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#### **Supplementary Material**

# Inhibition effect of kaolinite on antibiotic resistance genes development in *Escherichia coli* induced by sublethal ampicillin and its molecular mechanism

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#### Protein extraction and digestion

Based on data of the antibiotic tolerance assay, the strains induced on the last day were selected for proteomic analysis. The strain was grown in LB media containing antibiotics at 37  $^{\circ}$  C, and the antibiotic concentration was set as the 1/2 MIC value of the strains, and the strains were incubated in the LB media containing kaolinite. The strains were also incubated in kaolinite-free media as a control.

At the early stationary phase, the cells were harvested and washed twice with ice-cooled phosphate-buffered saline (PBS) solution and pelleted. The collected pellets were immediately stored at -80 °C and used for protein extraction. The mortar was pre-cooled, the sample was thoroughly ground and 1 ml of lysis buffer (1 mL/0.1 g; 7 M urea, 2 M thiourea, 4% CHAPS, and 1% Protease Inhibitor Cocktail) was added, dissolved and transferred to a 1.5 mL EP tube, sonicated (700w, 1.5s interval, 1s, ultrasonic 10min), 12000r / min, Centrifuge for 20 minutes and collect the supernatant. The supernatant was dispensed into 4 tubes of about 250 µL per tube and precipitated by adding 1 mL of acetone at -20 ° C overnight. The precipitated protein was centrifuged at 12000 r / min for 20 minutes at 4 ° C, the supernatant was decanted, and dried to obtain a processed protein mass. Add appropriate amount of lysis buffer (without SDS) to dissolve the protein. Ultrasonic dissolution (100 W, 0.8 sec, 0.8 sec, ultrasound, repeated once), after sonication at 4 ° C, 12000 r / min, centrifugation for 20 minutes, aspirate the supernatant and transfer to a new EP tube.

#### Characterization of E.coli-mineral complexes and minerals.

The X-ray diffraction patterns (XRD) of E.coli -mineral complexes or minerals were recorded by a powder diffractometer (Bruker, German) under Cu ka radiation( $\lambda = 0.154$  nm, Ni-filter) with a scan range of 3°-80°, a step size of 0.02° and a measuring time of 19.2 s per step.

The Fourier transform infrared (FTIR) spectra was recorded using a FTIR spectrometer Vertex-33 (Bruker, German) with the KBr pellet technique. Solid-state NMR spectra were collected on a Bruker Avance III HD 600 spectrometer. Al spectra were obtained at 104.26 MHz (14.095 T) with a 1s pulse delay. Si spectra were obtained at 79.49 MHz (14.095 T) with a 5s pulse delay. The spinning speed was 8 kHz.

The X-ray photoelectron spectra (XPS) were measured with an X-ray photoelectron spectrometer (Kratos, Axis Ulra DLD) and C1s (284.6 eV) was chosen as the reference. The pass energy was 40 eV and conventional Al K (1486.6 eV) anode radiation source was used as the excitation source. The C  $_{1s}$  and N  $_{1s}$  high resolution spectra and survey spectra of the samples were recorded. The C  $_{1s}$  and N  $_{1s}$  peak decomposition was performed using mixed Gaussian–Lorentzian (80/20) line shape with XPSPEAK software.

For SEM observation, after 48 h of incubation, the E.coli /mineral complexes or minerals were harvested and fixed with 2.5 % (v/v) glutaraldehyde in PBS at  $4^{\circ}$ C overnight, then E.coli /mineral complexes or minerals alone (without the addition of bacteria) were washed with 0.1 M PBS followed by 5 steps of dehydration with 30, 50,

70, 90 and 100 % ethanol solution for 10 min at each dehydration step. Thereafter, the pellets were resuspended in tertbutyl alcohol (TBA) for 20 min and then freeze-dried for at least 24 h. Dry specimens were gold coated for 90 s under vacuum in a JFC-1600 sputter coater (JEOL, Japan) to improve conductivity. The SEM images were obtained by a Zeiss EV0 LS10 scanning electron microscope (Carl Zeiss, Germany).



