Non-linear response of soil N₂O emissions to nitrogen fertiliser in a cotton-fallow rotation in sub-tropical Australia

Clemens Scheer^{A,B}, David W. Rowlings^A, and Peter R. Grace^A

^AInstitute for Future Environments, Queensland University of Technology, Brisbane, Qld 4000, Australia. ^BCorresponding author. Email: clemens.scheer@qut.edu.au

Abstract. Nitrogen (N) fertiliser is a major source of atmospheric nitrous oxide (N₂O), and over recent years there has been growing evidence for a non-linear, exponential relationship between N fertiliser application rate and N₂O emissions. However, there is still a high level of uncertainty around the relationship of N fertiliser rate and N₂O emissions for many cropping systems. We conducted year-round measurements of N₂O emission and lint yield in four N-rate treatments (0, 90, 180 and 270 kg N ha⁻¹) in a cotton–fallow rotation on a black vertosol in Australia. We observed a non-linear exponential response of N₂O emissions to increasing N fertiliser rates with cumulative annual N₂O emissions of 0.55, 0.67, 1.07 and 1.89 kg N ha⁻¹ for the four respective N fertiliser rates, but no N response to yield occurred above 180 kg N ha⁻¹. The annual N₂O emission factors induced by N fertiliser were 0.13, 0.29 and 0.50% for the 90, 180 and 270 kg N ha⁻¹ treatments respectively, significantly lower than the IPCC Tier 1 default value of 1.0%. This nonlinear response suggests that an exponential N₂O emissions model may be more appropriate for estimating emission of N₂O from soils cultivated to cotton in Australia. It also demonstrates that improved agricultural N-management practices can be adopted in cotton to substantially reduce N₂O emissions without affecting yield.

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Introduction

Nitrous oxide (N₂O) contributes to the greenhouse effect due to its high global warming potential (nearly 300 times greater than carbon dioxide over a 100-year horizon) and is also the largest ozone depleting substance of the 21st century (IPCC 2007; Ravishankara et al. 2009). Increased use of nitrogen (N) fertiliser and animal manure are the main sources of atmospheric N₂O. Globally, croplands are responsible for 66% of total anthropogenic N₂O emissions and these emissions are predicted to double by 2050 (Davidson and Kanter 2014). In soils, N₂O is mainly produced by the microbial processes of autotrophic nitrification (oxidation of ammonium to nitrate) and heterotrophic denitrification (reduction of nitrate to N₂O and ultimately N_2). The magnitude of these emissions is strongly affected by soil conditions and agricultural management, in particular the addition of N fertiliser (Butterbach-Bahl et al. 2013).

Total added N fertiliser is the most important predictor of N_2O emissions from cropping land, and early studies showed a linear relationship between N input and direct N_2O emission from the soil (Bouwman 1996). Consequently, the IPCC adopted a linear relationship for its Tier 1 emission factor (EF) methodology (IPCC 1997, 2006). This EF approach assumes that a fixed amount of N added as fertiliser is transformed and emitted as N_2O , and is currently still being used for most national greenhouse gas (GHG) inventories (IPCC 2006). However, the relationship between N fertiliser rate and N_2O emission is complex and few studies have measured N_2O emissions under more than two N fertiliser rates to establish clear functional

relationships. Consequently, there is still a high degree of uncertainty in the relationship between N fertiliser rate and N_2O emission for many cropping systems.

In recent years there has been growing evidence for a nonlinear, exponential relationship from a range of cropping systems, particularly for fertiliser rates that greatly exceed crop requirements (Hoben *et al.* 2011; Kim *et al.* 2013; McSwiney and Robertson 2005). In a recent global meta-analysis of 78 different studies, Shcherbak *et al.* (2014) found a general trend of exponentially increasing N₂O emissions as N fertiliser rates increased above crop N demand for the majority of crop types examined. However, such data do not currently exist for cotton cropping systems. A non-linear response of N₂O emissions to N fertiliser additions means that N₂O emission factors are not constant and will depend on N input rates. A clear understanding of this relationship is crucial for both N₂O inventory estimates and the design of effective mitigation strategies.

