## Greenhouse gas (N<sub>2</sub>O and CH<sub>4</sub>) fluxes under nitrogen-fertilised dryland wheat and barley on subtropical Vertosols: risk, rainfall and alternatives

Graeme D. Schwenke<sup>A,D</sup>, David F. Herridge<sup>B</sup>, Clemens Scheer<sup>C</sup>, David W. Rowlings<sup>C</sup>, Bruce M. Haigh<sup>A</sup>, and K. Guy McMullen<sup>A</sup>

<sup>A</sup>Tamworth Agricultural Institute, New South Wales Department of Primary Industries, Tamworth, NSW 2340, Australia.

<sup>B</sup>School of Environmental and Rural Science, University of New England, Armidale, NSW 2351, Australia.

<sup>C</sup>Institute for Future Environments, Queensland University of Technology, Brisbane, Qld 4000, Australia.

<sup>D</sup>Corresponding author. Email: graeme.schwenke@dpi.nsw.gov.au

**Abstract.** The northern Australian grains industry relies on nitrogen (N) fertiliser to optimise yield and protein, but N fertiliser can increase soil fluxes of nitrous oxide (N<sub>2</sub>O) and methane (CH<sub>4</sub>). We measured soil N<sub>2</sub>O and CH<sub>4</sub> fluxes associated with wheat (*Triticum aestivum*) and barley (*Hordeum vulgare*) using automated (Expts 1, 3) and manual chambers (Expts 2, 4, 5). Experiments were conducted on subtropical Vertosol soils fertilised with N rates of  $0-160 \text{ kg N ha}^{-1}$ .

In Expt 1 (2010), intense rainfall for a month before and after sowing elevated  $N_2O$  emissions from N-fertilised (80 kg N ha<sup>-1</sup>) wheat, with 417 g N<sub>2</sub>O-N ha<sup>-1</sup> emitted compared with 80 g N<sub>2</sub>O-N ha<sup>-1</sup> for non-fertilised wheat. Once crop N uptake reduced soil mineral N, there was no further treatment difference in N<sub>2</sub>O. Expt 2 (2010) showed similar results, however, the reduced sampling frequency using manual chambers gave a lower cumulative N<sub>2</sub>O. By contrast, very low rainfall before and for several months after sowing Expt 3 (2011) resulted in no difference in N<sub>2</sub>O emissions between N-fertilised and non-fertilised barley. N<sub>2</sub>O emission factors were 0.42, 0.20 and -0.02 for Expts 1, 2 and 3, respectively. In Expts 4 and 5 (2011), N<sub>2</sub>O emissions increased with increasing rate of N fertiliser. Emissions were reduced by 45% when the N fertiliser was applied in a 50 : 50 split between sowing and mid-tillering, or by 70% when urea was applied with the nitrification inhibitor 3,4-dimethylpyrazole-phosphate.

Methane fluxes were typically small and mostly negative in all experiments, especially in dry soils. Cumulative  $CH_4$  uptake ranged from 242 to 435 g  $CH_4$ - $Cha^{-1}$  year<sup>-1</sup>, with no effect of N fertiliser treatment. Considered in terms of  $CO_2$  equivalents, soil  $CH_4$  uptake offset 8–56% of soil N<sub>2</sub>O emissions, with larger offsets occurring in non-N-fertilised soils.

The first few months from N fertiliser application to the period of rapid crop N uptake pose the main risk for  $N_2O$  losses from rainfed cereal cropping on subtropical Vertosols, but the realisation of this risk is dependent on rainfall. Strategies that reduce the soil mineral N pool during this time can reduce the risk of  $N_2O$  loss.

Additional keywords: 3,4-dimethylpyrazole-phosphate (DMPP), ENTEC, Hordeum vulgare, Triticum aestivum.

Received 17 November 2015, accepted 9 March 2016, published online 21 June 2016

#### Introduction

Direct greenhouse gas (GHG) emissions from Australia's agricultural sector total 81 Mt carbon dioxide equivalents (CO<sub>2</sub>-e) annually, or 15% of net national GHG emissions of 547 Mt CO<sub>2</sub>-e (Commonwealth of Australia 2015*b*). The agriculture sector is the primary source of emissions of both methane (CH<sub>4</sub>), essentially associated with meat and wool production, and nitrous oxide (N<sub>2</sub>O), predominantly associated with grain cropping (Dalal *et al.* 2003). Methane emissions from enteric fermentation (i.e. grazing animals) account for ~70% of all agricultural emissions, and N<sub>2</sub>O a further 15–20%. Concerns over the impacts of increasing GHG concentrations in the atmosphere and resultant effects on global warming and climate

change have prompted the Australian agricultural sector to sponsor research into methodologies for improved GHG mitigation. These methodologies will then be incorporated into industry plans for environmentally sustainable production that maintains or increases farm profitability and market access (Rodriguez *et al.* 2003; Gunasekera *et al.* 2007; Blumenthal *et al.* 2008).

The concentration of  $N_2O$  in the atmosphere is increasing, largely because of increased anthropogenic N inputs into the soil. Emissions of  $N_2O$  from the soil originate from the biological processes of nitrification and denitrification, which utilise inorganic N from any source (Bremner 1997). Susceptibility to  $N_2O$  loss is primarily determined by the The Australian Government publishes quarterly national inventories of the sources and magnitude of GHG emissions (Commonwealth of Australia 2015*b*) as well as comprehensive guidelines on how the emissions are calculated (Commonwealth of Australia 2015*a*). For the inventories, emissions of N<sub>2</sub>O associated with grain cropping are calculated by using an emissions factor (EF), which is the proportion of N input subsequently emitted to the atmosphere as N<sub>2</sub>O, applied to estimated amounts of N fertiliser and other N inputs (IPCC 2006). Australia uses a country-specific EF of 0.2% for N fertiliser used in rainfed cropping (Commonwealth of Australia 2015*a*), a value based on an analysis by Shcherbak and Grace (2014) of recent Australian field research.

Those same Australian studies indicate that the percentage of fertiliser N applied to grain crops and emitted as N<sub>2</sub>O varies substantially. The array of values reflects, among other factors, rate of fertiliser N, crop type, tillage practice, stubble management, soil texture, soil pH, and environment – particularly rainfall and temperature (Barker-Reid *et al.* 2005; Barton *et al.* 2008, 2010, 2013; Li *et al.* 2008; Officer *et al.* 2010; Wang *et al.* 2011; Schwenke *et al.* 2015). An analysis of Australian work in which conservation tillage was used indicates an overall average EF of 0.28% from a range 0.01–0.77%. The EFs for subtropical dryland cropping in northern New South Wales (NSW) and southern Queensland (average 0.48%) were higher than for the remainder of Australian grain-growing areas (average 0.07%).

Li et al. (2008) suggested that it was inappropriate to use a single default EF to estimate N2O emissions associated with grain cropping, with crop type, management and environmental factors needing to be taken into account in the calculations. Nitrous oxide is emitted episodically, largely in association with rainfall events. For example, there were 8-10 spikes of N<sub>2</sub>O emissions during the 3 years of field monitoring in the study by Wang et al. (2011), with the spikes much greater in magnitude for N-fertilised treatments than for unfertilised treatments. Those authors concluded that emissions mainly occurred when the water-filled pore space (WFPS) was >50% and the daily air temperature was >10°C. Other studies have also reported temporal patterns of N<sub>2</sub>O emissions (Barton et al. 2008, 2011; Officer et al. 2010; Schwenke et al. 2015). Therefore, the occurrence of rainfall episodes coinciding with other conditions conducive to high N2O emissions will determine the EF for any particular situation.

From an industry point of view, an emissions intensity index (EI) can be a more effective tool than EF for comparing management options to optimise both crop yield and  $N_2O$ emissions (Scheer *et al.* 2012). The EI is calculated as the ratio of  $N_2O$  emitted to grain produced. Simply aiming to minimise  $N_2O$  emissions with no regard to continued profitable grain production will produce strategies that are unlikely to be adopted by grain growers. Instead, using a yield-scaled emissions approach such as EI should ensure that proposed  $N_2O$ -mitigation strategies are both agronomically efficient and economically profitable (Van Groenigen *et al.* 2010). Substantially more data from direct field measurements of  $N_2O$  emissions are needed so that EFs and EIs can be applied that are at least more specific to region, crop type and soil (Ogle *et al.* 2014).

Apart from soils used for wetland rice cultivation. agricultural soils are typically considered a sink, not a source, of atmospheric CH<sub>4</sub>, but limited availability of data means that CH<sub>4</sub> uptake by soil microorganisms is currently not considered in Australian GHG inventory calculations (Dalal et al. 2008). Methanotrophic activity tends to be greater in forest soils than grasslands, with dryland arable soils typically having much lower uptake potential (Le Mer and Roger 2001). Conditions favouring greater CH<sub>4</sub> uptake by cultivated soils include low WFPS, temperatures >10°C, greater aeration, more gas-diffusion pathways (less compaction), non-acidic pH and low salinity (Dalal et al. 2008). Fine-textured soils typically have low CH<sub>4</sub> oxidation rates due to their low porosity and high water retention, which combine to restrict diffusion of CH<sub>4</sub> into the soil profile (Dutaur and Verchot 2007). However, long-term notill management that improves soil structure can enhance gas diffusivity and CH<sub>4</sub> uptake (Plaza-Bonilla et al. 2014). The addition of ammonium to soils through fertilisers may temporarily inhibit CH<sub>4</sub> uptake through competition between some methanotrophic groups and ammonium oxidisers for oxygen in the soil atmosphere. Long-term N-fertiliser use can also cause long-lasting reductions in methanotrophic activity (Hütsch et al. 1993). Chemical additives used to inhibit soil nitrification may either enhance (Weiske et al. 2001) or inhibit (Syamsul Arif et al. 1996) soil CH<sub>4</sub> uptake.

Globally, CH<sub>4</sub> uptake in cultivated soils averages  $0.92 \text{ kg CH}_4$ -C ha<sup>-1</sup> year<sup>-1</sup> across a range of boreal, temperate and tropical climates (Dutaur and Verchot 2007). Soil CH<sub>4</sub> flux data from cultivated Australian soils are available for tropical sugarcane soils in Queensland (Weier 1999; Denmead *et al.* 2010), temperate winter crops on a sandy soil in Western Australia (Barton *et al.* 2010, 2013), and subtropical winter wheat on a clay soil (Wang *et al.* 2011). There is a need for substantially more field-based data on CH<sub>4</sub> fluxes from a diversity of soil types, climatic conditions and potentially influential management factors. Such data would be used in national GHG inventories and, where CH<sub>4</sub> uptake exceeds CH<sub>4</sub> emission, might be used to offset some of the N<sub>2</sub>O emissions produced during the same cropping period (Wang *et al.* 2011).

