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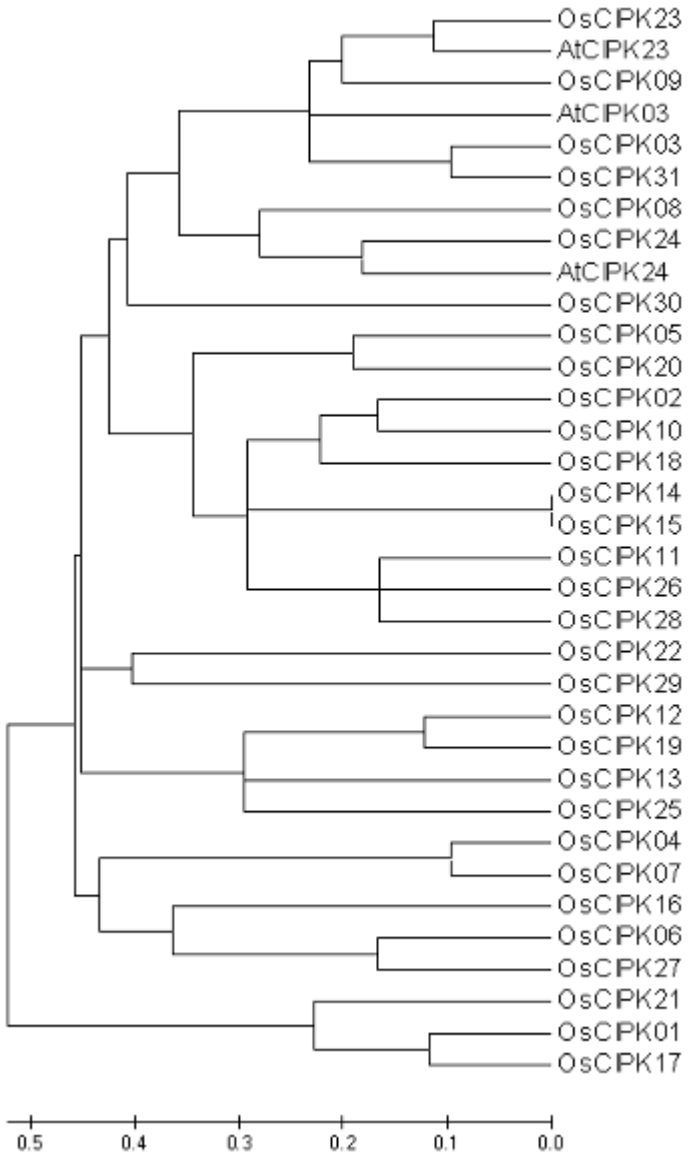

Fig S1. Phylogenetic tree of the rice CIPK family and three CIPKs from *Arabidopsis*. The OsCIPK03 sequence was aligned with all 31 rice CIPKs and three functional *Arabidopsis* CIPKs (AtCIPK03, AtCIPK23 and AtCIPK24). The dendrogram was constructed using the MEGA 3 software.