Fig. S1. High-resolution C 1s spectra of ampicillin + *E.coli* + kaolinite (A) and *E.coli* + kaolinite (C), and high-resolution N 1s spectra of ampicillin + *E.coli* + kaolinite (B) and *E.coli* + kaolinite (D).



Fig. S2. Venn diagram showing the overlaps of the differentially expressed proteins in two experimental groups

Targeted genes	Primers Sequence (5'to 3')	Annealing temperature
		(°C)
acrA	F:CAGATCACAACCGAGCTTCC	60
	R:GCTTCGATGTCGCTACCTTC	
marA	F:AATACATCCGCAGCCGTAAG	60
	R:TGAAGGTTCGGGTCAGAGTT	
marR	F:CAACCTTTCCACCCAGCCTT	60
	R:TGCTCTGCTCTATCCGCTGC	
tetB	F:ACGTGATAATACAGATACCGAA	60
	R:CACAAAGGCTTGGAATACTGA	
blaTEM	F:CCCCGAAGAACGTTTTCCAA	60
	R:TAGTGTATGCGGCGACCGAG	
blaFOX	F:AACATGGGGTATCAGGGAGATG	60
	R:CAAAGCGCGTAACCGGATTGG	
tetE	F:TTTGGTGGCGGTATGATTGC	60
	R:TGCCAGCCCGTTAATAGCAG	
tetZ	F:CCTTCTCGACCAGGTCGG	60
	R:ACCCACAGCGTGTCCGTC	
tetA	F: GCTACATCCTGCTTGCCTTC	60
	R:CATAGATCGCCGTGAAGAGG	

# Table S1. Targeted antibiotic resistance genes and their primer sequence

16S rRNA gene

## F:ATGGYTGTCGTCAGCTCGTG 60

#### R:GGGTTGCGCTCGTTGC

Table S2. Fitting of kinetic models for the adsorption of tetracycline (TET), ampicillin (AMP) and mixed antibiotics by kaolinite in aqueous solutions at pH of 7.0, 37°C (initial antibiotic concentration, 16 mg  $L^{-1}$ ; kaolinite dosage, 10 g  $L^{-1}$ ).

	Pseudo-first-order model			~	Pseudo-second-order model		
Adsorbates	$k_1$	<i>q</i> e	$R^2$	$q_{\rm e, exp}$ (mg g <sup>-1</sup> )	<i>k</i> <sub>2</sub>	<i>q</i> e	$R^2$
	$(\min^{-1})$	$(mg g^{-1})$			$(g mg^{-1} min^{-1})$	$(mg g^{-1})$	
TET	0.1492	0.9033	0.9328	0.9248	0.2898	0.9464	0.9998
AMP	0.3638	0.4602	0.9103	0.4672	1.0426	0.4721	0.9986
TET in mixed	0 2262	0 6975	0.0119	0 6012	1 0057	0 6091	0.0001
solution	0.3362	0.08/5	0.9118	0.0912	1.0937	0.0981	0.9991
AMP in mixed	0 1939		0.0265	0 2472	1 1450	0 2522	0.0006
solution	0.1656		0.9203	0.3472	1.1430	0.5552	0.9996

	Citable	Gene		
Group	Accession	Name	AVG.	FLAG
AMP vs. control	P00370	gdhA	2.1289515	UP
AMP vs. control	P00448	sodA	0.25613	DOWN
AMP+KA vs. control	P00448	sodA	0.2697959	DOWN
AMP+KA vs. control	P00562	metL	1.995601	UP
AMP+KA vs. control	P00968	carB	0.663919	DOWN
AMP vs. control	P02918	mrcA	0.6340377	DOWN
AMP+KA vs. control	P07862	ddlB	1.599626	UP
AMP+KA vs. control	P08337	mutT	1.8709995	UP
AMP vs. control	P09155	rnd	0.6576648	DOWN
AMP+KA vs. control	POA6L9	hscB	0.4659021	DOWN
AMP vs. control	P0A7A2	gpmB	1.513818	UP
AMP+KA vs. control	P0A7R5	rpsJ	0.5547198	DOWN
AMP+KA vs. control	P0A7W7	rpsH	0.6226237	DOWN
AMP+KA vs. control	P0A8B5	ybaB	0.5728507	DOWN
AMP+KA vs. control	P0A8K5	yaeP	0.5917874	DOWN
AMP vs. control	P0A908	mipA	0.3224485	DOWN
AMP vs. control	P0A968	cspD	1.729835	UP
AMP vs. control	P0A9H7	cfa	0.6670613	DOWN
AMP vs. control	P0A9M2	hpt	0.6166016	DOWN