We report data from five experiments conducted during a 2-year period in northern NSW, in which we used both automated and manual chambers to monitor fluxes of  $N_2O$  and  $CH_4$  associated with the production of N-fertilised wheat and barley. The automated chamber system provides high temporal-measurement resolution of GHG fluxes that can be readily related to changes in soil factors, particularly temperature and moisture. Chamber numbers, however, are restricted (in our case to 12), in turn restricting the number of treatments and/or replicates that can be monitored at any one time. On the other hand, manual chambers deployed at a particular site or experiment need not be as restricted in number and may therefore prove useful in assessing potential emissions-reducing management options.

#### Materials and methods

Five field-plot experiments were conducted in the winter cropping seasons of 2010 and 2011. Expts 1, 2 and 3 were at the Tamworth Agricultural Institute (31.152°S, 150.982°E) near Tamworth, NSW. Expt 4 (31.484°S, 150.177°E) and Expt 5 (31.171°S, 150.806°E) were on nearby commercial farms. All experiments were on neutral–alkaline Vertosols (Isbell 2002) of medium–heavy clay content.

## Experiments 1 and 3: automated chambers in wheat (2010) and barley (2011)

The site for Expts 1 and 3 was described by Schwenke *et al.* (2015), along with field methods of soil and plant sampling, laboratory analysis, monitoring of soil-water content and environmental conditions, and data interpretation.

Plots were 12 m long and 6 m wide. The experimental crops were wheat (Triticum aestivum cv. Crusader) in 2010 and barley (Hordeum vulgare cv. Shepherd) in 2011. All plots were sown with a no-till planter at a row spacing of 0.25 m. Wheat was sown on 27 July 2010. The previous crop (2009) was chickpea (Cicer arietinum). Wheat plots were machineharvested on 15 December 2010. Following the weed-free summer-autumn fallow, barley was sown on 27 June 2011 and it was harvested on 16 December 2011. Rates of applied N fertiliser were 0 or  $80 \text{ kg N} \text{ ha}^{-1}$  (wheat) and 0 or  $60 \text{ kg N} \text{ ha}^{-1}$ (barley). Rates were calculated as the projected crop N demand minus the pre-sowing soil mineral N supply as determined by soil-core testing to 1.5 m depth (Herridge 2011). Urea-N was side-banded at sowing at a 0.1-m offset to each plant row and a soil depth of 0.05-0.1 m. Weeds, insect pests and plant diseases were controlled chemically by using appropriate compounds at approved rates when necessary. The 2010 wheat and 2011 barley were grown within a larger crop rotation trial organised as a randomised complete block design with four replicates.

Fluxes of  $N_2O$  and  $CH_4$  were measured in three of the four replicates. One chamber (0.5 m by 0.5 m by 0.15 m high) was deployed in each treatment of three replicate plots from July 2010 to June 2011 for wheat, and from June 2011 to July 2012 for barley. Chambers were secured to bases pushed 0.1 m into the soil. Each base covered two crop rows and two N-fertiliser bands. During crop growth, chambers were moved weekly to new locations within each plot to minimise the impact of the chamber on plant growth. Chamber height was increased with extensions so that the chamber covered the crops as they grew, to include  $N_2O$  emitted through plant transpiration (Chang *et al.* 1998). At maturity, plants growing within the chamber bases were hand-harvested, the grain was removed, and the plant residues were returned to the same base.

We used a 12-chamber automated gas-measuring system (Scheer *et al.* 2011) on four treatments within the larger rotation trial. Data from only two of the four treatments are presented in this paper. The automatically operated chambers in the first replicate were closed for 60 min, during which time four separate samples of air from the chamber headspace were collected at 15-min intervals. Samples were analysed immediately in an *in-situ* gas chromatograph (8610C; SRI Instruments, Torrance, CA, USA) fitted with an electron-capture detector for N<sub>2</sub>O measurement and a flame-ionisation

detector for  $CH_4$  measurement. After the closure period, the chambers in the first replicate opened and those in the second replicate closed for 60 min, followed by the chambers in the third replicate 60 min later. After 180 min, the cycle restarted, giving eight measurements per chamber per day.

The N<sub>2</sub>O and CH<sub>4</sub> concentrations in the four air samples from each chamber during each closure period were regressed against closure time. For N2O fluxes, we used the HMR routine, developed by Pedersen et al. (2010), which selects the most appropriate model for flux estimation based on the data from each measurement period. The routine first fitted a non-linear model to the data. Where this fit was not statistically significant, the linear model was then fitted. If neither model was statistically significant, a slope of zero was assigned. For CH<sub>4</sub> fluxes, we used linear regression, with a slope of zero assigned where  $r^2$ <0.8. The slope of the selected regression was used in the calculation of N<sub>2</sub>O and CH<sub>4</sub> fluxes (Schwenke and Haigh 2016). Chamber temperature was measured by using a thermocouple probe in one chamber in each replicate set of four. Excessive heat (>55°C) within a chamber triggered a cessation of the current measurement and the chambers opened. Similarly, chambers opened and measurements were suspended when rainfall intensity exceeded 0.4 mm in 5 min. Altogether, ~25% of all possible measurement opportunities (eight per day) in Expts 1 and 3 were missed through heat, rainfall, equipment removal for planting or harvesting, or equipment servicing or breakdown. Cumulative N<sub>2</sub>O and CH<sub>4</sub> emitted during the experiment were calculated by linear interpolation between daily average fluxes.

Aboveground plant and grain samples (two 0.5-m rows per plot) were collected before harvest by hand-cutting, then dried at 70°C for 48 h, weighed and analysed for total N concentration (%N) by combustion analysis (EA1112; Thermo Finnigan, San Jose, CA, USA). Grain protein was calculated by multiplying grain %N by a factor of 5.7. All results for grain yield and protein were standardised to a moisture content of 12% before treatments were compared. After harvest, all plant residue material from a plot was distributed evenly across that same plot.

Surface and subsurface samples (0-0.1 and 0.1-0.2 m) for soil mineral N (ammonium and nitrate) were collected in a crossrow transect of five 0.05-m-diameter cores per plot. For deeper samples (0.2–0.3, 0.3–0.6, 0.6–0.9, 0.9–1.2 and 1.2–1.5 m), two cores were collected per plot. Surface and subsurface samples were collected approximately monthly and deeper cores at presowing and post-harvest. Soil bulk density was determined by the small-core volumetric-ring method (Cresswell and Hamilton 2002). Soil-water content was determined by gravimetric analysis at each time of soil sampling. Ammonium-N and nitrate-N in filtered (Whatman 42) soil extracts (2 M KCl) that were prepared on the day of sampling were determined by standard colourimetric analyses with a flow injection analyser (Lachat Instruments, Loveland, CO, USA). Soil moisture was also monitored by using a site-calibrated ThetaProbe ML2x soil moisture sensor (Delta-T Devices, Burwell, UK) at 0.05 m depth in the soil within each chamber.

Weather conditions measured at the site included ambient air temperature and relative humidity (Tinytag Plus 2: TGP-4500; Gemini Data Loggers, Chichester, UK), daily rainfall (RainLogger; RainWise, Trenton, ME, USA), and soil temperature at 0.05 and 0.15 m depths (Tinytalk II; Gemini Data Loggers).

# Experiment 2: manual chambers in wheat (Tamworth, 2010)

Experiment 2 was adjacent to Expts 1 and 3. Plots were 12 m long and 2 m wide. Wheat (cv. Crusader) was sown on 27 July 2010. The experiment was part of a larger, randomised complete block experiment with four replicates. Results presented here are for 4-m-wide plots that grew chickpea in 2009 and were then divided in half lengthwise, with one half receiving N fertiliser in 2010 and the other half no N fertiliser. Emissions of N<sub>2</sub>O and CH<sub>4</sub> were measured weekly on average (range 1–16 days between samplings) during the first 2 months, then every 12 days on average for the remainder of the year. Crop agronomy, and soil and plant sampling, were conducted as described for Expt 1. Manual measurements of GHG emission were described previously (Schwenke et al. 2015). The chambers were the same dimensions as the automated chambers used in Expt 1 and included plants. Chamber bases were inserted 0.1 m into the soil and remained in place throughout the experiment but chambers were removed after sampling. Upon chamber closure, 18-mL air samples were collected in pre-evacuated 12=mL glass Exetainer vials (Labco, UK) after 0, 30 and 60 min. All samples were later analysed using a laboratory gas chromatograph with detectors similar to those used in Expt 1.

# Experiment 4: manual chambers in wheat (Spring Ridge, 2011)

Wheat (cv. EGA Gregory) was sown on a Black Vertosol on 21 June 2011 and harvested on 5 December 2011. Plots were 9 m long and 1.5 m wide (5 rows at 0.3-m spacing). Nitrogen fertiliser treatments were: 0, 40, 80 and 160 kg N ha<sup>-1</sup> applied as urea side-banded at sowing; a split urea application (40 kg N ha<sup>-1</sup> at sowing+40 kg N ha<sup>-1</sup> as broadcast urea at Z31, mid-tillering; Zadoks *et al.* 1974) and 80 kg N ha<sup>-1</sup> of DMPP (3,4-dimethylpyrazole-phosphate) coated urea (ENTEC<sup>TM</sup>, Incitec-Pivot Fertilisers, Melbourne, Australia) at sowing. All treatments were replicated four times within a randomised block design.

Fluxes of N<sub>2</sub>O and CH<sub>4</sub> were measured by using manual chambers on 14 occasions during crop growth. Chambers were 0.25-m-diameter PVC cylinders positioned centrally over the fertiliser band, then pushed into the ground to a depth of 0.1 m, leaving 0.2 m above the soil surface. At the time of sampling, a lid fitted with a rubber O-ring was placed on the top of each chamber. Several chambers were sampled immediately after fitting the lids to give an average ambient gas concentration that was used for that day's flux calculations. Using preevacuated, 12-mL glass Exetainer (Labco, Lampeter, UK) vials, we collected 20 mL chamber air through a rubber septum in the chamber lid at 0, 30 and 60 min post-closure. These samples were analysed as for Expt 2. Cumulative N<sub>2</sub>O emitted during the experiment was calculated by multiplying the average daily flux rate of consecutive samples by the number of days between those consecutive sample collections. However, where consecutive samples were collected at more

than 3 days apart, the intervening daily fluxes were calculated by halving each previous day's flux. High-temporal resolution data from a previous auto chamber experiment at the site showed that this calculation avoided overestimating fluxes during extended dry periods after rainfall events and dry periods leading up to a wet period with high fluxes (Schwenke *et al.* 2015). Cumulative fluxes of  $CH_4$  were calculated by linear interpolation between sampling days.

Daily rainfall was measured at the site using a tippingbucket rain logger (RainWise). Soil was sampled to 1.5 m depth at eight locations across the trial area before sowing, then approximately monthly to 0.1 m in each plot during the crop's growth. Samples were processed and analysed for mineral N as in Expt 1. Biomass was measured at plant maturity (Z99) by collecting all aboveground plant matter in two 0.5-m lengths of row per plot. Cut samples were dried at 70°C for 48 h then weighed and analysed for %N as per Expts 1 and 3.