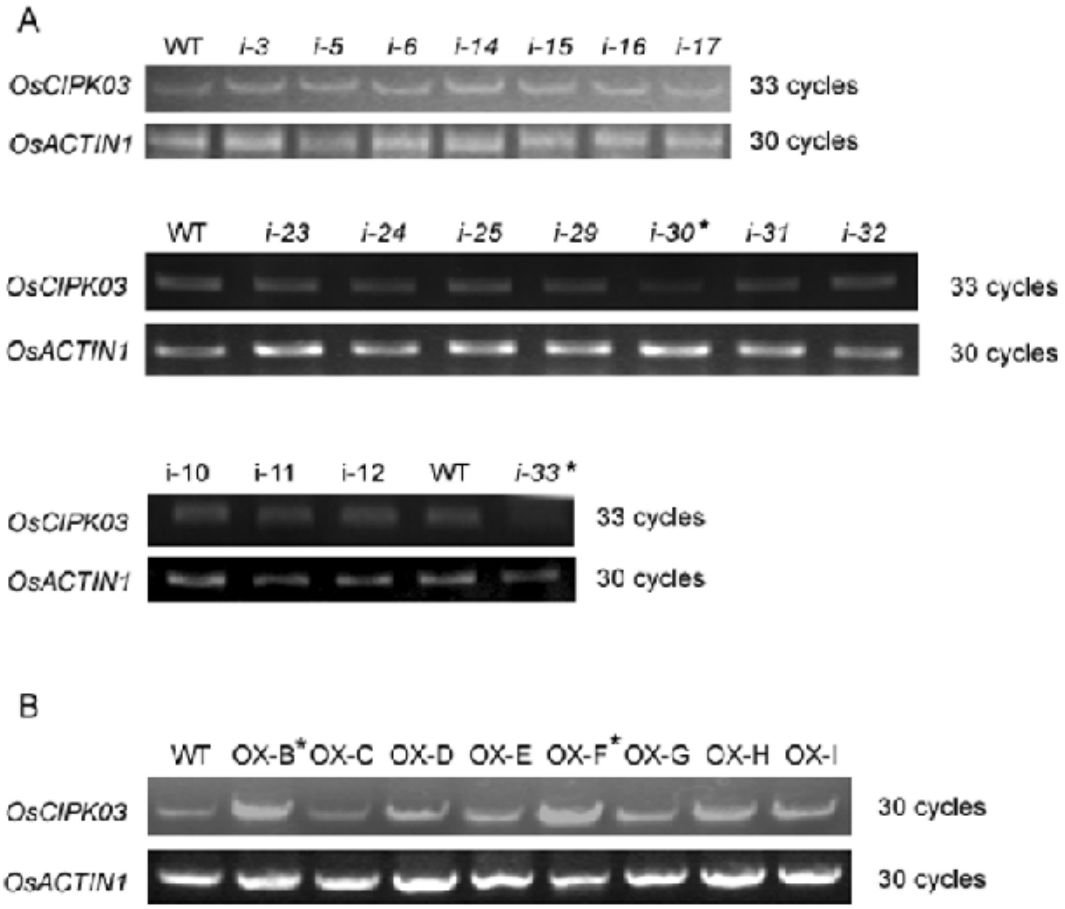

Fig S2. Analysis of the *OsCIPK03* transcript levels in T0 transgenic rice plants. (A) Semi-quantitative RT-PCR analysis of *OsCIPK03* transcript levels in the wild-type and eighteen RNAi transgenic lines. The *Actin1* gene was used as a loading control. (B) Semi-quantitative RT-PCR analysis of *OsCIPK03* transcript levels in the wild-type and eight overexpression transgenic lines. The *Actin1* was used as a loading control. The reactions were amplified for 33 cycles for *OsCIPK03* expression in RNAi transgenic lines and 30 cycles for others. The selected lines are marked with a black asterisk.

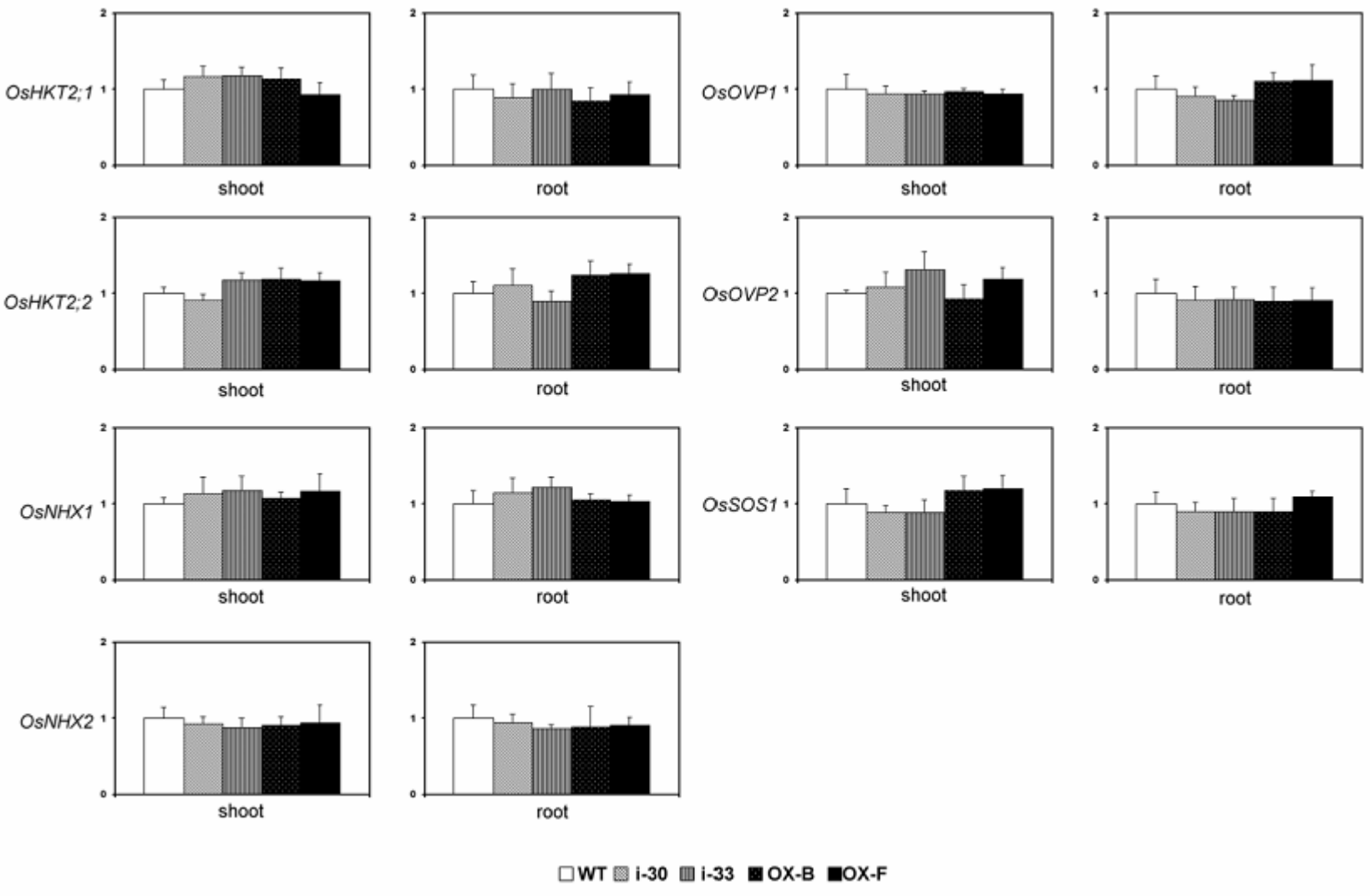
OsHKT2;1
OsHKT2;2
OsNHX1
OsNHX2
OsOVP1
OsOVP2
OsSOS1
shoot
root
0
1
2
WT
i-30
i-33
OX-B
OX-F