Cotton production provides an ideal opportunity to investigate the relationship between N₂O emissions and N fertiliser inputs due to high N fertiliser (up to 400 kg N ha⁻¹) and irrigation inputs, and significant emissions of N₂O from irrigated cotton fields have been reported (Scheer *et al.* 2013). In Australia, cotton is grown on almost 600 000 ha in the inland regions of northern New South Wales and southern Queensland. However, there are still limited data available on the effect of fertiliser N rate on N₂O emissions and to date no emission factors have been reported for a cotton–fallow rotation in Australia over a full year. Consequently, this field-based study measured N₂O emissions from a typical cotton–fallow rotation with four N fertiliser rates over one year in southeast Queensland, Australia. The two main objectives of the study were: (i) to investigate N_2O emissions (including EFs) and lint yield from a cotton–fallow rotation in response to increasing N fertiliser rate; and (ii) test the assumption that N_2O emissions increase linearly in response to N fertiliser rates.

Material and methods

Study site

The field experiment was conducted during the 2010/11 cotton season at the Agri-Science Queensland, Department of Employment, Economic Development and Innovation (DEEDI) Kingsthorpe Research Station. The station is located in the Darling Downs region ~140 km west of Brisbane (27°31'S, 151°47'E, 431 m above mean sea level). Prior to the present study, the field site was used for an irrigation study under a cotton-wheat rotation for which crop residues were removed from the plots after harvest (Scheer et al. 2012, 2013). The field site included an overhead sprinkler irrigation system and so there were no ridges or furrows. The region has a sub-tropical climate with an average annual precipitation of 630 mm (1990–2010) (Commonwealth Bureau of Meteorology, www. bom.gov.au/climate) with most rainfall during October-March in the summer crop growing season. The soil at the site is classified as a haplic, self-mulching, black vertosol using the Australian Soil Classification (Isbell 2002). It has a heavy clay texture (76% clay) in the 1.5-m root zone profile, with a distinct change in soil colour from brownish black (10YR22) in the top 90 cm to dark brown (7.5YR33) deeper in the profile. The soil is formed in an alluvial fan of basalt rock origin, slowly permeable, with a surface slope of ~0.5%. Physical and chemical characteristics of the soil profile are shown in Table 1.

Experimental design

The fertiliser response trial was set up as a randomised complete block design. Each block was replicated three times, with four N fertiliser rates in each block. Each experimental plot was 13 m wide \times 20 m in length, with the cotton crop (Gossypium *hirsutum* L. cv. Bollgard[®] II) planted in a north–south orientation. A buffer zone (4 m) was planted between plots and an access track (4 m) was located at the centre of the research area. Cotton was planted on 5 November 2007 at a density of ~17 seeds m⁻¹, a depth of 4 cm and a row spacing of 1 m. The aim was to get an established stand of 11–12 plants m⁻¹, which is the recommended density for Bollgard[®] II. Weeds were controlled by a combination of manual chipping and a chemical

Table 1. Physical and chemical soil characteristics of the experimental site Kingsthorpe Research Station, Queensland, Australia

Soil property	0–10 cm	
Organic carbon (%)	1.6	
Total nitrogen (%)	0.16	
pH (H ₂ O)	7.3	
Texture (USDA)	Clay	
Cation exchange capacity $(meq^+/100 g)$	70	
Clay (%)	76	
Silt (%)	16	
Sand (%)	6	

control (1 kg ha⁻¹ of glyphosate). Only N fertiliser was applied at sowing and evenly broadcasted by hand to each replicate plot to prevent possible inaccuracy in N₂O measurements due to uneven fertiliser distribution.