## Experiment 5: manual chambers in barley (Bithramere, 2011)

Barley (cv. Shepherd) was sown on a Red Vertosol on 9 June 2011 and harvested on 30 November 2011. Plots were 9 m long and 1.5 m wide (five rows at 0.3-m spacing). Nitrogen fertiliser treatments were 0, 30, 60 and 90 kg N ha<sup>-1</sup> applied as urea side-banded at sowing, a split urea application treatment  $(30 \text{ kg N ha}^{-1} \text{ at sowing} + 30 \text{ kg N ha}^{-1} \text{ as broadcast urea at Z31, mid tillering}), and 60 kg N ha<sup>-1</sup> of DMPP-coated urea at sowing. All treatments were replicated four times within a randomised block design.$ 

Fluxes of  $N_2O$  and  $CH_4$  were measured on 13 separate occasions during the crop growth as in Expt 4. Plant and soil samples were also collected and analysed as in Expt 4.

#### N<sub>2</sub>O emission factors and emission intensity

Emission factors for N<sub>2</sub>O were calculated by first subtracting the N<sub>2</sub>O emitted from the non-N-fertilised crop from that emitted by the N-fertilised crop, then dividing the net amount by the fertiliser N applied. In Expts 1–3, this was calculated for the crop-only period (EF<sub>crop</sub>, sowing–harvest) and for the full year after N fertiliser application (EF<sub>year</sub>). In Expts 4 and 5, EF was calculated for the crop period only. The N<sub>2</sub>O EI (gN<sub>2</sub>O-N t<sup>-1</sup> grain) was calculated as the cumulative N<sub>2</sub>O emitted over the measurement period, divided by the grain yield from the same plot.

#### Non-CO<sub>2</sub> GHG balance

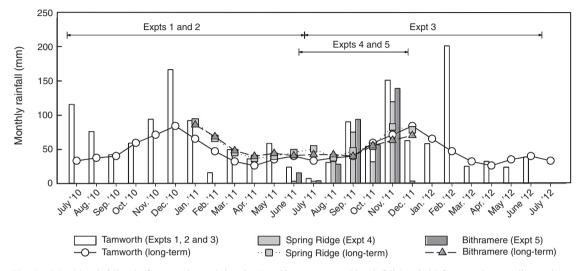
Using the cumulative  $N_2O$  and  $CH_4$  emission totals, an annual non- $CO_2$  GHG balance was calculated for each treatment in each trial as follows:

Non-CO<sub>2</sub> GHG balance (kg CO<sub>2</sub>-e ha<sup>-1</sup>year<sup>-1</sup>)  
= 
$$(298 \times \text{kg N}_2\text{O-N ha}^{-1}\text{year}^{-1} \times 44/28)$$
  
+  $(25 \times \text{kg CH}_4\text{-C ha}^{-1}\text{year}^{-1} \times 16/12)$ 

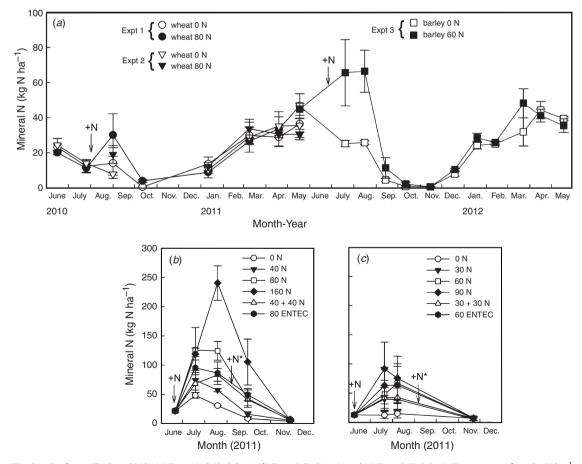
The conversion factors of 298 and 25 were used to convert  $N_2O$  and  $CH_4$ , respectively, to  $CO_2$ -e (Forster *et al.* 2007).

#### Statistical analyses

Statistical comparisons of treatment results were made using the REML with spatial analysis procedure in GENSTAT 16th edition (VSN International Ltd, Hemel Hempstead, UK). Individual treatment means were tested for difference using the least significant difference (l.s.d.) test at a probability level of P=0.05. Possible correlations between daily gas fluxes and environmental factors (air temperature, soil



**Fig. 1.** Monthly rainfall at the five experimental sites (bars) and long-term monthly rainfall (symbols) from nearby recording stations. Totals for Expts 4 and 5 include only rainfall after sowing and before harvesting, not the entire month as in Expts 1–3.



**Fig. 2.** Surface soil mineral N in (*a*) Expts 1-3 (0–0.2 m), (*b*) Expt 4 (0–0.1 m), and (*c*) Expt 5 (0–0.1 m). Treatments refer to kg N ha<sup>-1</sup> as standard urea applied at sowing or as a split application, or DMPP-coated urea (ENTEC) applied at sowing.

temperature at depths of 0.05 and 0.15 m, rainfall and WFPS) were investigated by using the multiple linear regression procedure in GENSTAT 16th edition.

#### Results

### Rainfall

Monthly rainfall deviated considerably from long-term averages at the experimental sites (Fig. 1). At the Tamworth site, rainfall in the weeks preceding sowing of Expts 1 and 2 was three times the long-term July average for the site (93rd percentile), and continued at a high intensity into August (86th percentile). Rainfall throughout the remainder of the growing season was near normal until harvest in December when the monthly total was double the long-term average (94th percentile). By contrast, there was minimal rainfall before, during and for 2 months after the sowing of Expts 3, 4 and 5 in June–July 2011 (33rd and 16th percentiles). Only in September and November 2011 were the monthly rainfalls well above average (86th and 91st percentiles).

Total in-crop (sowing-harvest) rainfall was 476, 476, 380, 303 and 338 mm for Expts 1–5, respectively. However, much of the above-average rainfall recorded in November 2011 occurred just before harvest and thus did not benefit crop production; for example, 86 mm rainfall occurred in the final week before harvest at Expt 5. During the post-harvest fallow to the end of the 12-month emissions-measuring period, Expts 1 and 2 had an additional 315 mm rainfall, and Expt 3 an additional 400 mm.

#### Soil mineral N

Pre-sowing mineral N concentrations (0-1.5 m) were uniform across the sites used for Expts 1-3 (data not shown). Postharvest sampling to 1.5 m depth showed no significant treatment effects on mineral N at any depth in the soil profile.

In all experiments, treatment effects of N fertiliser application on surface-soil mineral N (0-0.1 and 0-0.2 m)were evident for 1-3 months after application (Fig. 2), depending on the experimental conditions. In Expts 1 and 2 (Fig. 2a), soil mineral N in the surface declined between June and July 2010 as a result of the intense rainfall and waterlogging conditions between these sample times. Although sampling in late August 2010 showed significant treatment differences in soil mineral N, levels were much less than expected considering the amount of N applied; therefore, much of the applied N had been either lost or taken up by the crop. The next sampling in October found little remaining evidence of treatment effects. Following harvest in December 2010, soil mineral N increased in all treatments during the 6-month fallow period, with no effect of the N fertiliser application in July 2010.

Mineral N in the surface soil of Expt 3 (Fig. 2a) showed a large effect of fertiliser N throughout the dry period until September 2011, when rainfall and crop uptake reduced available N to nil by November 2011. Mineralisation of soil organic matter and crop residues rapidly replenished soil mineral N during the summer fallow, but no significant effects of the N fertiliser applied in June 2011 were detected post-harvest.

Surface soil mineral N in Expts 4 and 5 (Fig. 2*b*, *c*) showed effects of the fertiliser N applied at sowing throughout crop growth. However, treatment differences had disappeared by harvest. In Expt 4, the strategies of split N ( $40 + 40 \text{ kg N ha}^{-1}$ ) and DMPP-coated urea ( $80 \text{ kg N ha}^{-1}$ ) applications tended to reduce soil mineral N, particularly the ammonium component (not shown), in samples collected during July and August 2011 compared with the same rate of N added as urea at sowing ( $80 \text{ kg N ha}^{-1}$ ). These apparent trends were not statistically significant (P > 0.05). In September 2011, despite similar total mineral-N levels, the treatment with DMPP-coated urea had

Table 1. Crop N uptake, grain yield, grain protein and crop-residue N in Expts 1–3
Grain yield and protein data are presented from machine-harvest, and N-removal data from hand-harvests. For each parameter,
comparisons are between nil-N and N-applied treatments within experiments: $*P < 0.05$ ; n.s., no significant difference ( $P > 0.05$ )

	Expt	1 wheat	1	2 wheat rate $(kg N ha^{-1})$	Expt 3 barley	
	0	80	0	80	0	60
Crop N uptake $(kg N ha^{-1})$	136	147n.s	161	174n.s	75	106*
Grain yield $(t ha^{-1})$	3.1	3.0n.s.	3.2	3.0n.s.	3.7	3.8n.s. <sup>A</sup>
Grain protein (%)	13.5	14.1b*	13.7	13.9n.s.	10.6	13.2*
N removed in grain $(kg N ha^{-1})$	55	57n.s.	58	56n.s.	60	77*
Crop residue N $(kg N ha^{-1})$	54	66n.s.	53	74n.s.	16	29n.s.

<sup>A</sup>A significant yield difference was found in hand-harvest results, with  $0N=3.6 \text{ tha}^{-1}$  and  $60N=4.1 \text{ tha}^{-1}$ .

Table 2. Effects of rate of fertiliser N applied as urea on biomass, grain yield and grain protein of wheat in Expt 4Within rows, means followed by the same letter are not significantly different (P > 0.05)

	N fertiliser rate (kg N ha <sup><math>-1</math></sup> )							
	0	40	80	160	40+40 split	80 DMPP-coated		
Dry matter (Z99) (t ha <sup>-1</sup> )	11.2a	15.1b	15.4b	14.0b	15.3b	15.8b		
Grain yield $(t ha^{-1})$	4.4a	5.3b	5.6c	5.2b	5.2b	5.3b		
Grain protein (%)	10.5a	11.2b	11.9c	12.5d	12.0c	11.7c		

 
 Table 3. Effects of rate of fertiliser N applied as urea on biomass, grain yield and grain protein of barley in Expt 5

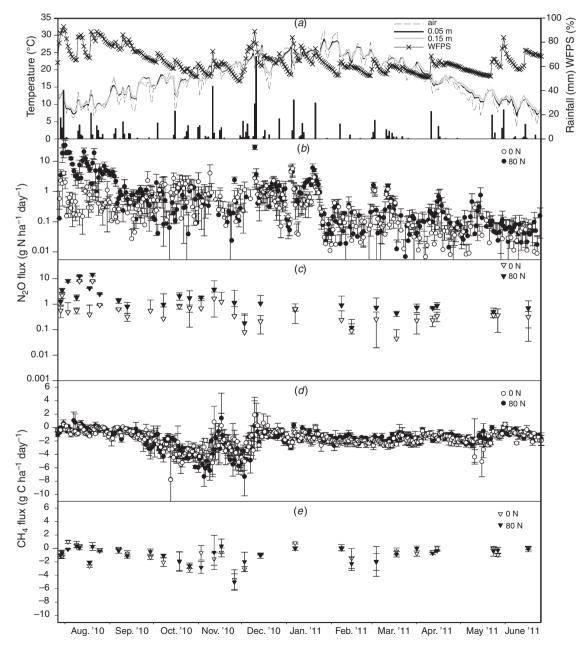
Within rows, means followed by the same letter are not significantly different (P > 0.05)

	0	30	60	90	30+30 split	60 DMPP- coated
Dry matter (Z99) (t ha <sup><math>-1</math></sup> )	6.0a	7.6b	8.9b	9.9c	9.9c	9.2c
Grain yield (tha <sup>-1</sup> ) Grain protein (%)		3.6ab 8.0 ab		4.9c 10.3c	4.8c 8.7bc	4.6c 10.1c

significantly more ammonium-N than the urea at sowing and the split N treatments, demonstrating its effectiveness in delaying nitrification. For Expt 5, there were no significant N treatment effects on soil mineral N at any sampling date.

