

## Supplementary Material

### **Signal transducer and activator of transcription (STAT) 1 and STAT3 are expressed in the human ovary and have Janus kinase 1-independent functions in the COV434 human granulosa cell line**

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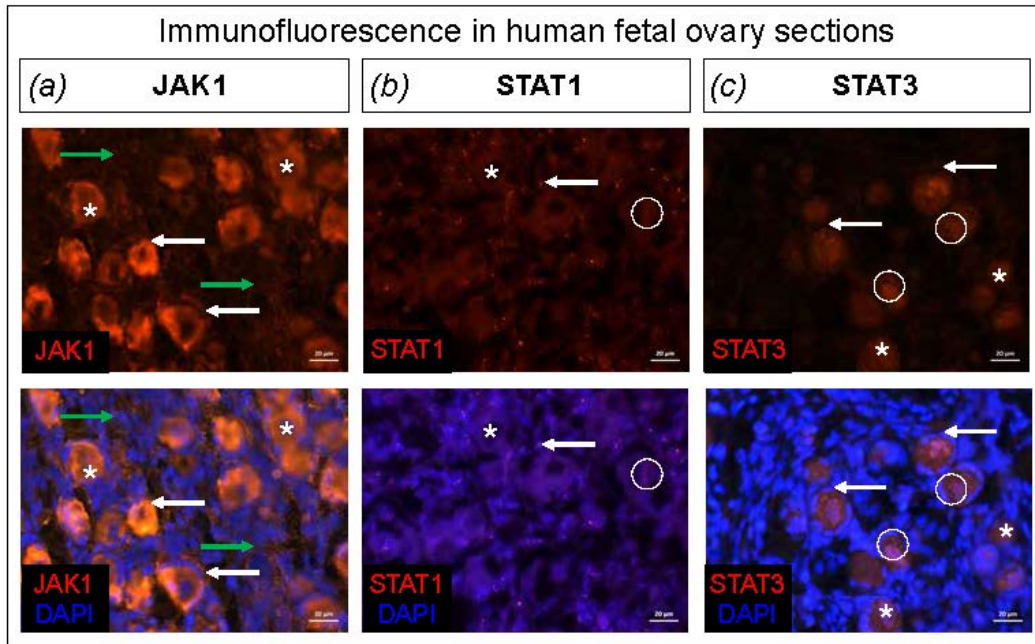
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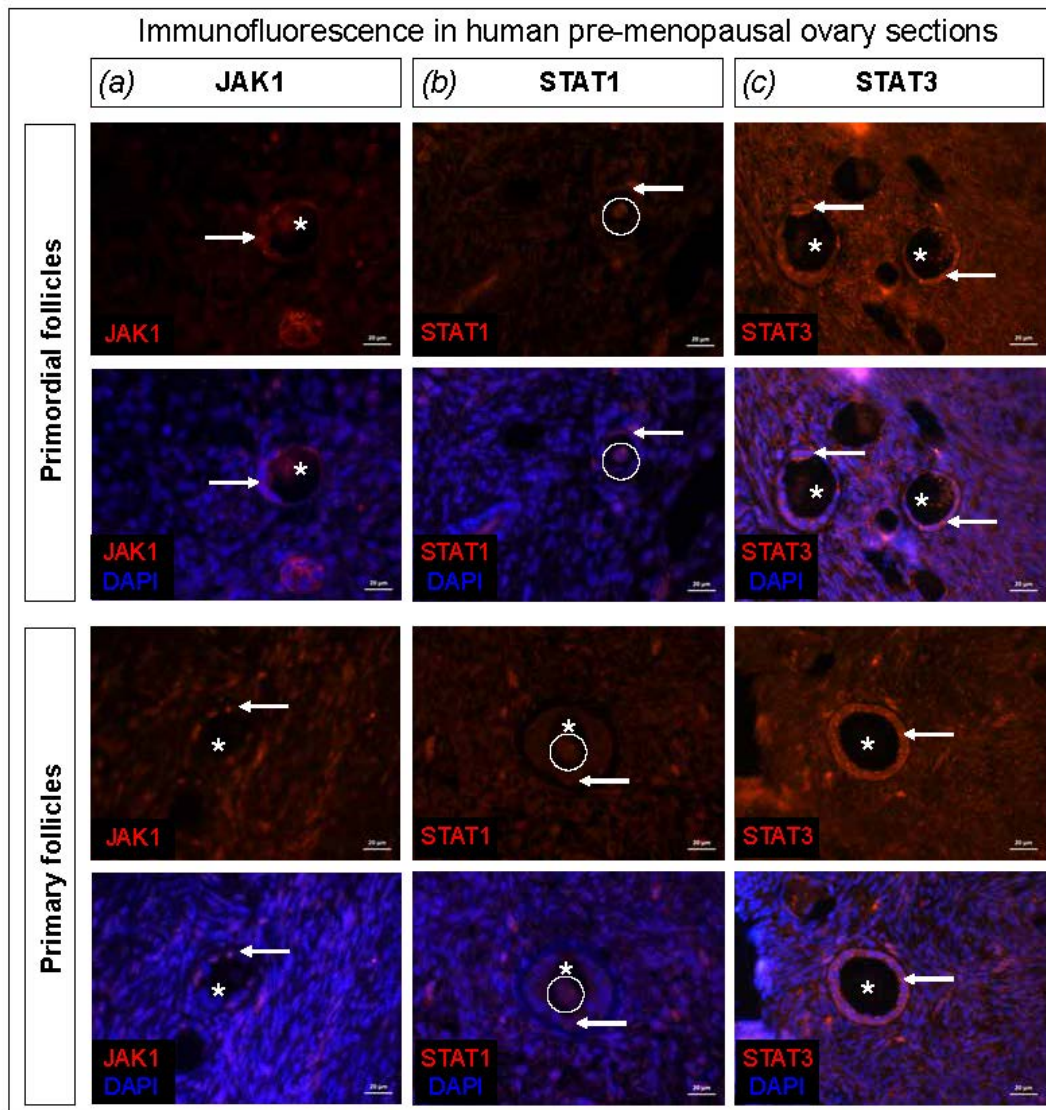
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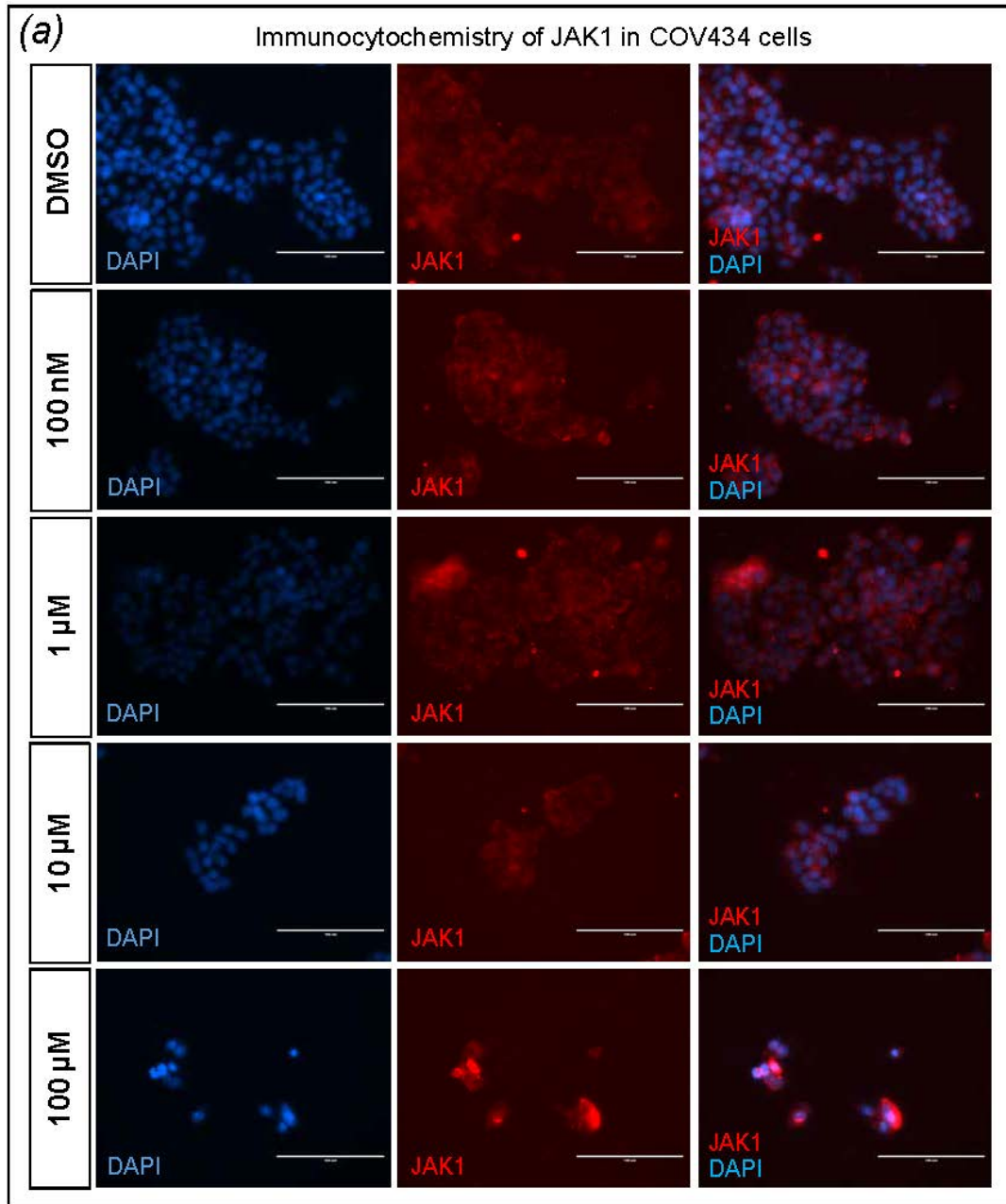