The N fertiliser treatments were

- 1. Zero nitrogen fertiliser (0N) i.e. no added fertiliser;
- 90 kg N ha⁻¹ (90N) 90 kg N ha⁻¹ urea basal application at planting (4 November 2010);
- 180 kg N ha⁻¹ (180N) 90 kg N ha⁻¹ urea basal application at planting (4 November 2010) and 90 kg N ha⁻¹ urea in two side dressings (5 January and 3 March 2011);
- 270 kg N ha⁻¹ (270N) 90 kg N ha⁻¹ urea basal application at planting (4 November 2010) and 180 kg N ha⁻¹ urea in two side dressings (5 January and 3 March 2011).

The plots could be irrigated individually with bore water using a hand-shifted sprinkler and partial-circle sprinkler heads to avoid irrigating adjacent plots. However, during this study, growing season rainfall exceeding the typical water demand (600–800 mm) of cotton and irrigation did not occur.

N₂O flux measurement

The N₂O fluxes were measured over an entire year, including the cotton cropping season from 5 November 2010 to 9 June 2011 and the following fallow phase from 9 June to 15 November 2011. Emissions were measured using the closed chamber technique using quality criteria as outlined by de Klein and Harvey (2013) and Parkin and Venterea (2010). This method uses a gas-tight chamber which encloses soil for a given interval. The chamber consists of a frame inserted a few centimetres into the soil and a lid that is fixed to the frame throughout the sampling period. Chamber enclosure is achieved by a sealed gasket at the lower edge of the lid. We used cylindrical PVCchambers with an inner diameter of 22.5 cm and a height of 20 cm that were randomly inserted between the plant rows (which were 1 m apart) in each plot (i.e. the measurements did not account for potential N₂O emissions directly from the cotton plants). The volume of each chamber was $\sim 0.008 \text{ m}^3$ and the cross-sectional area was 0.04 m². Fluxes were measured by collecting air samples from the chamber head space. Of headspace air, 20 mL was drawn through a septum into gas-tight 20-mL polypropylene syringes at 0, 30 and 60 min after the soil was covered and inserted into evacuated vials (Exetainers[®]). Chamber temperature was monitored during the measurement using an electronic temperature sensor. The gas samples were then analysed for N₂O using a gas chromatograph (Shimadzu GC-2014, Kyoto, Japan) equipped with an electron capture detector.

The N₂O flux rates were measured from three replicated chambers per experimental plot (n=9 per treatment) to minimise the error associated with the spatial variability of N₂O emissions over the cotton growing period. The number of chambers was decreased to one replicated chamber within each treatment per experimental plot (n=3 per treatment) over the fallow period when only small fluxes (<5.0 g N₂O-N ha⁻¹ day⁻¹) were observed in all treatments. Sampling frequency was optimised according to the recommendations of Reeves and Wang (2015). Specifically, measurements were conducted three times a

week immediately after fertilisation followed by heavy rainfall events and weekly over the remaining period, which was expected to provide a highly accurate estimate ($\pm 10\%$ error) compared with measurements with a sub-daily temporal resolution (Reeves and Wang 2015). Fluxes were measured once on each sampling day during 0900–1100 hours, which has been shown to best approximate the daily mean N₂O flux (Reeves and Wang 2015).

Ancillary measurements

An EnviroStation (ICT International Pty Ltd, Armidale, NSW, Australia) electronic weather station was installed at the research site to measure local weather variables. The station recorded both daily and hourly values of solar radiation, air temperature (maximum, minimum and average), relative humidity, wind speed and rainfall. Volumetric soil water content was measured in the surface soil (0-10 cm) in each experimental plot at each gas sampling using a hand-held MP406 standing wave soil moisture probe (ICT International Pty Ltd) that was calibrated for the soil at the research site. Water-filled pore space (WFPS) was calculated by dividing volumetric water content by total porosity. Total porosity was calculated as $[1 - (bulk density/particle density)] \times 100\%$] using measured soil bulk density data (arithmetic means of four samples) and an assumed particle density of 2.65 g cm⁻³ (Barton et al. 2008). Additionally, at the beginning of the growing season, bulk soil samples were taken from each plot by combining 5-10 soil cores (0-10 cm depth) and analysed for soil texture, total carbon (C%) and total nitrogen (N%). Seed cotton and lint yield were determined in each plot by harvesting a 2.5 m length of two cotton rows outside the chamber area by hand and the seed cotton was ginned.