#### Crop production, N uptake and grain yield

Nitrogen uptake by the wheat and barley in Expts 1 and 2 was not affected by N fertiliser, whereas in Expt 3, N fertiliser resulted in significantly higher crop N uptake (Table 1). Crop (aboveground) residue N was similarly unaffected by N fertiliser in Expts 1 and 2 but was higher with N application in Expt 3 (Table 1).



**Fig. 3.** Expts 1 and 2. Daily measurements of (*a*) rainfall, air and soil (0.05 and 0.15 m) temperatures, and water-filled pore space (WFPS); N<sub>2</sub>O flux in (*b*) Expt 1 and (*c*) Expt 2; and CH<sub>4</sub> flux in (*d*) Expt 1 and (*e*) Expt 2. Data in (*b*)–(*e*) are mean daily results ( $\pm$  standard error) of the three treatment replicates. Treatments refer to kg N ha<sup>-1</sup>.

In 2010, wheat grain yields were reduced because of late sowing caused by rain delays, waterlogging after sowing, a late frost coinciding with flowering, and rainfall during crop maturation (Table 1). These factors led to low harvest indices of 0.26 (Expt 1) and 0.22 (Expt 2). There was no significant effect of N fertiliser on grain yields in either experiment, but grain protein contents were high and, in Expt 1, were significantly increased with added N fertiliser.

Crop production in 2011 (Expt 3) was reduced by low available soil water at sowing and very dry conditions for the first 2 months post-sowing (Fig. 1). As a result, crop biomass was below that of 2010, although grain yields were similar across the two seasons (Table 1). The 2011 harvest index was 0.42. Fertiliser N addition in 2011 significantly increased yield in hand-harvested samples (although not in machine-harvested results), grain protein, and grain N removal (Table 1).

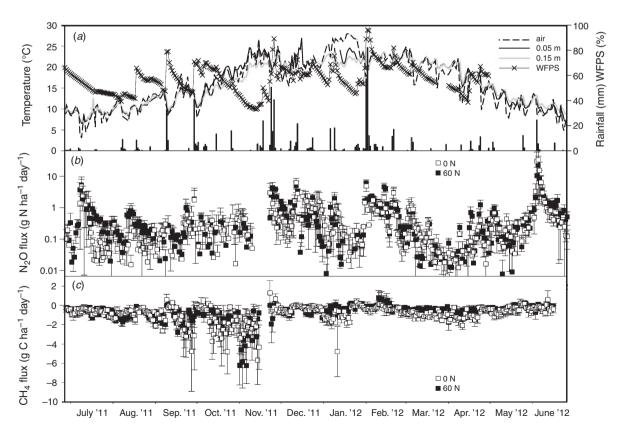
In Expts 4 and 5, mature crop biomass, grain yield and grain protein were increased by N fertiliser above the nil N treatment. In Expt 4, although all fertiliser N treatments produced similar biomass,  $80 \text{ kg N ha}^{-1}$  applied at sowing produced the highest grain yield, higher than the treatments with split N or the DMPP-coated urea with the same amount of N applied (Table 2). All three  $80 \text{ kg N ha}^{-1}$  strategies gave similar grain protein results, whereas the  $160 \text{ kg N ha}^{-1}$  rate significantly increased grain protein. The treatments with split N application and DMPP-coated urea did not influence grain yield or grain protein in Expt 5 but did result in greater

mature biomass than the equivalent N rate all applied at sowing (Table 3).

#### Daily N<sub>2</sub>O fluxes

In general, N<sub>2</sub>O emissions were very low, interspersed with brief periods of higher emission activity after heavy rain fell on saturated soils with either freshly decomposing crop residue or recently added N fertiliser. Daily N<sub>2</sub>O fluxes (gN<sub>2</sub>O-Nha<sup>-1</sup> day<sup>-1</sup>) averaged 1.0 in Expt 1 (range -1.7 to 33.5), 1.6 in Expt 2 (range -1.8 to 14.4), 0.6 in Expt 3 (range -1.0 to 33.0), 4.2 in Expt 4 (range -1.7 to 26.5), and 9.1 in Expt 5 (range -0.7 to 118.4).

Nitrous oxide fluxes in the first 2 months after N application were greater in the N-fertilised treatments of Expts 1 and 2 than in the corresponding nil-N treatments (Fig. 3*b*, *c*). Sowing and N fertiliser application on 27 July 2010 was followed by 68 mm rainfall over 3 days on an already-saturated soil, so soil WFPS remained >70% for much of the first 2 months post-sowing (Fig. 3*a*). From October 2010 onwards, the soil dried as the crop grew and N<sub>2</sub>O emissions remained low, with no difference in daily N<sub>2</sub>O flux between N-fertilised and non-fertilised treatments. In November and December 2010, isolated heavy rainfall events temporarily increased WFPS and N<sub>2</sub>O emission, but only for 1–2 days. Post-harvest N<sub>2</sub>O emissions were very low, with daily fluxes only temporarily exceeding 1 g N<sub>2</sub>O-N ha<sup>-1</sup> day<sup>-1</sup> following significant rainfall. Daily N<sub>2</sub>O emissions



**Fig. 4.** Expt 3. Daily measurements of (*a*) rainfall, air and soil (0.05 and 0.15 m) temperatures, and water-filled pore space (WFPS); (*b*) N<sub>2</sub>O flux (log scale); and (*c*) CH<sub>4</sub> flux. Data in (*b*) and (*c*) are mean daily results ( $\pm$  standard error) from three replicates of the two fertiliser-N treatments. Treatments refer to kg N ha<sup>-1</sup>.

measured in Expt 2 were generally similar to those in Expt 1 (Fig. 3c).

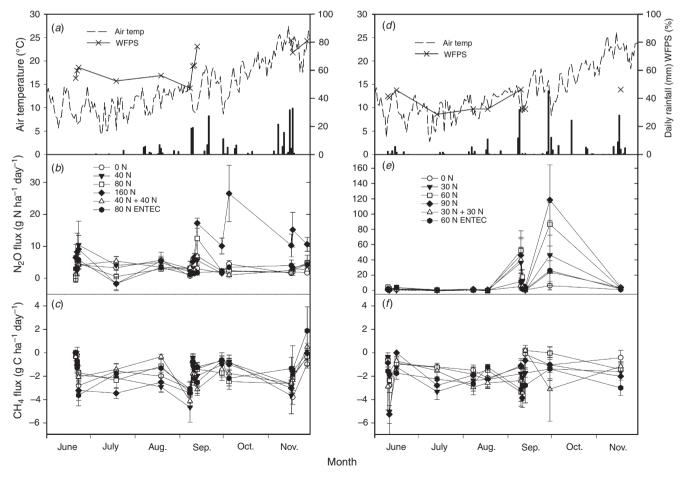
During the June–December 2011 crop period of Expt 3, there were only 7 days when soil WFPS exceeded 70% (Fig. 4*a*). It was not until February 2012 that more than 3 consecutive days showed soil WFPS >70%. As a result, daily N<sub>2</sub>O fluxes were low throughout the year of measurement, with only a single spike in daily N<sub>2</sub>O flux >10 g N<sub>2</sub>O-N ha<sup>-1</sup> day<sup>-1</sup> during June 2012 (Fig. 4*b*). There was no measurable impact of N fertiliser addition on N<sub>2</sub>O flux at any stage in Expt 3.

Daily N<sub>2</sub>O fluxes in Expts 4 and 5 were low for the first half of the growing season (Fig. 5*b*, *e*) owing to dry conditions, with elevated emissions recorded only after heavy rainfall (28 mm for Expt 4, 45 mm for experiment 5; Fig. 5*a*, *d*) in the later stages of the experimental period. In Expt 4, rain in early September 2011 resulted in treatment differences in N<sub>2</sub>O emissions. The 160 kg N ha<sup>-1</sup> treatment produced significantly higher N<sub>2</sub>O emissions than all other treatments for the remainder of the experiment. There were also two occasions in September when N<sub>2</sub>O flux was significantly greater in the 80 kg N ha<sup>-1</sup> treatment than the treatments with split N or DMPP-coated urea. In Expt 5, the large variation amongst replicates meant that there were no statistically significant differences between N rate treatments on most sampling occasions. However, there was a consistent trend across sampling dates for N<sub>2</sub>O fluxes in the order 90 kg N ha<sup>-1</sup> > 60 kg N ha<sup>-1</sup> > 30 kg N ha<sup>-1</sup> > split N, 30 + 30 N ha<sup>-1</sup> > DMPP-coated urea, 60 kg N ha<sup>-1</sup>.

#### Daily CH<sub>4</sub> fluxes

Emissions of CH<sub>4</sub> were below the detection limit or negative across all experiments; therefore, for the majority of measurements there was a net uptake of atmospheric CH<sub>4</sub>. Daily CH<sub>4</sub> fluxes (g CH<sub>4</sub>-C ha<sup>-1</sup> day<sup>-1</sup>) averaged -1.7 in Expt 1 (range -7.8 to 1.9), -0.9 in Expt 2 (range -5.0 to 1.0), -0.8 in Expt 3 (range -6.2 to 1.3), -1.6 in Expt 4 (range -4.7 to 1.9), and -1.9 in Expt 5 (range -5.3 to 0.2).

