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Vitamin or salicylic acid treatments versus the adverse effects of copper stress on photosynthesis and some related activities of wheat seedlings

AM Hamada

Botany Department, Faculty of Science, Assiut University, Assiut, Egypt

Fax: 02 088 312564, e-mail: Hamada@aun.eun.eg

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Introduction

Copper is an essential microelement in biological systems, participating in a number of physiological processes as a cofactor or as part of the prosthetic group of many enzymes, primarily oxidoreductases (Clarkson and Hanson, 1980). Excess copper may result in membrane damage (Kennedy and Gonsalves, 1987), suppression of enzyme activity (Walker and Webl, 1981), cell death (Meyer and Heath, 1987) and inhibition of both light reactions of photosynthesis (Lanaras *et al.*, 1993).

In autotrophic organisms, although it is well known that vitamins are endogenously synthesized (De Gara *et al.*, 1993), yet exogenous application of these organic compounds exerted mostly positive effects on plant growth, CO₂ uptake and protein synthesis (Arrigoni *et al.*, 1997). Also, exogenous application of SA decreased the effects of low temperature, salinity stresses in some plants (Al-Hakimi and Hamada, 2001; Janda *et al.*, 1999). Thus in the present work it is intended to follow the role of AsA, B₁ and SA treatment in protection of young wheat plant against excess Cu stress.

Materials and methods

Five-day-old wheat (*Triticum aestivum* L.) seedlings were grown hydroponically in aerated Hoagland solution in a green house under natural light for 2 weeks. Copper treatment was performed by supplementing the nutrient solution with an increasing concentration of copper ions in the form of CuSO₄ (5, 10, 20 and 40 mg L⁻¹ Cu⁺⁺) and the plants were left for 3 days. Some of the plants were pre-treated with 100 ppm AsA, B₁ or SA as sodium salicylate added to the hydroponic solution for only 1 day. At the end of the experimental period fresh shoots and roots were then dried in an aerated oven at 70C. The content of chlorophylls a, b and carotenoids were determined spectrophotometrically (Metzner *et al.*, 1965). Net photosynthetic rate and dark respiration rate were determined manometrically (Umbreit *et al.*, 1959).

Results

An increase in Cu concentration from 5 to 40 mg L⁻¹ resulted in a marked inhibition of seedling growth; both fresh and dry weights of shoot and root systems decreased (Fig. 1). Pretreatment with AsA, B₁ or SA resulted, in most cases, in a pronounced increase in the production of fresh and dry matter yields in shoots and roots of Cu stressed wheat plants as compared with those of untreated plants (Fig. 1).

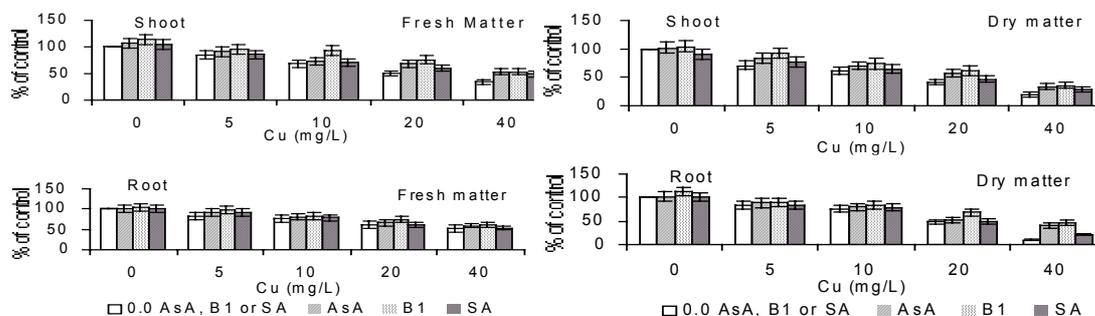


Fig. 1: The action of AsA, B₁ and SA treatments in ameliorating the adverse effects of Cu stress on fresh and dry matter of shoots and roots of wheat plants. Values in parentheses represent \pm SD.

Also the retarding effect of Cu stress on the biosynthesis of pigments fractions (Fig. 2) was partially or completely alleviated in seedlings pretreated with the AsA, B₁ or SA.