**Fig. S1.** Protein localisation of JAK1, STAT1 and STAT3 in human foetal ovarian sections. Representative protein localisation was analysed using immunofluorescence for JAK1, STAT1 and STAT3 in human foetal ovarian sections (Supplementary Figure 1a, 1b and 1c). JAK1 and STAT3 protein was expressed in both the oocyte (white asterisks) and the granulosa cells (white arrows) of primordial follicles. JAK1 was faintly detected in the stromal and interstitial cells of the foetal ovary (identified by green arrows). STAT1 showed weak protein expression in the surrounding granulosa cells, however distinct STAT1 foci were detected in the nucleus of oocytes (identified by white circles in Supplementary Figure 1b). Aggregates of STAT3 protein were observed in the nucleus of the oocytes (identified by white circles in Supplementary Figure 1c). All sections were counter-stained with DAPI (blue). Scale bar in all images is 20  $\mu\text{m}$ .

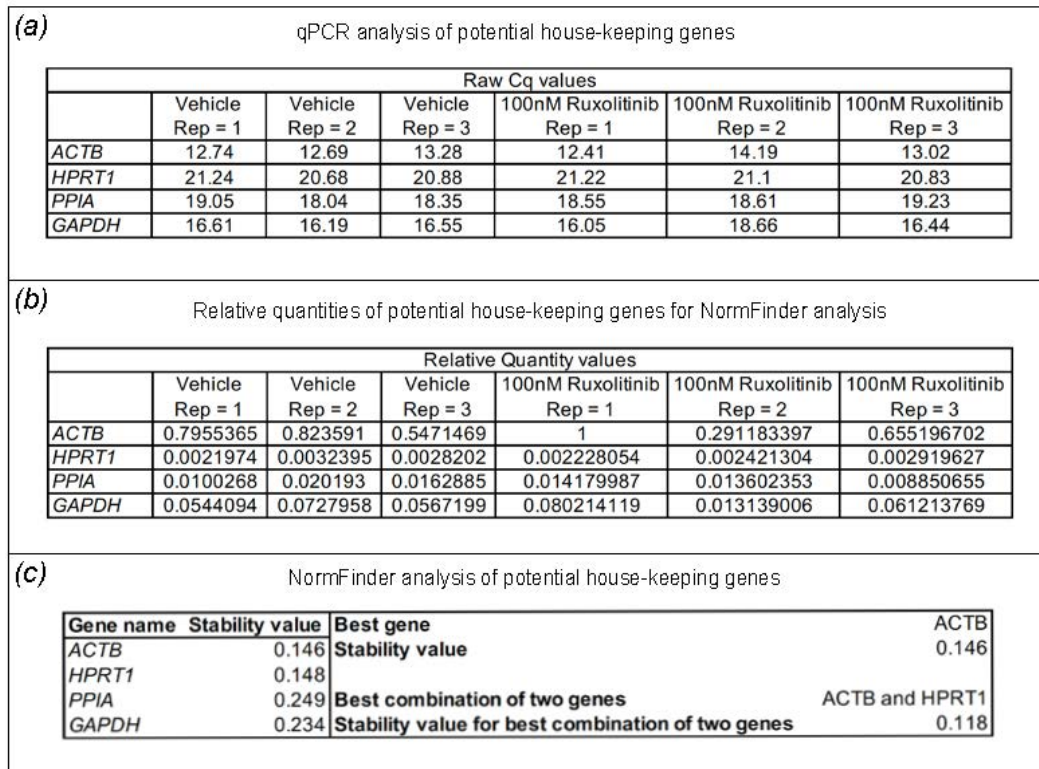