# Table S3. A selection of proteins

AMP+KA vs. control	P0AA37	rluA	0.5889997	DOWN
AMP vs. control	P0AAC8	iscA	0.3500046	DOWN
AMP+KA vs. control	P0AAC8	iscA	0.6344143	DOWN
AMP+KA vs. control	P0AAD6	sdaC	11.912925	UP
AMP+KA vs. control	P0AAN9	iraP	0.6198615	DOWN
AMP vs. control	P0AB24	efeO	0.3383362	DOWN
AMP+KA vs. control	P0AB24	efeO	0.2977567	DOWN
AMP+KA vs. control	P0AB55	yciI	0.5579008	DOWN
AMP+KA vs. control	P0ABQ4	folA	0.6638064	DOWN
AMP vs. control	P0ABU9	tolQ	0.4609569	DOWN
AMP vs. control	P0AC33	fumA	2.3014665	UP
AMP vs. control	P0ACB2	hemB	0.5406323	DOWN
AMP vs. control	P0ACD4	iscU	0.1527631	DOWN
AMP vs. control	P0ADA1	tesA	0.6025851	DOWN
AMP vs. control	P0ADS9	yggN	0.5495642	DOWN
AMP+KA vs. control	P0AEJ4	envZ	2.5256486	UP
AMP vs. control	P0AEL0	fdoI	1.614427	UP
AMP vs. control	P0AFH2	oppB	1.535024	UP
AMP vs. control	P0AFL3	ppiA	0.6252788	DOWN
AMP vs. control	P0AFM2	proX	0.4529168	DOWN
AMP+KA vs. control	P0AG90	secD	3.7172824	UP
AMP+KA vs. control	P0DMC5	rcsC	1.682692	UP

AMP+KA vs. control	P12282	moeB	1.698406	UP
AMP vs. control	P14175	proV	0.2040199	DOWN
AMP+KA vs. control	P14175	proV	0.5705512	DOWN
AMP vs. control	P15877	gcd	0.1455577	DOWN
AMP+KA vs. control	P15877	gcd	0.4864468	DOWN
AMP vs. control	P16703	cysM	0.497119	DOWN
AMP+KA vs. control	P17117	nfsA	2.4119754	UP
AMP vs. control	P21829	ybhA	2.0896505	UP
AMP vs. control	P24555	<i>ptrB</i>	0.6672593	DOWN
AMP vs. control	P28904	treC	3.3114514	UP
AMP vs. control	P31660	prpC	0.645661	DOWN
AMP vs. control	P33030	yeiR	0.6311982	DOWN
AMP vs. control	P33221	purT	3.0211049	UP
AMP+KA vs. control	P36672	treB	9.5069551	UP
AMP+KA vs. control	P37908	yfjD	1.5780295	UP
AMP+KA vs. control	P39180	flu	0.2929394	DOWN
AMP vs. control	P39451	adhP	0.3722012	DOWN
AMP+KA vs. control	P39451	adhP	0.4943579	DOWN
AMP+KA vs. control	P39830	ybaL	1.7952095	UP
AMP+KA vs. control	P45395	kdsD	0.6397416	DOWN
AMP vs. control	P63020	nfuA	0.6340377	DOWN
AMP+KA vs. control	P63177	rlmB	0.6368226	DOWN

AMP vs. control	P64534	rcnB	0.6698917	DOWN
AMP vs. control	P69874	potA	0.5155783	DOWN
AMP+KA vs. control	P76116	yncE	0.3435725	DOWN
AMP+KA vs. control	P76149	sad	2.4334674	UP
AMP+KA vs. control	P76217	astD	0.4024045	DOWN
AMP+KA vs. control	P76621	csiD	0.3362383	DOWN
AMP vs. control	P77252	ykgE	0.4051915	DOWN
AMP+KA vs. control	P77581	astC	0.2714537	DOWN