Fluxes of CH<sub>4</sub> in Expts 1, 2 and 3 became increasingly negative as the crop-growing season progressed and the soil became drier (Figs 3*d*, *e* and 4*c*). Significant rainfall events and high soil WFPS in November 2010 and September 2011 coincided with less negative or positive CH<sub>4</sub> fluxes. In both years of measurement, the CH<sub>4</sub> fluxes reverted to mostly negative again as the soil dried. Fluxes of CH<sub>4</sub> were more stable and generally less negative during the post-crop fallow than during the cropping period. Methane flux and WFPS were



**Fig. 5.** Expts 4 (left, a-c) and 5 (right, d-f). Daily measurements of (a, d) rainfall, air temperature, and water-filled pore space (WFPS);  $(b, e) N_2O$  flux; and  $(c, f) CH_4$  flux. Data in (b), (c), (e) and (f) are mean daily results ( $\pm$  standard error) of four replicates of six fertiliser-N treatments. Treatments refer to kg N ha<sup>-1</sup> as standard urea applied at sowing or as a split application, or DMPP-coated urea (ENTEC) applied at sowing.

significantly positively correlated, with WFPS accounting for 33% of variation in CH<sub>4</sub> fluxes in Expt 1 (P < 0.05) and 23% in Expt 3 (P < 0.05). Addition of N fertiliser did not significantly affect CH<sub>4</sub> flux in Expts 1–3.

In Expts 4 and 5 (Fig. 5*c*, *f*), CH<sub>4</sub> flux appeared to be positively related to soil moisture, but incomplete sampling of soil moisture during these experiments meant that statistical correlations could not be calculated. There was no significant effect of N fertiliser rate or application strategy on CH<sub>4</sub> flux, except during one mid-September sampling in Expt 5. In that case, the treatment with DMPP-coated urea at  $60 \text{ kg N ha}^{-1}$  had a significantly greater negative flux than treatments with  $60 \text{ kg N ha}^{-1}$  or split N at  $30 + 30 \text{ kg N ha}^{-1}$ , both of which had slightly positive fluxes on the same date.

# (Fig. 6*a*) but not in Expt 3 (Fig. 6*b*). Most of the treatment effect in Expts 1 and 2 occurred during the first 2–3 months after N fertiliser application, with only sporadic additions to the cumulative total throughout the remainder of the measurement period.

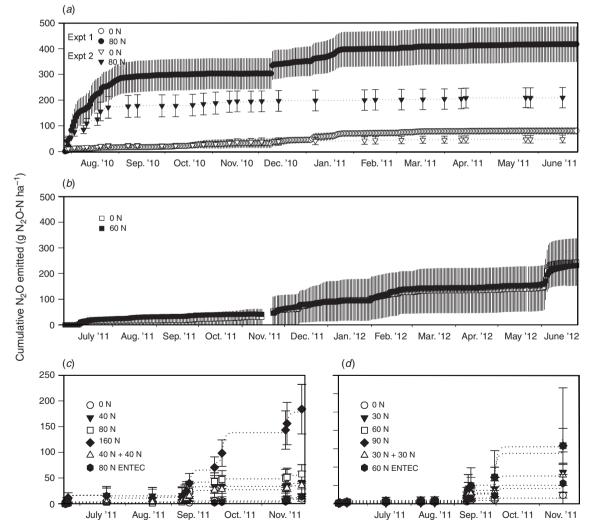
For Expts 4 and 5, there was a clear trend for increasing cumulative N<sub>2</sub>O emitted with increasing N rate (Figs 6*c*, *d* and 7*a*, *b*). DMPP-coated urea reduced N<sub>2</sub>O emissions by 76% (Expt 4) and 65% (Expt 5) compared with treatments where the same rate of urea was applied (Fig. 7*a*, *b*). Similarly, the strategy of splitting N reduced N<sub>2</sub>O emissions by 38% (Expt 4) and 51% (Expt 5) compared with treatments where the same rate of urea was applied all at sowing (Fig. 7*a*, *b*).

## *Cumulative CH*<sup>4</sup> uptake

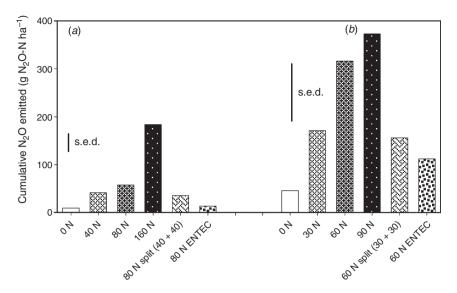
Cumulative  $N_2O$  emissions

Cumulative  $N_2O$  emissions for the year of measurement were significantly affected by N fertiliser application in Expts 1 and 2

Most daily  $CH_4$  fluxes were nil or negative, so the cumulative  $CH_4$  emitted during the measurement periods was negative; that is, there was a net uptake of atmospheric methane by the



**Fig. 6.** Cumulative N<sub>2</sub>O emitted in (*a*) Expts 1 and 2, (*b*) Expt 3, (*c*) Expt 4, and (*d*) Expt 5. Data are means ( $\pm$  standard error) of three (Expts 1–3) or four (Expts 4 and 5) replicates. Treatments refer to kg N ha<sup>-1</sup> as standard urea applied at sowing or as a split application, or DMPP-coated urea (ENTEC) applied at sowing.



**Fig. 7.** Cumulative N<sub>2</sub>O emitted in (*a*) Expt 4 and (*b*) Expt 5. Data are means of four replicates. P < 0.001 (Expt 4) and P = 0.11 (Expt 5). Standard error of difference (s.e.d.) shown for each experiment. Treatments refer to kg N ha<sup>-1</sup> as standard urea applied at sowing or as a split application, or DMPP-coated urea (ENTEC) applied at sowing.

soil in each experiment, with ranges of 242–435 g  $CH_4$ -C ha<sup>-1</sup> for Expts 1–3, and 181–366 g  $CH_4$ -C ha<sup>-1</sup> for Expts 4–5 (Fig. 8). There were no significant N fertiliser treatment effects on  $CH_4$  uptake in any experiment.

#### N<sub>2</sub>O emission factors

The similarity between  $EF_{crop}$  and  $EF_{year}$  in Expts 1 and 2 reflects the particular environmental conditions of these experiments, with N<sub>2</sub>O emissions primarily occurring during early crop growth (Table 4). In Expt 4, the EF for the treatment with DMPP-coated urea at  $80 \text{ kg N ha}^{-1}$  was significantly less than for the corresponding N rate of urea applied all at sowing  $(80 \text{ kg N ha}^{-1})$ , whereas the EF for the treatment with split N  $(40+40 \text{ kg N ha}^{-1})$  was statistically between the these two treatments.

The EI values reflected significant effects of N fertiliser applications in Expts 1 and 4, with more  $N_2O$  emitted per t grain when N fertiliser was applied in Expt 1 because of the seasonal conditions. However, in Expt 4, the inefficiency of the highest N rate was demonstrated by a higher EI than all other treatments (Table 4). The treatments with DMPP-coated urea in both Expts 4 and 5 had lower EIs than the equivalent N rates with ordinary urea, although these differences were not statistically significant.

#### Non-CO<sub>2</sub> GHG balance

The non-CO<sub>2</sub> GHG balance for the crop period varied from 10 to 150 kg CO<sub>2</sub>-e ha<sup>-1</sup> across Expts 1–3, and from 15 to 181 kg CO<sub>2</sub>-e ha<sup>-1</sup> for the full year (Table 5). The influence of CH<sub>4</sub> uptake in offsetting N<sub>2</sub>O emissions differed between experiments and treatments. In CO<sub>2</sub>-e terms, CH<sub>4</sub> uptake offset 8–11% of the N<sub>2</sub>O emitted from the 80 kg N ha<sup>-1</sup> treatment in Expts 1 and 2, and 39–56% of the N<sub>2</sub>O from the nil N treatment. In Expt 3, fertiliser N addition did not

significantly affect the 9% of  $N_2O$  emissions offset by the  $CH_4$  uptake.

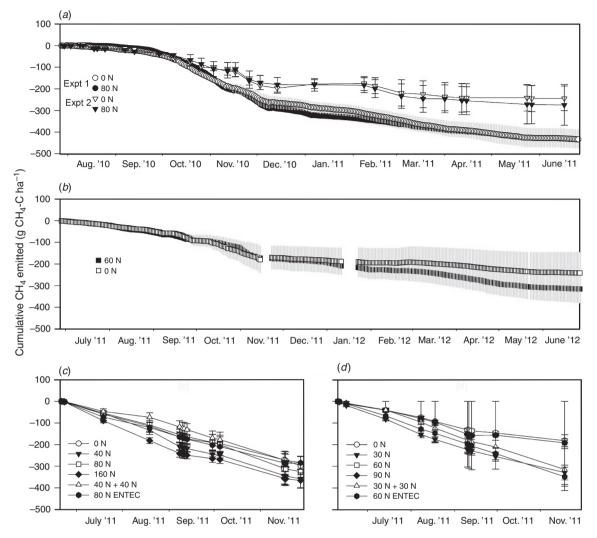
#### Discussion

#### N<sub>2</sub>O emissions in relation to rainfed grain production

Several studies have examined N<sub>2</sub>O emissions from N-fertilised dryland wheat on cracking clay soils in Australia, with total yearly N<sub>2</sub>O emissions ranging from  $0.50 \text{ kg N}_2\text{O-N ha}^{-1}$  in western Victoria (Officer *et al.* 2008) to  $0.90 \text{ kg N}_2\text{O-N ha}^{-1}$  in southern Queensland (Wang *et al.* 2011). By comparison, we measured  $0.42 \text{ kg N}_2\text{O-N ha}^{-1}$  for N-fertilised wheat and  $0.23 \text{ kg N}_2\text{O-N ha}^{-1}$  for N-fertilised wheat and  $0.23 \text{ kg N}_2\text{O-N ha}^{-1}$  for N-fertilised barley. Our EF values for wheat (0.42% in Expt 1, 0.20% in Expt 2, 0.06% in Expt 4) and barley (-0.02% in Expt 3, 0.30% in Expt 5) were considerably lower than the 3-year average EF of 0.77% for no-till, stubble-retained wheat measured by Wang *et al.* (2011). However, they were generally higher than the 0.14% for N-fertilised, non-irrigated wheat in the study by Officer *et al.* (2008).

The dissimilarity in cumulative  $N_2O$ -N totals between Expts 1 and 2 is likely the result of differences in sampling frequency (Barton *et al.* 2015), with manual sampling occurring weekly on average during the first 2 months, and then every 12 days on average for the remainder of the year. Large but transient daily  $N_2O$  emission peaks occurring between sampling days in the first 2 months may therefore have been missed. Additionally, there may have been days when the flux at the designated time of sampling was not representative of the daily average flux rate. The high-frequency data from Expt 1 are the more robust measurement.

None of the experiments reported here experienced average rainfall in the first few months after sowing and N fertiliser application (Fig. 1). In 2010 (Expts 1 and 2), rainfall for several days after sowing resulted in conditions of high soil WFPS, which produced significant  $N_2O$  emissions from the



**Fig. 8.** Cumulative CH<sub>4</sub> emitted in (*a*) Expts 1 and 2, (*b*) Expt 3, (*c*) Expt 4, and (*d*) Expt 5. Data are means ( $\pm$  standard error) of three (Expts 1–3) or four (Expts 4 and 5) replicates. Treatments refer to kg N ha<sup>-1</sup> as standard urea applied at sowing or as a split application, or DMPP-coated urea (ENTEC) applied at sowing.