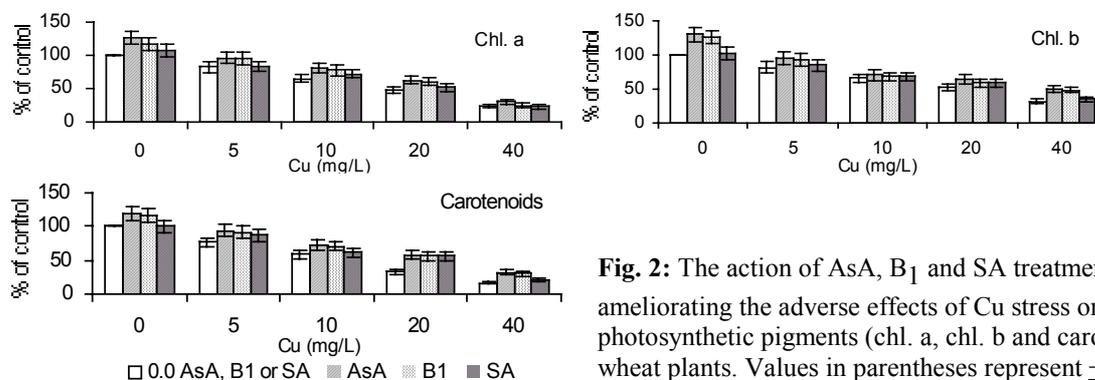


Fig. 2: The action of AsA, B₁ and SA treatments in ameliorating the adverse effects of Cu stress on photosynthetic pigments (chl. a, chl. b and carotenoids) of wheat plants. Values in parentheses represent \pm SD.

Excess Cu in the nutrient solution retarded the photosynthetic O₂ evolution (Fig. 3). In case of AsA, B₁ or SA pretreatments the net photosynthetic rate approached or exceeded those of the control especially under the low levels of Cu. Respiration of all Cu stressed plants was markedly stimulated with the rise of Cu level. With the application of AsA or B₁, the low levels of Cu failed to stimulate O₂ consumption. However SA pretreatment enhanced the stimulatory role of Cu stress in activation of dark respiration (Fig. 3).

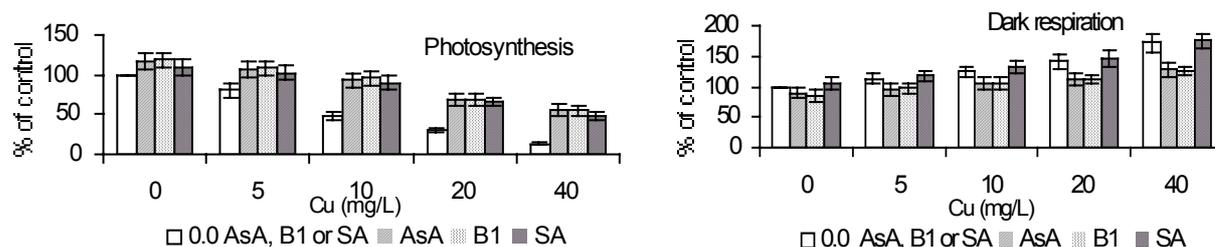


Fig. 3: The action of AsA, B₁ and SA treatments in ameliorating the adverse effects of Cu stress on net photosynthesis and dark respiration of wheat plants. Values in parentheses represent \pm SD.

Discussion

The response of wheat plants to the high levels of Cu was reflected in a manifest decrease in shoot and root growth. Vitamins or SA pretreatment alleviated completely or partially the adverse effects of Cu on growth of wheat seedlings. This means that each of the two vitamins or salicylate may act as a growth stimulant, which can play a role in reversing the effect of Cu on metabolic activities relevant to growth, like cell division and cell enlargement (Woolhouse, 1983).

The inhibited pigment biosynthesis due to Cu stress could be attributed to increased leaf senescence. Pretreatment of the seedlings with any of the two applied vitamins or SA alleviated this inhibitory effect, which may contribute directly to the effectiveness of the photosynthetic apparatus. In accordance with this, it was found that chloroplasts isolated from leaves, which were previously sprayed with AsA showed higher contents of chlorophylls than those isolated from untreated leaves (Choudhury *et al.*, 1992). Similarly, foliar application with B₁ helped some bleached plants resynthesise chlorophyll and consequently exerted positive effect on plant growth and productivity (Oertli, 1987).