Group	GO Classification	GO ID	GO Name	Gene ID	p-value
<u>^</u>				P00448,	-
				P02918,	
				P0AC33,	
				POACB2,	
				P15877,	
				P39451,	
				P69874,	
				P0A9M2,	
				P0ACD4,	
				P33030,	
				P63020,	
				POAAC8,	
				POAELO,	
				P14175,	
				P16703,	
				P33221,	
AMP vs.	molecular_functio	GO:004316		P21829,	
control	n	7	ion binding	P64534	0.002704
				P00370,	
				P00448,	
				P15877,	
AMP vs.	molecular_functio	GO:001649	oxidoreductase	P39451,	
control	n	1	activity	P0AEL0	0.014353
				P02918,	
AMP vs.	molecular_functio	GO:000823		POADA1,	
control	n	3	peptidase activity	P24555	0.010377
	1 1 6 2			P0AC33,	
AMP vs.	molecular_functio	GO:001682	1	POACB2,	0.051125
control	n	9	Iyase activity	P16/03	0.071127
	1 1 6 2	00.001.005		P0AC33,	
AMP vs.	molecular_functio	GU:001685	·····	PUAFL3,	0.016657
control	n	5	isomerase activity	PUA/A2	0.016657
	mologilar front	CO.000005	transmembrane	$\frac{1}{100} \frac{1}{100} \frac{1}$	
AMP VS.	molecular_functio	GU:002285	transporter	PUAFIMZ,	0.056570
	II molecular functio	/ CO:000267	activity	P141/3	0.030372
AIVIP VS.	noiecular_functio	GU:00036/ 7	DNA hinding	PUU448,	0.001616
control	11	1	transforms	LOUA209	0.081010
			activity		
	molecular functio	GO:001674	transferring acul	P31660	
control	n	6	groups	ΡΩΔΠΛ1	0.040761
AMP vs. control	molecular_functio n	GO:001674 6	transferase activity, transferring acyl groups	P31660, P0ADA1	0.040761

# Table S4. GO analysis

			transferase		
			activity,		
AMP vs.	molecular_functio	GO:001675	transferring	P02918,	
control	n	7	glycosyl groups	P0A9M2	0.027129
AMP vs.	molecular_functio	GO:001688		P69874,	
control	n	7	ATPase activity	P14175	0.016576
				P00448,	
				P0AB24,	
				P39451,	
				P0AFM2,	
				P63020,	
				POAELO,	
				P14175,	
				P28904,	
AMP vs.		GO:000695		P0A968,	
control	biological_process	0	response to stress	P64534	0.002064
				P00370,	
				P02918,	
				P0ACB2,	
				P0A9H7,	
				P0A9M2,	
				P0A7A2,	
				P16703,	
				P33221,	
AMP vs.		GO:000905	biosynthetic	P0A908,	
control	biological_process	8	process	P0A968	0.025641
				P0AC33,	
				P31660,	
				P0A9H7,	
				P0A9M2,	
				P0A7A2,	
AMP vs.		GO:004428	small molecule	POAELO,	
control	biological_process	1	metabolic process	P33221	0.017633
				P69874,	
				P0ABU9,	
				P0AFM2,	
AMP vs.		GO:000681		P0AFH2,	
control	biological_process	0	transport	P14175	0.012210
				POACB2,	
				P09155,	
			cellular nitrogen	P0A9M2,	
AMP vs.		GO:003464	compound	P33221,	0.072716
control	biological_process		metabolic process	PUA968	0.073712
AMP vs.	biological_process	GO:005118	cofactor	POACB2,	0.008928