N-fertilised treatment. Soil sampling conducted 1 month after sowing found less mineral N in the soil than would be expected from the amount added in fertiliser, indicating substantial N loss from the surface soil through denitrification and nitrate leaching. Because the N-fertilised soil did not affect postharvest N<sub>2</sub>O emissions, the critical period for N<sub>2</sub>O loss was the first 2–3 months after application of N fertiliser (Scheer *et al.* 2016).

By contrast, the N fertiliser applied at the 2011 sowing remained largely intact as soil mineral N for the next 2 months due to below-average moisture conditions (Fig. 1). Resumption of average, though not intense, rainfall during September led to rapid plant uptake of the soil mineral N (Fig. 2). Above-average rainfall in November temporarily increased N<sub>2</sub>O emissions but by then soil nitrate concentration was low (Fig. 2*a*), so N<sub>2</sub>O losses were small. As in 2010, there was minimal mineral N remaining post-harvest and no N-fertiliser treatment effect on crop residue N to cause any lasting treatment effects on postharvest N<sub>2</sub>O emissions. Therefore, the risk of early N<sub>2</sub>O loss after fertiliser N application was not realised in 2011 because of the dry conditions. Consequently, the measured EF was effectively nil.

Clearly, N<sub>2</sub>O emissions associated with fertiliser N applications for a particular site will very much depend on the seasonal rainfall conditions. Predicting emissions by using readily available information such as soil texture, rainfall and temperature profile will produce more accurate inventory results. In the meantime, the currently prescribed EF of 0.2% appears reasonable (Shcherbak and Grace 2014; Commonwealth of Australia 2015*a*).

#### N<sub>2</sub>O mitigation options for rainfed cereal cropping

Application of N fertiliser in excess of crop requirements can lead to exponentially increased  $N_2O$  emissions, as seen in Expts 4 and 5, and in many international studies (Scherbak *et al.* 2014). Even at optimum rates, emissions can still be substantial, so alternative strategies for N application need to be considered.

## Table 4. $N_2O$ emissions factor (EF, %) and $N_2O$ emissions intensity (EI, $g N_2O$ -N t<sup>-1</sup> grain) for each experiment

 $EF_{crop}$ , N<sub>2</sub>O emitted during crop growth divided by N added as urea × 100;  $EF_{year}$ , N<sub>2</sub>O emitted in the year since N addition divided by N added as urea × 100;  $EI_{crop}$ , N<sub>2</sub>O emitted during crop growth divided by grain produced. *P*-values for comparison between treatments within experiments and standard errors of difference (s.e.d.) are also presented

	N fertiliser (urea) $(kg N ha^{-1})$	EF <sub>crop</sub>	EFyear	EI <sub>crop</sub>
	Exp	ot 1		
2010-11	0	_	_	13
	80	0.38	0.42	115
Р				0.00
s.e.d.				13
	Exp	ot 2		
2010-11	0	_	_	12
2010 11	80	0.20	0.20	66
Р	00	0120	0120	0.08
s.e.d.				16
	Exp			
2011-12	0	4 5		21
2011-12	60	0.01	-0.02	20
Р	00	0.01	-0.02	0.88
s.e.d.				14
5. <b>c</b> . <b>u</b> .				14
	Exp	ot 4		
2011	0	_	_	2
	40	0.08	_	8
	80	0.06	_	10
	160	0.11	_	35
	40+40 split	0.03	—	7
D	80 DMPP-coated	0.01	—	3
P		0.20		0.01
s.e.d.		0.04		6.6
	Exp	ot 5		
2011	0	_	_	17
	30	0.42	_	53
	60	0.45	_	95
	90	0.36	_	98
	30+30 split	0.18	_	40
	60 DMPP-coated	0.11	_	29
Р		0.64		0.17
s.e.d.		0.28		36

Results from Expts 1 and 2 and from a study of N-fertilised canola at the same site (Schwenke *et al.* 2015) indicated that application of fertiliser N at sowing led to significantly higher N<sub>2</sub>O emissions for up to several months before crop growth depleted the pool of soil mineral N derived from fertiliser. Expts 4 and 5 explored two potential strategies to reduce this pool of soil mineral N. The strategies were (*i*) splitting the optimal N fertiliser 50 : 50 between application at sowing and topdressing at mid-tillering, and (*ii*) applying DMPP-coated urea at sowing at the optimal N fertiliser rate. These experiments coincided with abnormally dry conditions in 2011 where denitrification or leaching losses were minimal during the critical post-sowing period, so the full potential of these strategies could not be assessed within this study.

Cereal N uptake typically proceeds very slowly until tillering begins, and then increases rapidly to a maximum N

uptake rate during the jointing stage. Therefore, it is logical to delay some or all of the N fertiliser application until late tillering. Splitting the N application for cereal production has been found to increase grain yield, grain protein and fertiliser N recovery in many studies (IAEA 2000). Split N application is practised in many cereal-growing regions worldwide: however, it is rare in the study region. This is because <20% of the annual rainfall (600-800 mm) occurs in winter, winter rainfall events are highly variable with coefficients of variation of 46-80% (Webb et al. 1997), and forecast rain fronts during winter may not eventuate into sufficient rainfall for the crop to utilise the applied N (Dovle and Shapland 1991; Herridge 2011). Other studies of effects of split N application on N2O emissions have shown mixed results, depending on the timing of significant rainfall events in relation to N application (Weier 1999; Burton et al. 2008; Venterea and Coulter 2015), or the total N rate applied (Allen et al. 2008; Zebarth et al. 2008). Expts 4 and 5 in this study showed apparent reductions in emissions of 38-51% compared with a single N application at sowing, although grain yield was also reduced in Expt 4. Further field research is needed into the impacts of split N application on N<sub>2</sub>O emissions from rainfed cereal crops in Australia.

The application of DMPP-coated urea fertiliser (ENTEC) at sowing to delay nitrification has been reported in several cropping experiments (Weiske et al. 2001; Liu et al. 2013; De Antoni Migliorati et al. 2014; Scheer et al. 2014; Scheer et al. 2016). All studies showed that this treatment significantly reduced nitrate production from fertiliser and consequently reduced seasonal N2O emissions by 38-75% compared with standard urea, with deviations in effectiveness linked to soil moisture and temperature conditions (Chen et al. 2010; Menéndez et al. 2012). In our study, DMPP-coated urea reduced cumulative N<sub>2</sub>O emissions in Expts 4 and 5 by 65-75% compared with standard urea when both were applied at the same N rate at sowing. Our soil mineral N and N2O results also indicated good potential for these strategies to influence the outcome in a wetter season, but the 2011 season did not see waterlogging that would have caused potentially yield-reducing soil N loss through denitrification. Further field research is needed to quantify the impact of nitrification inhibition as a practical mitigation option in unusually wet cropping seasons.

#### CH<sub>4</sub> uptake associated with rainfed grain production

Methane uptake rather than emission dominated CH<sub>4</sub> fluxes in this study, with the rate of uptake negatively correlated with soil moisture; that is, dry soil absorbed more CH<sub>4</sub>. Daily CH<sub>4</sub> fluxes were within the ranges for CH<sub>4</sub> emission and uptake recorded for other Australian rainfed grain-cropping experiments (Barton *et al.* 2010, 2013; Wang *et al.* 2011). The annual cumulative CH<sub>4</sub>-C uptake in this study ranged from 242 to 435 g CH<sub>4</sub>-C ha<sup>-1</sup> year<sup>-1</sup> (Table 5), which was comparable to the 301–539 g CH<sub>4</sub>-C ha<sup>-1</sup> year<sup>-1</sup> uptake measured in a semi-arid sandy soil (Barton *et al.* 2013). However, other Australian grain-cropping studies have reported either no net CH<sub>4</sub> uptake (Barton *et al.* 2010), or net CH<sub>4</sub> emissions of 75–150 g CH<sub>4</sub>-C ha<sup>-1</sup> year<sup>-1</sup> (Wang *et al.* 2011).

There is evidence in the literature for the inhibition of soil  $CH_4$  uptake by applied N fertiliser (Dalal *et al.* 2008),

Table 5. Cumulative N<sub>2</sub>O, CH<sub>4</sub> and non-CO<sub>2</sub> greenhouse gas (GHG) balance for crop and full-year periods in Expts 1–3

Values are presented as N<sub>2</sub>O, CH<sub>4</sub> or CO<sub>2</sub> equivalents (CO<sub>2</sub>-e) per ha. *P*-values for comparison between treatments within experiments and standard errors of difference (s.e.d.) are also presented

N rate	Crop period						Crop period + post-crop fallow				
$(\text{kg}\text{ha}^{-1})$	Cumulative N <sub>2</sub> O Cumulati			ive CH <sub>4</sub> Non-CO <sub>2</sub> GHG		Cumulative N <sub>2</sub> O		Cumulative CH <sub>4</sub>		Non-CO2 GHG	
	$(g N_2O-N)$	(kg CO <sub>2</sub> -e)	(g CH <sub>4</sub> -C)	(kg CO <sub>2</sub> -e)	(kg CO <sub>2</sub> -e)	$(g N_2O-N)$	(kg CO <sub>2</sub> -e)	$(g CH_4-C)$	(kg CO <sub>2</sub> -e)	(kg CO <sub>2</sub> -e)	
					Expt 1						
0	39	18	-266	-9	10	80	38	-434	-15	23	
80	340	159	-292	-10	150	417	196	-435	-15	181	
Р	0.00	0.00	0.72	0.72	0.00	0.00	0.00	0.52	0.52	0.00	
s.e.d.	40	19	31	1.0	19	51	24	47	1.6	25	
					Expt 2						
0	39	18	-181	-6	12	48	23	-244	-8	15	
80	197	92	-197	-7	86	210	98	-275	-9	90	
Р	0.13	0.13	0.77	0.77	0.12	0.12	0.12	0.84	0.84	0.15	
s.e.d.	62	29	49	1.6	28	62	29	132	4.4	33	
					Expt 3						
0	73	34	-178	-6	28	247	115	-242	-8	107	
60	80	38	-184	-6	32	232	109	-316	-11	98	
Р	0.96	0.96	0.88	0.88	0.96	0.94	0.94	0.76	0.76	0.93	
s.e.d.	50	23	48	1.6	22	115	54	101	3.4	52	

but we found no such effects in our experiments. Likewise, Wang et al. (2011), with a similar soil type (Black Vertosol) and subtropical climate, also found no effect of long-term (38-year) N fertiliser addition on CH<sub>4</sub> fluxes during 3 years of continuous monitoring. In Expt 5, one sampling period indicated enhanced CH<sub>4</sub> uptake from soils fertilised with DMPP-coated urea compared with soils fertilised with standard urea at the same N rate. Weiske et al. (2001) found that DMPP-coated ammonium sulfate nitrate stimulated annual soil CH<sub>4</sub> uptake by an average of 28% during a 3-year study in Germany, with the treatment differences in 2 years lasting for several months after fertiliser application. Given the consistent and significant overall trend for soil CH4 uptake observed in this study, further field or laboratory studies should be targeted to isolate the effects of potentially influencing factors such as N fertiliser rate, weed management (herbicides, tillage operations), fertiliser type, and fertiliser inhibitors.