Statistical analysis and calculations

The N₂O emissions were calculated from the linear increase of the gas concentration at each sampling time (0, 30 and 60 min during the time of chamber closure), adjusted for area and volume of the chamber and corrected for chamber temperature and air pressure as described by Scheer *et al.* (2014). The coefficient of determination (R^2) for the linear regression was calculated and used as a quality check for the measurement. For $R^2 < 0.9$ ($R^2 < 0.7$ for small flux rates <5.0 g N₂O-N ha⁻¹ day⁻¹) the measurement was rejected and not used in subsequent analyses.

Annual N_2O emissions from each plot were calculated by integrating hourly losses with time. Days where fluxes were not measured were filled by linear interpolation across missing days. Emission factors of the N fertiliser applied to the soil were calculated using:

$$EF = \frac{N_2O-N(treatment) - N_2O-N(0N)}{Total N applied} \times 100\%$$

where EF is the percentage of the total fertiliser N applied that was emitted as N₂O-N, N₂O-N is the total N₂O over one year (kg N ha⁻¹ year⁻¹) for a N fertiliser rate and total N applied is the amount of N fertiliser applied (kg N ha⁻¹ year⁻¹). Effects of treatment on total emissions were assessed by two way analysis of variance, which estimated variability due to experimental block and treatments. The null hypothesis significance test for treatment was conducted using an *F*-ratio test. Treatment effects on average total emissions were compared statistically by comparison with a least significant difference calculated at 5% critical value. Statistical analysis was undertaken using SPSS 16.0 (SPSS Inc., USA).

Results

Seasonal variability of environmental and soil conditions

Over the one-year observation period, 1260 mm of rainfall was recorded at the study site. This rainfall was double the long-term annual precipitation (630 mm). The study site received exceptionally high rainfall over a two-month period from mid-November 2010 to mid-January 2011. Over 700 mm of rainfall was recorded during this two-month period, including several heavy rainfall events exceeding 50 mm on a single day (Fig. 1). Over 1000 mm of rainfall was recorded during the entire cotton cropping period, exceeding the typical water demand (600–800 mm) of cotton at the experimental site. Consequently there was no irrigation over the 2010/11 cotton cropping period. The mean air temperature during the study period was 18.4°C; maximum hourly air temperature (38.9°C) was recorded in November and minimum hourly air temperature (-5.9°C) in August 2011 (Fig. 1).

Soil WFPS in the surface soil (0-10 cm) varied over the year in response to rainfall. Soil WFPS was high over the cotton growing season (57-100%), due to frequent rainfall at the onset of the study and the high water holding capacity of the clay; whereas WFPS was significantly lower (38-91%) over the fallow period (Fig. 1). The lowest calculated WFPS (38%)was in August 2011 after an extended dry period.

Influence of N fertiliser rates on N_2O emissions and cotton yield

Average N_2O flux (over 1 year) was 5.2, 2.9, 1.8 and 1.5 g N_2O - $N ha^{-1} day^{-1}$ in the 270N, 180N, 90N and 0N treatments

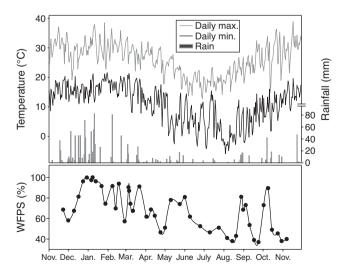


Fig. 1. Maximum and minimum hourly air temperature, daily precipitation and water filled pore space (WFPS) (0–10 cm) over the one-year observation period at the Kingsthorpe Research Station, Australia (November 2010 to November 2011).

respectively, corresponding to cumulative annual emissions of 1.89, 1.07, 0.67 and 0.55 kg of N emitted as N₂O (Table 2). There was a clear non-linear response of N₂O emissions to N fertiliser rates, which was best described by a non-linear exponential growth function ($R^2 = 0.99$, Fig. 2).

