unit, NSW Office of Environment and Heritage, Department of Premier and Cabinet, 59–61 Goulburn Street, Sydney, NSW 2000, Australia.

<sup>E</sup>Corresponding author. Email: bill.maher@canberra.edu.au

**Table S1. Arsenic concentrations measured in certified reference materials**

Certified reference material	<i>n</i>	Measured ( $\mu\text{g g}^{-1}$ )	Certified ( $\mu\text{g g}^{-1}$ )
DORM-2 Dogfish muscle	8	17.9 ± 0.6	18 ± 1.1
DOLT-1 Dogfish liver	8	9.9 ± 0.5	10.1 ± 1.4
NIST-1566a Oyster tissue	7	13.8 ± 0.5	14 ± 1.2
TORT-2 Lobster hepatopancreas	8	26 ± 2	24.6 ± 2.2
NIES-9 Sargasso	4	109 ± 5	115 ± 9
CRM-279 Sea lettuce ( <i>Ulva lactuca</i> )	5	3.6 ± 0.3	3.09 ± 0.2
CRM 402 White Clover	3	0.085 ± 0.008	0.093 ± 0.01

**Table S2. Principal components analysis of arsenic species' proportions in seagrass sediment, plant and animal tissues**

Factor loadings in bold have more influence on the sample location in three-dimensional space

Axis	Eigenvalues	Percentage variation	Cumulative percentage variation
PC1	$2.1 \times 10^3$	79.9	79.9
PC2	376	14.1	94
PC3	87.1	3.3	97.3
Variable	PC1	PC2	PC3
Arsenoriboside 1	0.172	0.279	0.234
Arsenoriboside 2	0.108	<b>0.407</b>	-0.112
Arsenoriboside 3	0.059	0.193	<b>-0.792</b>
Arsenoriboside 4	0.011	-0.028	-0.018
DMA	0.073	0.104	<b>0.466</b>
MA	0.060	0.094	0.288
Inorganic arsenic	<b>0.419</b>	<b>-0.798</b>	-0.072
AB	<b>-0.878</b>	-0.248	0.003
TMAO	-0.014	-0.003	0.001
AC	-0.005	0.000	0.003
TETRA	-0.004	0.001	-0.001

**Table S3. Method performance data for speciation analyses: (a) extraction and column recoveries for a pooled sample (mean  $\pm$  standard deviation,  $n = 4$ ) and (b) peak times and precision**

Limits of detection are based on 20- $\mu$ L injection,  $n = 10$ ,  $<0.01 \mu\text{g g}^{-1}$ , for all arsenic species except for inorganic arsenic, which is  $<0.001 \mu\text{g g}^{-1}$ .

Samples were spiked with standards to confirm peak identity and matrix effects. Recoveries of spikes are in the range 99–101 %,  $100 \pm 1$  %

( $n = 10$ )

(a)							
Sample	Methanol–water extraction				Column recoveries		
<i>Zostera capricornii</i>	117 $\pm$ 6				111 $\pm$ 5		
<i>Cystophora moniformis</i>	80 $\pm$ 4				99 $\pm$ 2		
<i>Saccrostrea glomerata</i>	78 $\pm$ 2				89 $\pm$ 4		
<i>Acanthopagrus australis</i>	95 $\pm$ 2				100 $\pm$ 1		
(b)							
Cations							
Arsenic species	AB	AS1	TMAO	AC	TETRA		
Peak time ( $n = 5$ )	3.6 $\pm$ 0.1	4.10 $\pm$ 0.08	5.2 $\pm$ 0.1	6.50 $\pm$ 0.07	8.1 $\pm$ 0.1		
Precision ( $n = 5$ )							
Concentration ( $\mu\text{g L}^{-1}$ )	10	1	0.2	0.05	0.2		
CV (%)	<1	<15	<8	<4	<4		
Anions							
Arsenic species	As <sup>3+</sup>	DMA	MA	AS2	As <sup>5+</sup>	AS3	AS4
Peak time ( $n = 5$ )	2.5 $\pm$ 0.1	3.10 $\pm$ 0.08	5.1 $\pm$ 0.1	7 0.0 $\pm$ 0.1	10.0 $\pm$ 0.5	13.5 $\pm$ 0.8	31 $\pm$ 1
Precision ( $n = 5$ )							
Concentration ( $\mu\text{g L}^{-1}$ )	10	0.2	0.2	1	0.5	1	1
CV (%)	<1	<10	<5	<15	<6	<15	<15