control		6	metabolic process	P0ACD4,	
				P63020,	
				P0A7A2,	
				P0AAC8	
			generation of	P0AC33,	
			precursor	P31660,	
AMP vs.		GO:000609	metabolites and	P0A7A2,	
control	biological_process	1	energy	P0AEL0	0.004253
				P0ACD4,	
				P63020,	
AMP vs.		GO:000679	sulfur compound	P0AAC8,	
control	biological_process	0	metabolic process	P16703	0.002282
				P31660,	
AMP vs.		GO:000662	lipid metabolic	P0A9H7,	
control	biological_process	9	process	P0ADA1	0.07657
			cellular	P0ACD4,	
AMP vs.		GO:002260	component	P63020,	
control	biological_process	7	assembly	P0AAC8	0.00228
				P00370,	
				P00448,	
				P0AC33,	
				P0ACB2,	
				P0A9H7,	
				P0A9M2,	
				P0ACD4,	
				P63020,	
				P0AAC8,	
				P28904,	
				P33221,	
				P77252,	
				P0A968,	
AMP vs.	cellular_componen	GO:000582		P21829,	
control	t	9	cytosol	P64534	0.002856
				P00370,	
				P00448,	
				P31660,	
				P09155,	
				P0A9H7,	
				P0A9M2,	
				P16703,	
				P28904,	
AMP vs.	cellular_componen	GO:000573		P0A968,	
control	t	7	cytoplasm	P24555	0.001548
AMP vs.	cellular_componen	GO:000588	plasma	P02918,	0.064852

control	t	6	membrane	P15877, P69874, P0ABU9, P0AFM2, P0AEL0, P0AFH2, P14175 P69874,	
				P0AFM2,	
AMP vs.	cellular_componen	GO:003299	protein-containin	POAELO,	0.01000
control	t cellular componen	I GO:000562	g complex	P14175	0.018925
control	t	2	intracellular	P09155	0.036610
control	t	2	external	10/155	0.030010
AMP vs	cellular componen	GO:003031	encansulating		
control	t	2	structure	P0A908 P00562, P00968, P07862, P15877, P39451, P76621, P77581,	0.026043
				P00448, P17117, P37908,	
AMP+K A vs. control	molecular_functio n	GO:004316 7	ion binding	P45395, P0AAC8, P0ABQ4, P0AEJ4, P0DMC5, P14175, P08337, P12282 P00562,	0.004116
AMP+K		00.001/10		P15877, P39451, P00448, P17117, P76149,	
A vs.	molecular_functio	GO:001649 1	oxidoreductase	PUABQ4, P76217	0 008817
AMP+K	molecular functio	GO:002285	transmembrane	P0AG90.	0.00001/
A vs.	n	7	transporter	P36672,	0.021228

control			activity	P14175, P39830, P0AAD6	
AMP+K				P0A/W/, P0A7R5,	
A vs.	molecular_functio	GO:000372		P63177,	
control	n	3	RNA binding	P0AA37	0.003005
				P00562,	
AMP+K				P36672,	
A vs.	molecular_functio	GO:001630		POAEJ4,	
control	n	1	kinase activity	P0DMC5	0.014757
AMP+K				P00448,	
A vs.	molecular_functio	GO:000367		P76116,	
control	n	7	DNA binding	P0A8B5	0.065263
AMP+K				POAEJ4,	
A vs.	molecular_functio	GO:000487	signal transducer	P0DMC5,	0.001007
control	n	1	activity	P141/5	0.001927
AMP+K		CO 001 (07		P00968,	
A VS.	molecular_functio	GO:001687	1:	PU/862,	0.000716
	Π	4	ingase activity	P12282	0.009/10
AMP+K	molecular functio	CO(000272)	structural	$\mathbf{D} \cap \mathbf{A} \mathbf{T} \mathbf{W} \mathbf{T}$	
A vs.	n	5	ribosome	POA7W7,	0.011476
$\Delta MP + K$	11	5	noosome	I UA/KJ	0.011470
A vs	molecular functio	GO·001685		P45395	
control	n	3	isomerase activity	P0AA37	0.081795
• on a of		c		P00562.	01001770
				P00968.	
				P07862,	
				P77581,	
				P0A7R5,	
				P45395,	
				P0ABQ4,	
				P0DMC5,	
AMP+K				P08337,	
A vs.		GO:000905	biosynthetic	POAAN9,	
control	biological_process	8	process	P12282	0.017867
				P39451,	
				P76621,	
				P00448,	
				P0AB24,	
AMP+K				P0AEJ4,	
A vs.		GO:000695		P0DMC5,	
control	biological_process	0	response to stress	P14175,	0.008636