**Fig. S2.** Protein localisation of JAK1, STAT1 and STAT3 in human pre-menopausal ovarian sections. Representative protein localisation was analysed using immunofluorescence for JAK1, STAT1 and STAT3 in human pre-menopausal ovarian sections (Supplementary Figure 2a, 2b and 2c). Both primordial and primary follicles were identified in the ovary sections (according to Gougeon's criteria (Gougeon 1996)). JAK1 showed protein expression in both the oocyte (white asterisks) and the granulosa cells (white arrows) of primordial follicles (Supplementary Figure 2a). In primary follicles, however, JAK1 was restricted to the granulosa cells, and was undetectable in the oocyte (Supplementary Figure 2a). STAT1 showed weak protein expression in the

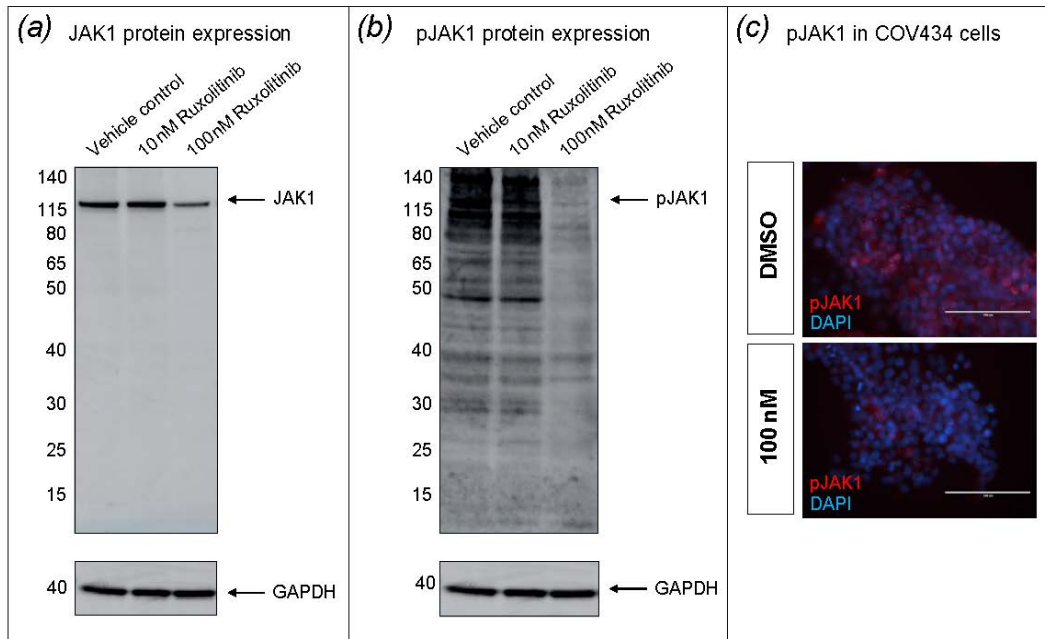
oocyte and surrounding granulosa cells (Supplementary Figure 2*b*). Aggregates of STAT1 protein were observed in the nucleus of the oocytes (identified by white circles in Supplementary Figure 2*b*). STAT3 protein was intensely localised to the pre-granulosa and granulosa cells of both follicle types (Supplementary Figure 2*c*). All sections were counter-stained with DAPI (blue). Scale bar in all images is 20  $\mu\text{m}$ .