There was no significant difference in annual N_2O emissions from 0N and 90N treatments, but there was a significant increase with increasing fertiliser rate for 180N and 270N treatments. EFs were 0.13, 0.29 and 0.50% of the total amount of mineral N applied to the plots for the 90N, 180N and 270N treatments respectively (Table 2).

There was a significant effect of N fertiliser rate on average lint yield. Lint yield was highest in the 270N with 1.34 tha^{-1} and was not significantly different to the 1.24 tha^{-1} of the 180N treatment. There was also no significant difference in lint yield between the 0N (0.89 tha^{-1}) and the 90N (1.05 tha^{-1}) treatments, but average yield was significantly lower in the 0N than the 180N and 270N treatments (Table 2). Lint percentage of the seed cotton was ~43% and did not vary with N fertiliser rate. These yields were lower than the average cotton yield in Australia in the 2010/11 season (1.5 tha^{-1}) (CRDC 2014), but within the range of yields reported from previous cotton trials at the same site (Payero 2010; Scheer *et al.* 2013).

Table 2. Average and annual N_2O fluxes, cotton lint yield and emission factors from four fertiliser rate treatments with standard error (s.e.) of the means and least significant difference (l.s.d.) at 5% critical value

	Average N ₂ O flux $(g N ha^{-1} day^{-1})$	Annual N ₂ O flux $(kg N ha^{-1} year^{-1})$	Lint yield $(t ha^{-1})$	Emission factor (%)
0N	1.50	0.55	0.89	_
90N	1.82	0.67	1.05	0.13
180N	2.93	1.07	1.24	0.29
270N	5.18	1.89	1.34	0.50
s.e.	0.33	0.12	0.06	0.04
1.s.d.	1.31	0.48	0.28	0.13

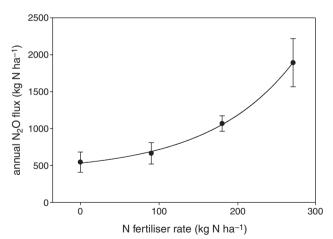


Fig. 2. N₂O emissions vs N fertiliser rate over the cotton–fallow rotation at the Kingsthorpe Research Station, Queensland. Cumulative annual N₂O emissions (g N ha⁻¹) vs N fertiliser rate (kg N ha⁻¹) for four N fertiliser rate treatments (0, 90, 180 and 270 kg N ha⁻¹); error bars indicate the standard error of the mean over the measurement period. The exponential growth function N₂O=411.2+121.1e^(0.0093 × N rate) was the best-fit equation (R^2 =0.99).