				P08337,	
				POAAN9	
				P00968,	
				P0A7R5,	
				P63177,	
				P76149,	
				P0ABQ4,	
AMP+K			cellular nitrogen	P0DMC5,	
A vs.		GO:003464	compound	P0AA37,	
control	biological_process	1	metabolic process	POAAN9	0.029258
				P00562,	
				P00968,	
				P77581,	
AMP+K			cellular amino	P76149,	
A vs.		GO:000652	acid metabolic	P0ABQ4,	
control	biological_process	0	process	P76217	0.004231
				P0AG90,	
				P36672,	
AMP+K				P14175,	
A vs.		GO:000681		P39830,	
control	biological_process	0	transport	P0AAD6	0.013514
				P39451,	
				P77581,	
AMP+K				P76149,	
A vs.		GO:000905		P76217,	
control	biological_process	6	catabolic process	POAAN9	0.012319
				P0AG90,	
AMP+K				P36672,	
A vs.		GO:005508	transmembrane	P39830,	
control	biological_process	5	transport	P0AAD6	0.083458
AMP+K				P0AEJ4,	
A vs.		GO:000716	signal	P0DMC5,	
control	biological_process	5	transduction	P14175	0.004307
AMP+K				P0A7W7,	
A vs.		GO:004225	ribosome	P63177,	
control	biological_process	4	biogenesis	P0AA37	0.007586
AMP+K				P00968,	
A vs.		GO:004428	small molecule	P45395,	
control	biological_process	1	metabolic process	P0ABQ4	0.084281
				P00562,	
				P00968,	
AMP+K				P07862,	
A vs.	cellular_componen	GO:000582		P0A7W7,	
control	t	9	cytosol	P00448,	0.010827

				P0A7R5,	
				P17117.	
				P63177.	
				P0A6L9.	
				P0A8B5.	
				POAAC8.	
				POABO4.	
				POAB55.	
				P12282	
				P0AG90,	
				P15877.	
				P36672,	
				P37908,	
				POAEJ4,	
				P0DMC5,	
AMP+K				P14175,	
A vs.	cellular_componen	GO:000588	plasma	P39830,	
control	t	6	membrane	P0AAD6	0.053972
				P00968,	
				P07862,	
				P00448,	
AMP+K				P63177,	
A vs.	cellular_componen	GO:000573		P0A8B5,	
control	t	7	cytoplasm	P0AAN9	0.012118
				P0AG90,	
AMP+K				P0A7R5,	
A vs.	cellular_componen	GO:000562		POAEJ4,	
control	t	2	intracellular	P0DMC5	0.009071
AMP+K				P00968,	
A vs.	cellular_componen	GO:003299	protein-containin	P0A6L9,	
control	t	1	g complex	P14175	0.039097
AMP+K					
A vs.	cellular_componen	GO:000584		P0A7W7,	
control	t	0	ribosome	P0A7R5	0.014418
AMP+K					
A vs.	cellular_componen	GO:000557	extracellular		
control	t	6	region	P39180	0.006644
AMP+K			external		
A vs.	cellular_componen	GO:003031	encapsulating		
control	t	2	structure	P39180	0.027103

	KEGG			
Group	Pathway	Pathway Name	Gene ID	p-value
			P69874	
			P0AFM2	
AMP vs.			P14175	
control	eco02010	ABC transporters	P0AFH2	0.011696
AMP vs.				
control	eco00550	Peptidoglycan biosynthesis	P02918	0.014547
AMP vs.		Alanine, aspartate and glutamate		
control	eco00250	metabolism	P00370	0.02725
			P0AC33	
			P15877	
			P39451	
			P33221	
AMP vs.			P16703	
control	eco01130	Biosynthesis of antibiotics	P0A7A2	0.001264
			P0AC33	
			P0AEL0	
AMP vs.			P16703	
control	eco01200	Carbon metabolism	P0A7A2	0.0016
AMP vs.			P02918	
control	eco01501	beta-Lactam resistance	P0AFH2	0.000308
AMP vs.				
control	eco00030	Pentose phosphate pathway	P15877	0.025677
AMP vs.				
control	eco00640	Propanoate metabolism	P31660	0.039301
			P0ACB2	
			P0AC33	
			P15877	
			P39451	
			P0A9M2	
			P33221	
AMP vs.			P16703	
control	eco01110	Biosynthesis of secondary metabolites	P0A7A2	0.000524
AMP vs.				
control	eco00020	Citrate cycle (TCA cycle)	P0AC33	0.019777
AMP vs.				
control	eco00270	Cysteine and methionine metabolism	P16703	0.02725
AMP vs.			P16703	
control	eco01230	Biosynthesis of amino acids	P0A7A2	0.067943
AMP vs.				
control	eco00626	Naphthalene degradation	P39451	0.00038