**Fig. S3.** Optimisation of Ruxolitinib treatment doses for JAK1 inhibition. The doses of Ruxolitinib trialled were based on the IC<sub>50</sub> value of Ruxolitinib to inhibit JAK1 and previous studies using Ruxolitinib in whole ovary culture (Sutherland *et al.* 2018). Representative JAK1 protein localisation was analysed in COV434 cells following treatment with DMSO as a vehicle control, and increasing doses of Ruxolitinib (labelled) (Supplementary Figure 4a). Cells were counterstained with DAPI to mark the nucleus (blue). The scale bar in all images is 100 μm.



**Fig. S4.** Selection of human house-keeping primers for qPCR analysis. COV434 cells were treated for 72 h with 100 nM Ruxolitinib or vehicle control. The raw Cq values for *ACTB*, *HPRT1*, *PPIA* and *GAPDH* are shown in Supplementary Figure 3a, along with the relative quantities of mRNA in Supplementary Figure 3b. The relative quantities were used in the NormFinder analysis (Andersen *et al.* 2004) to identify the most suitable house-keeping gene for normalisation of the COV434 cell data. Three biological replicates of vehicle control and Ruxolitinib-treated cells were used in the analysis.



**Fig. S5.** Optimisation of Ruxolitinib treatment doses for JAK1 inhibition using immunoblotting and immunocytochemistry. As Ruxolitinib is a pan JAK1/JAK2 inhibitor, JAK1 and JAK2 protein levels with Ruxolitinib treatment were examined to determine the efficacy of the inhibitor. Representative blots showing protein expression levels in COV434 cells treated with 100 nM Ruxolitinib or vehicle control for JAK1 (Supplementary Figure 5a) and pJAK1 (Supplementary Figure 5b) are shown. Visible decreases in both JAK1 and pJAK1 are observed at the 100 nM Ruxolitinib dose (Supplementary Figure 5a and Supplementary Figure 5b). Representative pJAK1 protein localisation was analysed in cultured COV434 cells following 100 nM Ruxolitinib treatment, with DMSO as a vehicle control (Supplementary Figure 5c). Decreases in phosphorylated JAK1 were observed in Ruxolitinib treatment conditions. The scale bar in all images is 100  $\mu\text{m}$ .

**Table S1. Primer sequences for real-time polymerase chain reaction**

*JAK1*, Janus kinase 1; *STAT1*, signal transducer and activator of transcription 1;  
*STAT3*, signal transducer and activator of transcription 3; *ACTB*,  $\beta$ -actin

Gene	Primer sequence (5'-3')	Annealing temperature (°C)	Efficiency in COV434 cells (AU)
<i>JAK1</i>	Forward: AGACTTGTGAATACGTTAAAAGAAGGA	59	2.03
	Reverse: AAAGCTTGTCCGATTGGATG		
<i>STAT1</i>	Forward: CTTACCCAGAATGCCCTGAT	65	1.78
	Reverse: CGAACTTGCTGCAGACTCTC		
<i>STAT3</i>	Forward: GGTCTGGCTGGACAATATCATT	65	1.87
	Reverse: GAGGCTTAGTGCTCAAGATGG		
<i>ACTB</i>	Forward: TGTGGCATCCACGAAACTACC	65	1.96
	Reverse: ACATCTGCTGGAAGGTGGACA		

**Table S2. Immunofluorescence results showing relative expression of candidate proteins in COV434 cells**

*JAK1*, Janus kinase 1; *STAT1*, signal transducer and activator of transcription 1;  
*STAT3*, signal transducer and activator of transcription 3

Protein	Cell compartment	Relative expression of each protein compared to the cell region (immunofluorescence)
<i>JAK1</i>	Nucleus	High
	Cytoplasm	Medium
<i>STAT1</i>	Nucleus	High
	Cytoplasm	High
<i>STAT3</i>	Nucleus	Medium
	Cytoplasm	Medium