The results of the present work and those obtained by other investigators (Maksymiec and Baszyski, 1996) clearly demonstrate that high applications of Cu inhibit the photosynthetic activity. These results were generally accompanied by reciprocal variations in the dark respiration activity (stimulated). Seedling pretreatment with AsA, B₁ or SA was generally effective in antagonizing partially or completely the inhibitory effects of Cu stress on the net photosynthetic rate. On the other side, the stimulatory effect on the dark respiration rate with the rise of Cu level, could be regarded as a repairing system for the cellular damage linked in some way to the suppression of growth (Lambers, 1979). On the contrary, pretreatment with SA enhanced the stimulatory effects of Cu stress on dark respiration of seedlings. These increased values of dark respiration rate indicate that SA could provoke alteration very often associated with plant responses to stressful conditions (Rhoads and McIntosh, 1991).

From the preceding results and discussion, it can be concluded that pretreatment of wheat seedlings with AsA, B₁ or SA could alleviate the inhibitory effects of Cu stress and also stimulate the growth *via* enhancement of the photosynthetic rate.

References

- Al-Hakimi AMA, Hamada AM (2001) Counteraction of salinity stress on wheat plants by grain soaking in ascorbic acid, thiamin or sodium salicylate. *Biologia Plantarum* **44**, 253-261.
- Arrigoni O, Calabrese G, De Gara L, Bitonti M, Lio R (1997) Correlation between changes in cell ascorbate and growth of *Lupinus albus* seedlings. *Journal Plant Physiology* **150**, 302-308.
- Choudhury NK, Aslam M, Huffaker RC (1992) Aging of cell free chloroplasts. Photoprotection of pigments and photochemical activity by zeaxanthin. *Plant Physiology* **99**, 26-35.
- Clarkson DT, Hanson JB (1980) The mineral nutrition of higher plants. *Annual Review of Plant Physiology* **31**, 239-298.
- De Gara L, Paciolia C, Liso P, Stefani A, Blanco A, Arrigoni O (1993) Ascorbate metabolism in mature pollen grains of *Dasypyrum villosum* (L.) Borb. During imbibition. *Journal Plant Physiology* **141**, 504-509.

- Janda T, Szalai G, Tari I, Paldi E (1999) Hydroponic treatment with salicylic acid decreases the effects of chilling injury in maize (*Zea mays*L.) plants. *Planta* **208**, 175-180.
- Kennedy CD, Gonsalves FAN (1987) The action of divalent zinc, cadmium, mercury, copper and lead on the trans-root potential and H⁺ efflux of excised roots. *Journal of Experimental Botany* **38**,800-817.
- Lambers H (1979) Efficiency of root respiration in relation to growth, morphology and soil composition. *Physiologia Plantarum* **46**, 194-202.
- Lanaras T, Moustakas M, Symeonidis L, Diamantoglou S, Karataglis S (1993) Plant metal content, growth responses and some photosynthetic measurements on field-cultivated wheat growing on ore bodies enriched in Cu. *Physiologia Plantarum* **88**, 307-314.
- Maksymiec W, Baszyski T (1996) Chlorophyll fluorescence in primary leaves of excess Cu-treated runner bean plants depends on their growth stages and the duration of Cu-action. *Journal of Plant Physiology* **149**, 196-200.
- Metzner H, Rau H, Sengen H (1965) Untersuchungen zur Synchronisierbarkeit einzelner Pigment Mangel Mutanten von *Chlorella*. *Planta* **65**, 186-194.
- Meyer SLF, Heath MC (1987) A comparison of the death induced by fungal invasion or toxic chemicals in cowpea epidermal cells. I. Cell death induced by heavy metal salts. *Canadian Journal of Botany* **66**, 613-623.
- Oertli JJ (1987) Exogenous application of vitamins as regulators for growth and development of plants. A review. *Z. Pflanzenernähr Bodenk* **150**, 375-391.
- Rhoads DN, McIntosh L (1991) Isolation and characterization of *Matum guttatum* (Schett). *Proceedings National Academic Science USA* **88**, 2122-2126.
- Umbreit WW, Burries RH, Stauffer JE (1959) Manometric Techniques. A Manual Describing Methods Applicable to the Study of Tissue Metabolism. Burgess Publishing Co., Minneapolis.
- Walker CD, Webl J (1981) Copper in plants. Forms and behaviours. In soils and Plants (JF Loneragan, AD Robson and RD Graham, eds), pp.189-212. Academic Press, London. ISBN 0-12-455520-9.
- Woolhouse HW (1983) Toxicity and tolerance in the responses of plants to metals. In: Lage, O.L. *et al.* (eds.): Encyclopedia of Plant Physiology, New Series, Vol. **12C**, pp. 245-300. Springer-Verlag, Berlin and New York.