Seasonal variability of N₂O emissions

The N₂O emissions showed typical high temporal and spatial variability as frequently reported for soil N₂O fluxes (Fig. 3). Seasonal N₂O fluxes during the cotton cropping period showed a high degree of spatial variability, with the coefficients of variation across replicate chambers (n=9) in the range of 20-53%. The majority of N₂O fluxes occurred over the cotton growing period when there was a clear influence of fertilisation events on N2O emissions. Highest emissions occurred after heavy rainfall following fertilisation and there was a significant effect of the N fertiliser rate on the magnitude of the flux. For example, the first N₂O peak occurred following the first rainfall after planting on 18 November, reaching 18.2 g N₂O-N ha⁻¹ day⁻¹ in the 90N, 180N and 270N treatments and was significantly lower $(7.1 \text{ g N}_2\text{O-N} \text{ ha}^{-1} \text{ day}^{-1})$ in the 0N treatment. In the 0N and 90N treatments, for which no additional fertiliser was applied after planting, there were no further N2O emission peaks; however, in the 180N and 270N treatments, two additional emission peaks occurred after the application of N fertiliser and subsequent rainfall in January and March 2011. Overall highest emissions were observed on 13 January and reached $72 \text{ g } \text{N}_2\text{O-N} \text{ ha}^{-1} \text{ day}^{-1}$ in the 270N treatment after the application of 90 kg N urea ha⁻¹ on 5 January was followed by heavy rainfall events (50 mm on 6 January and 78 mm on 11 January). The magnitude of this 'emission pulse' was significantly lower $(28.1 \text{ g N}_2\text{O-N ha}^{-1} \text{ day}^{-1})$ in the 180N treatment, for which only 45 kg N urea ha⁻¹ was applied on 5 January. From April onwards there were no significant differences in N₂O emissions between the treatments and only small fluxes ($< 5.0 \text{ g N}_2\text{O-N ha}^{-1} \text{ day}^{-1}$) were observed in all treatments.

Discussion

Annual N₂O emissions (0.55–1.9 kg N₂O-N ha⁻¹) were in good agreement with values reported from other cotton systems in Australia (Macdonald *et al.* 2015; Scheer *et al.* 2013), but at the lower end of emissions from cotton farming systems in other countries. In northern China, Liu *et al.* (2010) observed annual emissions of 2.6 kg-N ha⁻¹ year⁻¹ from irrigated cotton fertilised with 66 kg N ha⁻¹ and Scheer *et al.* (2008) reported seasonal emissions of 0.9–6.5 kg N ha⁻¹ from a range of different irrigated cotton systems in Uzbekistan (fertiliser rates of 162.5–250 kg N ha⁻¹). These results confirm that the N₂O emission potential from cotton on vertosols in Australia is generally low, most likely due to limited availability of labile carbon in the soil and the neutral to alkaline soil pH, which restricts denitrification activity and increases the N₂/N₂O emissions ratio (Scheer *et al.* 2012).

This study showed a non-linear exponential response of N_2O emissions to N fertiliser rates that may be typical for fertilised cotton systems on black vertosols in Australia. In our experiment, high rates of N fertiliser led to increased emissions of N_2O with no significant effect on yield. We observed a clear non-linear response of N_2O emissions to N fertiliser rates. There was no significant difference in cumulative N_2O emissions between the 0N and 90N treatments, indicating that soil microbes responsible for N_2O production had limited access to N. The N_2O fluxes in the 180N and 270N treatments

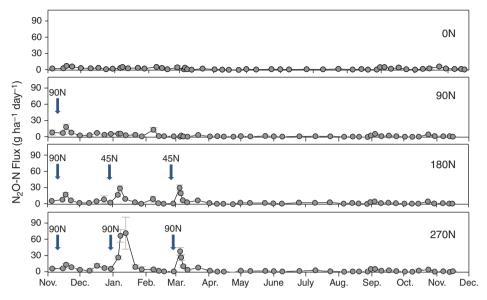


Fig. 3. Daily N₂O fluxes for the four fertiliser rate treatments (0N, 90N, 180N and 270N) over the cotton-fallow rotation at the Kingsthorpe Research Station, Queensland. Error bars indicate the standard error of the means (n=9 for the cotton growing period, n=3 for the fallow period). Arrows indicate the amount and timing of N fertiliser applications.

increased by 60 and 184% respectively, compared with the 90N treatment. Previous studies reported both linear and nonlinear responses of cumulative N₂O emissions to N fertiliser rates (Hoben et al. 2011; Lebender et al. 2014; Liu et al. 2012; Ma et al. 2010; McSwiney and Robertson 2005). However, recent metadata analyses suggest a non-linear response of direct N2O emissions to increased N additions (Kim et al. 2013; Shcherbak et al. 2014). It is not entirely clear what drives the relationship of N₂O emissions to N input in different agro-ecosystems. In theory, a linear response is expected in N-limited systems where N₂O emission is primarily controlled by the competition of plants vs microbes for available N. A non-linear exponential response is expected as soon as N fertiliser application exceeds plant demand, and then small increases in N fertiliser rates will result in disproportionally higher N₂O fluxes at higher N application rates. Once N addition increases beyond the capacity of soil microbes to take up and utilise N, the rate of increase for N₂O production would slow and finally reach a steady state (Kim et al. 2013).