### Table S5 KEGG Pathway

AMP vs.				
control	eco00350	Tyrosine metabolism	P39451	0.002223
AMP vs.			P39451	
control	eco00010	Glycolysis / Gluconeogenesis	P0A7A2	0.005164
AMP vs.				
control	eco00071	Fatty acid degradation	P39451	0.006287
AMP vs.				
control	eco00500	Starch and sucrose metabolism	P28904	0.030512
AMP vs.				
control	eco00910	Nitrogen metabolism	P00370	0.015789
AMP vs.		C	P0A9M2	
control	eco00230	Purine metabolism	P33221	0.036567
AMP vs.				
control	eco01040	Biosynthesis of unsaturated fatty acids	P0ADA1	0.00094
AMP vs.		5		
control	eco00220	Arginine biosynthesis	P00370	0.009023
		8	P0ACB2	
			POAC33	
			D15877	
			D02019	
			F02916	
			P39451	
			P00370	
			P0A9M2	
			P33221	
			P0AEL0	
AMP vs.			P16703	
control	eco01100	Metabolic pathways	P0A7A2	0.00838
AMP vs.		Glyoxylate and dicarboxylate		
control	eco00630	metabolism	P0AEL0	0.044983
AMP vs.				
control	eco00860	Porphyrin and chlorophyll metabolism	P0ACB2	0.014547
AMP vs.		Chloroalkane and chloroalkene		
control	eco00625	degradation	P39451	0.00038
AMP vs.		Cationic antimicrobial peptide (CAMP)		
control	eco01503	resistance	P0AFL3	0.033922
AMP vs.			P0AEL0	
control	eco00680	Methane metabolism	P0A7A2	0.001253
AMP vs.				
control	eco02024	Quorum sensing	P0AFH2	0.099429
AMP vs	2200 <u>202</u> 1	Glycine serine and threonine		0.0 <i>77</i> 1 <b>27</b>
control	eco00260	metabolism	ΡΩΔ7Δ2	0 03568
	CC000200	metabolism	1 UA / A2	0.05500
AIVIE VS.	01000	Dependencies of exempting operations	D20451	0 000001
	00670	Degradation of aromatic compounds	r 37431	0.005477
AMP vs.	eco006/0	One carbon pool by folate	P33221	0.005477

control				
AMP vs.				
control	eco00620	Pyruvate metabolism	P0AC33	0.068139
			P0ACB2	
			P0AC33	
			P39451	
			P0ABU9	
			P0AEL0	
AMP vs.		Microbial metabolism in diverse	P16703	
control	eco01120	environments	P0A7A2	0.000778
			P00968	
			P15877	
			P39451	
			P77581	
			P00562	
			P07862	
			P76149	
AMP+KA			P45395	
vs.			P76217	
control	eco01100	Metabolic pathways	P0ABQ4	0.027714
			P17117	
AMP+KA			P39451	
vs.		Microbial metabolism in diverse	P00562	
control	eco01120	environments	P76149	0.058375
AMP+KA			P39180	
vs.			P0AEJ4	
control	eco02026	Biofilm formation - Escherichia coli	P0DMC5	0.001716
AMP+KA			P15877	
vs.			P39451	
control	eco01110	Biosynthesis of secondary metabolites	P00562	0.246142
AMP+KA			P15877	
vs.			P39451	
control	eco01130	Biosynthesis of antibiotics	P00562	0.096783
AMP+KA				
VS.	00220	A · · · · · · · · · · · · · · ·	P//581	0.001002
control	eco00330	Arginine and proline metabolism	P/621/	0.001083
AMP+KA		Alexing constate and shitemate	D00069	
VS.		Alanine, aspartate and glutamate	P00968	0.002244
control	eco00250	metabolism	P/6149	0.002244
AMP+KA			DO 4 7117	
VS.	02010	Dihosoma	PUA/W/	0 026405
	eco03010	KIUOSOIIIE		0.020483
AMP+KA		True component and and	PUAEJ4	0 121000
vs.	eco02020	i wo-component system	PUDMIC5	0.131989