Fig S3. Expression patterns of seven functional genes in transgenic plants under salt stress. Transcript levels of *OsHKT2;1*, *OsHKT2;2*, *OsNHX1*, *OsNHX2*, *OsOVP1*, *OsOVP2* and *OsSOS1* were determined by RT-PCR from total RNA isolated from the shoots and roots of 2-week-old plants, respectively. The plants were treated with 200 mM NaCl for 24 h. The levels of the corresponding genes in the WT control are set to 1. The experiments were repeated three times, and similar tendency was observed. The expression levels shown here were according to one measurement. Error bars are standard deviations of three technical repeats.

Table S1. List of primers used in the study

The primers used for plasmid construction were listed as follows:	
OsCIPK03 overexpression construction	5'-TCCCCCGGGGATGTATAAGGCTAAAAGGAC-3' and 5'-TCCCCCGGGTTAGCTCATATCCTCAGC-3' (*Sma I* site underlined)
OsCIPK03 RNAi construction	5'-CACCCAGCCAACCTTGATGAATG-3' and 5'-CTCATATCCTCAGCCGCATC-3'
The primers used for semi-quantitative RT-PCR were listed as follows:	
Actin1	5'-TGGCATCTCTCAGCACATTCC-3' and 5'-TGCACAATGGATGGGTCAGA-3'
OsCIPK03	5'-GGAATTCATGAATGCCTTTGAGTTG-3' and 5'-ATAAGAATGCGGCCGCTTAGCTCATATCCTCAGC-3'
OsCIPK31	5'-CAGGCACTGAATCTGGACAA-3' and 5'-GCTACTCTACGGCGAACACC-3'
The primers used for quantitative real-time PCR were listed as follows:	
Actin1	5'-TGGCATCTCTCAGCACATTCC-3' and 5'-TGCACAATGGATGGGTCAGA-3'
eEF1α	5'-TTTCACTCTTGGTGTGAAGCAGAT-3' and 5'-GACTTCCTTCACGATTTCATCGTAA-3'
OsCIPK03	5'-ATAAGAATGCGGCCGCGATGTATAAGGCTAAAAGG-3' and 5'-ATAAGAATGCGGCCGCCATCAAGGTTGGCTGCTC -3'
RD29A	5'-TGGATCAAACAGAGGAACCA-3' and 5'-CATCTTAGTCGCACCATTCTCA-3'
Rab16	5'-GCCAGTTCCAGCCGATGA-3' and 5'-GGTGTCGGTGGTGGTGGT-3'
CatA	5'-GGATGACACCAAGACATG-3' and 5'-TCACGTTGAGCCTATTCG-3'

OsVHA-B	5'-ATTGACAGGCAGCTGCAT-3' and 5'-GCAATGTCCATGCTAGGT-3'
OsAKT1	5'-TGCCAGTTGTTGCGTTTG-3' and 5'-ATATGCAGTGCGGTATGCC-3'
OsHKT2;1	5'-TCCATCGACTGCTCACTCA-3' and 5'-TGTTGTCGATGGTGGTAAGTACA-3'
OsHKT2;2	5'-CAACGCGGTCTTTGTGATAG-3' and 5'-CCTACATTCCCATATGCACTGA-3'
OsNHX1	5'-CTGTCGTTCTTTTTAGCACTATGG-3' and 5'-GGTGACAGGATGGCCTGA-3'
OsNHX2	5'-TTATCTGTTTGCCACGAGCA-3' and 5'-CCTGCCGAAGTACAGCTTTT-3'
OsOVP1	5'-CCCAAGAAACCCAGCTGTC-3' and 5'-CATAGGAACCAAAGAGATCTGACC-3'
OsOVP2	5'-GTACGGGGACTACCTCATCG-3' and 5'-CGTGAAGAGAAATGATGTTGCT-3'
OsSOS1	5'-TTGCTGTCAGCTACATTGCTTT-3' and 5'-CCAGTGTCATGACGGTCAAA-3'