#### Net non-CO<sub>2</sub> GHG emissions

Rainfed grain-cropping systems have mostly been evaluated for their impacts on N<sub>2</sub>O emissions, with CH<sub>4</sub> and CO<sub>2</sub> fluxes not always measured or reported. Barton *et al.* (2013) found that cumulative CH<sub>4</sub> uptake increased with lime application in a wheat–wheat rotation, but they did not offset this against N<sub>2</sub>O emitted from the same treatments and did not report CO<sub>2</sub> fluxes. Cumulatively, the beneficial effects of CH<sub>4</sub> uptake by the soils in this study offset the environmentally harmful effects of the N<sub>2</sub>O emitted by 9–56%, with proportionally greater offsets occurring in the non-N fertilised plots that had lower N<sub>2</sub>O emissions. Wang *et al.* (2011) found no treatment effects on CH<sub>4</sub> flux and no clear trends towards either net CH<sub>4</sub> emission or consumption, but they did include the CH<sub>4</sub> flux data in a net GHG evaluation of their experimental treatments.

The net soil  $CO_2$  flux associated with cropping can also be considered when conducting a net GHG evaluation of soil–plant management options. However, the chambers used in this study necessarily included growing plants (to capture  $N_2O$  emitted in transpiration); therefore,  $CO_2$  flux measured in the chambers was a product of plant photosynthesis and plant–soil respiration. Although these data can be indicative of soil microbial activity (Barton *et al.* 2010), the inclusion of plant respiration precludes it from use in GHG balance calculations, as does the difficulty in accounting for the C added back into the soil as plant residues and exudates.

An alternative means of including CO<sub>2</sub> flux in a net GHG balance is to use an indirect estimate derived from long-term changes in soil organic C (SOC) stocks. For example, Wang et al. (2011) used 40 years of SOC data to calculate the net annual CO<sub>2</sub> fluxes from continuous wheat production on a subtropical Black Vertosol. They found a near-neutral response  $(9 \pm 196 \text{ kg CO}_2\text{-}e \text{ ha}^{-1} \text{ year}^{-1})$  in plots of continuous wheat under no-till, stubble-retained and N-fertilised conditions, compared with  $284 \pm 75 \text{ kg CO}_2$ -e ha<sup>-1</sup> year<sup>-1</sup> lost by the same system with no N fertiliser. Such a difference potentially outweighs the negative impacts of N2O emissions from Nfertiliser use when calculating the net GHG balance. We have not used CO<sub>2</sub> fluxes in our calculations because comparative CO<sub>2</sub> flux data do not exist for the no-till, stubble-retained, grainlegume-N-fertilised cereal cropping systems studied here. However, nearby long-term studies have found that SOC was stable over time in a cultivated, grain-legume-nil-N wheat rotation (Blair and Crocker 2000; Blair et al. 2006) after an initial decline (Holford et al. 1998), and potentially increasing in N-fertilised cereal-legume rotations with no-till and stubble retention (Young et al. 2005).

#### Conclusions

For a winter cereal crop, the critical period to target for strategic mitigation of  $N_2O$  emissions is between application of the N fertiliser at sowing and the period of rapid N uptake by the

growing crop after the onset of tillering. Although this risk period is present each year, actual N<sub>2</sub>O losses are realised only when rainfall intensity raises soil WFPS and denitrification activity increases. Possible management options to reduce N<sub>2</sub>O losses need further field investigation, with split applications of fertiliser N and urea coated with a nitrification inhibitor (DMPP) showing promise. The cereal crops studied here showed no post-harvest carry-over of the N applied at sowing; therefore, mineralisation of N-fertilised crop residues is unlikely to create further risk of N<sub>2</sub>O losses, as was the case with N-rich legumes and canola (Schwenke *et al.* 2015).

Methane uptake in this production system was largely unaffected by N-fertiliser use or application strategy, but was found to vary in response to soil WFPS. Although the uptake of CH<sub>4</sub> by soil benefits the overall GHG balance for a cropping paddock, its contribution relative to N<sub>2</sub>O emissions is generally small in an N-fertilised situation. However, where N<sub>2</sub>O emissions are low in drier cropping seasons, CH<sub>4</sub> uptake can almost completely offset the negative environmental effects of N<sub>2</sub>O emitted during a dryland cereal crop.

#### Acknowledgements

The authors gratefully acknowledge technical assistance from Kelly Leedham, Adam Perfrement, Jan Hosking, Rod Bambach, Zara Temple-Smith (NSW DPI), and Christian Brunk (QUT). This project was funded by NSW DPI and the GRDC, in conjunction with the Nitrous Oxide Research Program (NORP), which was funded by the Australian Department of Agriculture, Forests and Fisheries. We are indebted to Professor Peter Grace (QUT), the coordinator of NORP, and the other NORP network participants for their guidance and advice.

#### References

- Allen DE, Kingston G, Rennenberg H, Dalal RC, Schmidt S (2008) Nitrous oxide emissions from sugarcane soils as influenced by waterlogging and split N fertiliser application. In 'Proceedings 30th Annual Conference of the Australian Society of Sugar Cane Technologists'. Townsville, Qld. (Ed. RC Bruce) pp. 95–104. (Australian Society of Sugar Cane Technologists: Mackay, Qld)
- Barker-Reid F, Gates WP, Wilson KBR, Galbally IE, Meyer CP, Weeks IA, Eckard RJ (2005) Soil nitrous oxide emissions from rainfed wheat in SE Australia. In 'Science, control, policy and implementation. Proceedings Fourth International Symposium on Non-CO<sub>2</sub> Greenhouse Gases (NCGG-4)'. Utrecht, The Netherlands. (Ed. A van Amstel). (IOC Press: Amsterdam)
- Barton L, Kiese R, Gatter D, Butterbach-Bahl K, Buck R, Hinz C, Murphy DV (2008) Nitrous oxide emissions from a cropped soil in a semi-arid climate. *Global Change Biology. Bioenergy* 14, 177–192.
- Barton L, Murphy DV, Kiese R, Butterbach-Bahl K (2010) Soil nitrous oxide and methane fluxes are low from a bioenergy crop (canola) grown in a semi-arid climate. *Global Change Biology. Bioenergy* 2, 1–15. doi:10.1111/j.1757-1707.2010.01034.x
- Barton L, Butterbach-Bahl K, Kiese R, Murphy DV (2011) Nitrous oxide fluxes from a grain-legume crop (narrow-leafed lupin) grown in a semiarid climate. *Global Change Biology* 17, 1153–1166. doi:10.11 11/j.1365-2486.2010.02260.x
- Barton L, Murphy DV, Butterbach-Bahl K (2013) Influence of crop rotation and liming on greenhouse gas emissions from a semi-arid soil. *Agriculture, Ecosystems & Environment* 167, 23–32. doi:10.1016/j.agee. 2013.01.003
- Barton L, Wolf B, Rowlings D, Scheer C, Kiese R, Grace P, Stefanova K, Butterbach-Bahl K (2015) Sampling frequency affects estimates of

annual nitrous oxide fluxes. Scientific Reports 5, 15912. doi:10.1038/ srep15912

- Blair N, Crocker GJ (2000) Crop rotation effects on soil carbon and physical fertility of two Australian soils. *Australian Journal of Soil Research* 38, 71–84. doi:10.1071/SR99064
- Blair N, Faulkner RD, Till AR, Crocker GJ (2006) Long-term management impacts on soil C, N and physical fertility: Part III: Tamworth crop rotation experiment. *Soil & Tillage Research* **91**, 48–56. doi:10.1016/ j.still.2005.11.003
- Blumenthal M, Umbers A, Day P (2008) 'A responsible lead: an environmental plan for the Australian grains industry.' (Grains Research and Development Corporation: Canberra, ACT). Available at http://www.grdc.com.au/uploads/documents/GRDC\_Environmental\_ Plan.pdf [verified 28 May 2016]
- Bremner JM (1997) Sources of nitrous oxide in soils. Nutrient Cycling in Agroecosystems 49, 7–16. doi:10.1023/A:1009798022569
- Burton DL, Zebarth BJ, Gillam KM, MacLeod JA (2008) Effect of split application of fertilizer nitrogen on N<sub>2</sub>O emissions from potatoes. *Canadian Journal of Soil Science* 88, 229–239. doi:10. 4141/CJSS06007
- Chang C, Janzen HH, Cho CM, Nakonechny EM (1998) Nitrous oxide emission through plants. *Soil Science Society of America Journal* 62, 35–38. doi:10.2136/sssaj1998.03615995006200010005x
- Chen D, Suter HC, Islam A, Edis R (2010) Influence of nitrification inhibitors on nitrification and nitrous oxide (N2O) emission from a clay loam soil fertilized with urea. *Soil Biology & Biochemistry* 42, 660–664. doi:10.1016/j.soilbio.2009.12.014
- Commonwealth of Australia (2015*a*) 'National Inventory Report 2013. Vol. 1.' (Commonwealth of Australia: Canberra, ACT)
- Commonwealth of Australia (2015*b*) 'Quarterly update of Australia's National Greenhouse Gas Inventory: September 2014.' (Commonwealth of Australia: Canberra, ACT)
- Cresswell HP, Hamilton GJ (2002) Bulk density and pore space relations. In 'Soil physical measurement and interpretation for land evaluation'. (Eds N McKenzie, K Coughlan, H Cresswell) pp. 35–58. (CSIRO Publishing: Melbourne)
- Dalal RC, Wang WJ, Robertson GP, Parton WJ (2003) Nitrous oxide emission from Australian agricultural lands and mitigation options: a review. Australian Journal of Soil Research 41, 165–195. doi:10.1071/ SR02064
- Dalal RC, Allen DE, Livesley SJ, Richards G (2008) Magnitude and biophysical regulators of methane emission and consumption in the Australian agricultural, forest, and submerged landscapes: a review. *Plant and Soil* 309, 43–76. doi:10.1007/s11104-007-9446-7
- De Antoni Migliorati M, Scheer C, Grace PR, Rowlings DW, Bell M, McGree J (2014) Influence of different nitrogen rates and DMPP nitrification inhibitor on annual N2O emissions from a subtropical wheat-maize cropping system. *Agriculture, Ecosystems & Environment* 186, 33–43. doi:10.1016/j.agee.2014.01.016
- Denmead OT, Macdonald BCT, Bryant G, Naylor T, Wilson S, Griffith DWT, Wang WJ, Salter B, White I, Moody PW (2010) Emissions of methane and nitrous oxide from Australian sugarcane soils. *Agricultural* and Forest Meteorology 150, 748–756. doi:10.1016/j.agrformet.2009. 06.018
- Doyle A, Shapland R (1991) Effect of split nitrogen applications on the yield and protein content of dryland wheat in northern New South Wales. Australian Journal of Experimental Agriculture 31, 85–92. doi:10.1071/EA9910085
- Dutaur L, Verchot LV (2007) A global inventory of the soil CH<sub>4</sub> sink. Global Biogeochemical Cycles 21, GB4013.
- Forster P, Ramaswamy V, Artaxo P, Berntsen T, Betts R, Fahey DW, Haywood J, Lean J, Lowe DC, Myhre G, Nganga J, Prinn R, Raga G, Schulz M, Van Dorland R (2007) Changes in atmospheric constituents and in radiative forcing. In 'Climate change 2007: The physical science

basis. Contribution of Working Group I to the Fourth Assessment Report of the Intergovernmental Panel on Climate Change'. (Eds S Solomon, D Qin, M Manning, Z Chen, M Marquis, KB Averyt, M Tignor, HL Miller). (Cambridge University Press: Cambridge, UK, and New York)