control				
		Chloroalkana and chloroalkana		
vs.	000625	degradation	D20451	0.000402
AMP+KA	eco00023	degradation	F 39431	0.000402
vs.				
control	eco02060	Phosphotransferase system (PTS)	P36672	0.047341
AMP+KA				
vs.	000 40		<b>D</b> 000.00	0.404055
control AMP+KA	eco00240	Pyrimidine metabolism	P00968	0.104275
vs.				
control	eco01502	Vancomycin resistance	P07862	0.001838
AMP+KA				
vs.				
control	eco02010	ABC transporters	P14175	0.424014
AMP+KA				
vs.				
control	eco00540	Lipopolysaccharide biosynthesis	P45395	0.027067
AMP+KA				
vs.				
control	eco03070	Bacterial secretion system	P0AG90	0.025452
AMP+KA		2		
vs.				
control	eco00790	Folate biosynthesis	P0ABO4	0.019416
AMP+KA	••••••		101122	01017110
VS.				
control	eco00473	D-Alanine metabolism	P07862	0 000402
AMP+KA	0000175		107002	0.000102
vs				
control	eco00071	Fatty acid degradation	P39451	0 006644
AMP+KA	0000071	Tutty dele degradation	1 57451	0.0000++
vs				
control	eco00270	Cysteine and methionine metabolism	P00562	0 028721
	0000270	Cystelle and metholine metabolism	100502	0.020721
vs.	000676	Nanhthalana degradation	<b>D</b> 20451	0.000402
	6000020	Naphthalene degradation	F 37431	0.000402
vs.	0001230	<b>Biosynthesis of amino acids</b>	D00562	0 255403
	6001230	biosynthesis of anniho acids	F00302	0.233403
vo.	acc01220	Degradation of aromatic compounds	D20/51	0 008516
	0001220	Sulfur rolay system	F 37431	0.000510
лиг+кА	eco04122	Summer relay system	r12282	0.00933

VS.				
$\Delta MP + K \Delta$				
VS.				
control	eco00350	Tyrosine metabolism	P39451	0.002351
AMP+KA		2		
vs.				
control	eco03060	Protein export	P0AG90	0.010596
AMP+KA				
VS.				
control	eco00300	Lysine biosynthesis	P00562	0.004988
AMP+KA				
VS.				
control	eco00633	Nitrotoluene degradation	P17117	0.001386
AMP+KA				
VS.	22200260	Glycine, serine and threonine	D00560	0 027579
	eco00260	metabolism	P00502	0.05/5/8
AMP+KA				
vs.	eco00500	Starch and sucrose metabolism	P36672	0.03215
AMP+KA	<b>cc</b> 000500	Staten and sucrose metabolism	1 30072	0.05215
VS.				
control	eco00261	Monobactam biosynthesis	P00562	0.001838
AMP+KA		2		
vs.				
control	eco00550	Peptidoglycan biosynthesis	P07862	0.015353
AMP+KA				
vs.				
control	eco00030	Pentose phosphate pathway	P15877	0.027067
AMP+KA				
VS.				
control	eco00670	One carbon pool by folate	P0ABQ4	0.005788
AMP+KA				
VS.	00010		D20451	0.05140
	eco00010	Glycolysis / Gluconeogenesis	P39451	0.05148
AMP+KA				
vs.	eco00650	Butanoate metabolism	P76140	0 037578
			1/0147	0.037378
VS.				
control	eco00760	Nicotinate and nicotinamide metabolism	P76149	0.014091