In this study, N₂O emissions were mainly controlled by the combined impact of fertilisation and rainfall. Highest emissions occurred after heavy rainfall following fertilisation and there was a significant effect of the N fertiliser rate on the magnitude of the flux. This is consistent with previous studies on N₂O emissions from cotton where highest emissions following irrigation or rainfall immediately after fertiliser N application have been reported (Liu *et al.* 2010; Scheer *et al.* 2008, 2013). Consequently, the non-linear increase in N₂O emissions was mainly caused by highly elevated emissions after the side dressing of N fertiliser in the 180N and 270N treatments at the beginning of January and beginning of March 2011 that were followed by heavy rainfall. Applying double the N rate in the 270N treatment (90 kg N ha⁻¹) in January increased the emission pulse by almost four-fold compared with the 180N

treatment (45 kg N ha⁻¹). The strong increase in N₂O emissions with the higher N fertiliser rate shows that at that stage the conditions were such that N supply greatly exceeded crop demand and other factors such as soil moisture or soil temperature were not limiting. This indicates that a large amount of unused N was available for soil microbes responsible for N₂O production, and resulted in a higher proportion of the applied N fertiliser being lost as N₂O to the environment. There was no significant difference in lint vield between the 180N and 270N treatments, suggesting an optimal fertiliser range of 180-270 kg N ha⁻¹ for maximum yield in the investigated cotton system. It should be noted that this was not a N-response trial to determine optimum N rates, which would require more N rates; however, the overall trend is in good agreement with previous results from Australian cotton systems where an optimal economic N rate of 200 kg N ha⁻¹ has been reported (Macdonald et al. 2015; Rochester 2012). Above the optimal fertiliser rate, N₂O emissions will increase exponentially with no significant effect on yield.

The annual N₂O EFs, induced by N fertiliser in the present study, were 0.13, 0.29 and 0.50% for the 90N, 180N and 270N treatments respectively (Table 2). These are lower than the IPCC default value used from global inventories (1% of N applied (IPCC 2006)) but in reasonable agreement with the EF for irrigated cotton (0.5%) used by the Australian Government for their national GHG Inventory report (ANGA 2010). It is also at the lower end of EFs reported for other irrigated cotton systems with 0.12–4.0% (Liu *et al.* 2010; Macdonald *et al.* 2015; Scheer *et al.* 2008). This highlights the need for differentiated EFs that take the non-linear response to N fertiliser rates into account to reliably estimate emissions from different agricultural systems. More data is required to assess N₂O emissions as a function of added N for other intensively fertilised systems in Australia.

Conclusion

This study demonstrated a non-linear exponential response of N_2O emissions to N fertiliser rates that may be typical for fertilised cotton systems on black vertosols in Australia. Corresponding EFs increased from 0.13 to 0.50% when N fertiliser rates increased from 90 to 270 kg N ha⁻¹, but there was no significant increase in yield between the 180 and 270 kg N ha⁻¹ treatments. The study confirmed that an optimised fertiliser strategy can be adopted in cotton to substantially reduce N_2O emissions without affecting yield potential, corroborating previous studies in cotton systems. More studies on the effect of N fertiliser on N_2O emissions are required to develop N_2O response curves for other intensively fertilised systems in Australia. This study highlights the potential to reduce N losses to the environment by improved agricultural N-management practices.

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