- Gunasekera D, Kim Y, Tulloh C, Ford M (2007) Climate change impacts on Australian agriculture. *Australian Commodities: Forecasts and Issues* 14, 657–676.
- Herridge DF (2011) 'Managing legume and fertiliser N for northern grains cropping.' (Grains Research and Development Corporation: Canberra, ACT)
- Holford ICR, Schweitzer BE, Crocker GJ (1998) Comparative effects of subterranean clover, medic, lucerne, and chickpea in wheat rotations, on nitrogen, organic carbon, and moisture in two contrasting soils. *Soil Research* 36, 57–72. doi:10.1071/S97036
- Hütsch BW, Webster CP, Powlson DS (1993) Long-term effects of nitrogen fertilization on methane oxidation in soil of the broadbalk wheat experiment. *Soil Biology & Biochemistry* 25, 1307–1315. doi:10.1016/ 0038-0717(93)90045-D
- International Atomic Energy Agency (IAEA) (2000) 'Optimizing nitrogen fertilizer application to irrigated wheat: Results of a co-ordinated research project organized by the Joint FAO/IEAEA Division of Nuclear Techniques in Food and Agriculture 1994–1998.' (International Atomic Energy Agency: Vienna). Available at http://www-pub.iaea.org/MTCD/ publications/PDF/te\_1164\_prn.pdf [verified 28 May 2016]
- IPCC (2006) '2006 IPCC guidelines for national greenhouse gas inventories. Volume 4. Agriculture, forestry and other land use.' (Eds HS Eggleston, L Buendia, K Miwa T Ngara, K Tanabe). (Institute for Global Environmental Strategies: Hayama, Japan)
- Isbell RF (2002) 'The Australian soil classification.' Revised edn. (CSIRO Publishing: Melbourne)
- Le Mer J, Roger P (2001) Production, oxidation, emission and consumption of methane by soils: A review. *European Journal of Soil Biology* 37, 25–50. doi:10.1016/S1164-5563(01)01067-6
- Li Y, Chen D, Barker-Reid F, Eckard R (2008) Simulation of N<sub>2</sub>O emissions from rain-fed wheat and the impact of climate variation in southeastern Australia. *Plant and Soil* **309**, 239–251. doi:10.1007/s11 104-008-9672-7
- Liu C, Wang K, Zheng X (2013) Effects of nitrification inhibitors (DCD and DMPP) on nitrous oxide emission, crop yield and nitrogen uptake in a wheat-maize cropping system. *Biogeosciences* 10, 2427–2437. doi:10.5194/bg-10-2427-2013
- Menéndez S, Barrena I, Setien I, González-Murua C, Estavillo JM (2012) Efficiency of nitrification inhibitor DMPP to reduce nitrous oxide emissions under different temperature and moisture conditions. *Soil Biology & Biochemistry* 53, 82–89. doi:10.1016/j.soilbio.2012.04.026
- Officer SJ, Phillips F, Armstrong R, Kelly K (2008) Nitrous oxide emissions from dry-land wheat in south-eastern Australia. In 'Global issues paddock action. Proceedings 14th Australian Agronomy Conference'. Adelaide, South Australia. (Australian Society of Agronomy, The Regional Institute: Gosford, NSW). Available at http://www.regional. org.au/au/asa/2008/poster/farming-uncertain-climate/5831\_officersj. htm [verified 28 May 2016]
- Officer SF, Phillips F, Armstrong R, Graham C (2010) Nitrogen fertiliser increases nitrous oxide emissions from a semi arid Vertosol. In 'Proceedings 19th World Congress of Soil Science'. Brisbane, Qld. (Eds RJ Gilkes, N Prakongkep) pp. 168–171. (International Union of Soil Sciences: Vienna)
- Ogle S, Olander L, Wollenberg L, Rosenstock T, Tuniello F, Paustian K, Buendia L, Nihart A, Smith P (2014) Reducing greenhouse gas emissions and adapting agricultural management for climate change in developing countries: providing the basis for action. *Global Change Biology* 20, 1–6. doi:10.1111/gcb.12361
- Pedersen AR, Petersen SO, Schelde K (2010) A comprehensive approach to soil-atmosphere trace-gas flux estimation with static chambers.

*European Journal of Soil Science* **61**, 888–902. doi:10.1111/j.1365-2389.2010.01291.x

- Plaza-Bonilla D, Cantero-Martínez C, Bareche J, Arrúe J, Álvaro-Fuentes J (2014) Soil carbon dioxide and methane fluxes as affected by tillage and N fertilization in dryland conditions. *Plant and Soil* **381**, 111–130. doi:10.1007/s11104-014-2115-8
- Rodriguez D, Probert M, Meyer M, Galbally I, Howden M, Bennett A, Nussey R, Chen D, Strong W (2003) 'Background study into greenhouse gas emissions from the grains industry (DAV478).' (Grains Research and Development Corporation: Canberra, ACT). Available at http:// www.greenhouse.unimelb.edu.au/pdf\_files/GHGGrainsEmissions.pdf [verified 28 May 2016]
- Scheer C, Grace PR, Rowlings DW, Kimber S, Van Zwieten L (2011) Effect of biochar amendment on the soil-atmosphere exchange of greenhouse gases from an intensive subtropical pasture in northern New South Wales, Australia. *Plant and Soil* 345, 47–58. doi:10.1007/s11104-011-0759-1
- Scheer C, Grace P, Rowlings D, Payero J (2012) Nitrous oxide emissions from irrigated wheat in Australia: impact of irrigation management. *Plant and Soil* 359, 351–362. doi:10.1007/s11104-012-1197-4
- Scheer C, Rowlings DW, Firrel M, Deuter P, Morris S, Grace PR (2014) Impact of nitrification inhibitor (DMPP) on soil nitrous oxide emissions from an intensive broccoli production system in sub-tropical Australia. *Soil Biology & Biochemistry* 77, 243–251. doi:10.1016/j.soilbio.2014. 07.006
- Scheer C, Rowlings DW, De Antoni Migliorati M, Lester DW, Bell MJ, Grace PR (2016) Effect of enhanced efficiency fertilizers on nitrous oxide emissions in a sub-tropical cereal cropping system. *Soil Research* 54, 544–551. doi:10.1071/SR15332
- Scherbak I, Millar N, Robertson GP (2014) Global metaanalysis of the nonlinear response of soil nitrous oxide (N<sub>2</sub>O) emissions to fertilizer nitrogen. Proceedings of the National Academy of Sciences of the United States of America 111, 9199–9204. doi:10.1073/pnas.13224 34111
- Schwenke GD, Haigh BM (2016) The interaction of seasonal rainfall and nitrogen fertiliser rate on soil N<sub>2</sub>O emission rate, total N loss and crop yield of dryland sorghum and sunflower grown on sub-tropical Vertosols. Soil Research 54, 604–618. doi:10.1071/SR15286
- Schwenke GD, Herridge DF, Scheer C, Rowlings DW, Haigh BM, McMullen KG (2015) Soil N<sub>2</sub>O emissions under N<sub>2</sub>-fixing legumes and N-fertilised canola: A reappraisal of emissions factor calculations. *Agriculture, Ecosystems & Environment* **202**, 232–242. doi:10.1016/ j.agee.2015.01.017
- Shcherbak I, Grace P (2014) Determination of emission factors for estimating fertiliser-induced nitrous oxide emissions from Australia's rural production systems. Report to the Australian Department of the Environment.
- Syamsul Arif MA, Houwen F, Verstraete W (1996) Agricultural factors affecting methane oxidation in arable soil. *Biology and Fertility of Soils* 21, 95–102. doi:10.1007/BF00335999
- Van Groenigen JW, Velthof GL, Oenema O, Van Groenigen KJ, Van Kessel C (2010) Towards an agronomic assessment of N<sub>2</sub>O emissions: a case study for arable crops. *European Journal of Soil Science* 61, 903–913. doi:10.1111/j.1365-2389.2009.01217.x
- Venterea RT, Coulter JA (2015) Split application of urea does not decrease and may increase nitrous oxide emissions in rainfed corn. Agronomy Journal 107, 337–348. doi:10.2134/agronj14.0411
- Wang W, Dalal RC, Reeves SH, Butterbach-Bahl K, Kiese R (2011) Greenhouse gas fluxes from an Australian subtropical cropland under long-term contrasting management regimes. *Global Change Biology* 17, 3089–3101. doi:10.1111/j.1365-2486.2011.02458.x
- Webb AA, Grundy MJ, Powell B, Littleboy M (1997) The Australian sub-tropical cereal belt: soils, climate and agriculture. In 'Sustainable crop production in the sub-tropics: an Australian perspective'. (Eds AL

650 Soil Research

Clarke, PB Wylie) pp. 8–26. (Queensland Department of Primary Industries: Brisbane, Qld)

- Weier KL (1999) N<sub>2</sub>O and CH<sub>4</sub> emission and CH<sub>4</sub> consumption in a sugarcane soil after variation in nitrogen and water application. *Soil Biology & Biochemistry* **31**, 1931–1941. doi:10.1016/S0038-0717(99) 00111-X
- Weiske A, Benckiser G, Ottow JG (2001) Effect of the new nitrification inhibitor DMPP in comparison to DCD on nitrous oxide (N<sub>2</sub>O) emissions and methane (CH<sub>4</sub>) oxidation during 3 years of repeated applications in field experiments. *Nutrient Cycling in Agroecosystems* 60, 57–64. doi:10.1023/A:1012669500547
- Young R, Wilson BR, McLeod M, Alston C (2005) Carbon storage in the soils and vegetation of contrasting land uses in northern New South Wales, Australia. *Australian Journal of Soil Research* 43, 21–31. doi:10. 1071/SR04032
- Zadoks JC, Chang TT, Konzak CF (1974) A decimal code for the growth stages of cereals. *Weed Research* 14, 415–421. doi:10.1111/j.1365-3180.1974.tb01084.x
- Zebarth BJ, Rochette P, Burton DL, Price M (2008) Effect of fertilizer nitrogen management on N<sub>2</sub>O emissions in commercial corn fields. *Canadian Journal of Soil Science* 88, 189–195. doi:10.4141/CJSS06010