

Supplementary Material

Steroids in the equine oviduct: synthesis, local concentrations and receptor expression

Hilde Nelis^A, Julie Vanden Bussche^B, Bartosz Wojciechowicz^E, Anita Franczak^E, Lynn Vanhaecke^B, Bart Leemans^A, Pieter Cornillie^D, Luc Peelman^C, Ann Van Soom^A and Katrien Smits^{A,F}

^AGhent University, Department of Reproduction, Obstetrics and Herd Health, Faculty of Veterinary Medicine, Salisburylaan 133, 9820 Merelbeke, Belgium.

^BGhent University, Department of Veterinary Public Health and Food Safety, Laboratory of Chemical Analysis, Faculty of Veterinary Medicine, Salisburylaan 133, 9820 Merelbeke, Belgium.

^CGhent University, Department of Nutrition, Genetics and Ethology, Faculty of Veterinary Medicine, Heidestraat 19, 9820 Merelbeke, Belgium.

^DGhent University, Department of Morphology, Faculty of Veterinary Medicine, Salisburylaan 133, 9820 Merelbeke, Belgium.

^EUniversity of Warmia and Mazury, Department of Animal Physiology, Faculty of Biology and Biotechnology, Oczapowskiego St. 1A, 10-719 Olsztyn, Poland.

^FCorresponding author. Email: katrien.smits@ugent.be

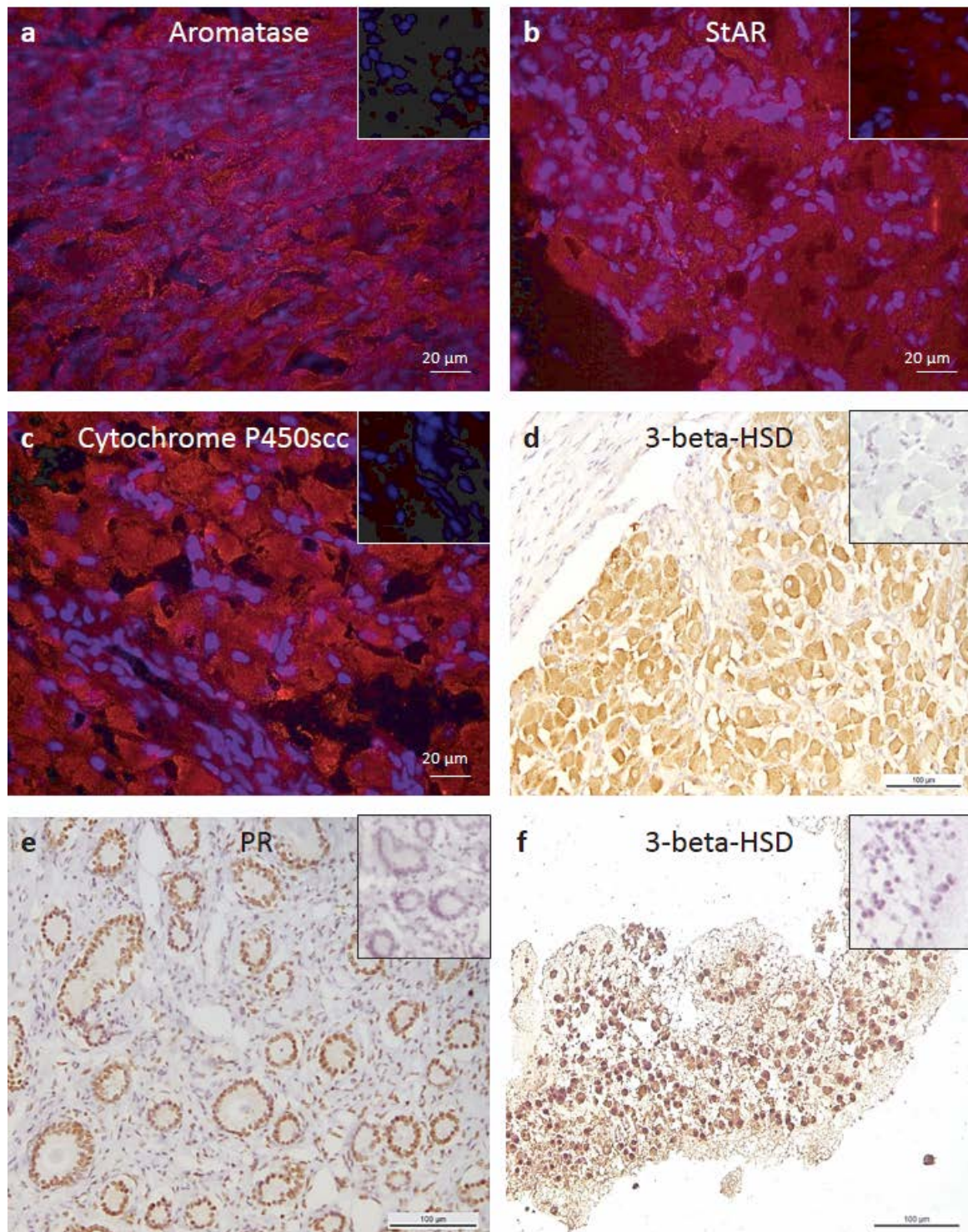


Fig. S1. Epifluorescent (a–c) and immunohistochemically (d–f) positive tissue control pictures and negative control slides without primary antibody (right upper corners) in the equine oviduct for aromatase (a), StAR (b), cytochrome P450scc (c), 3-beta-HSD (d, f) and PR (e). Positive control tissue was equine follicular wall (a), corpus luteum (b–d) and endometrium (e). 3-Beta-HSD staining (brown) with hematoxylin counterstaining (purple) of equine granulosal cells (f). Both the cytoplasm and the nuclei of cumulus cells stain strongly. Bar a–c = 20 µm, bar d–f = 100 µm.

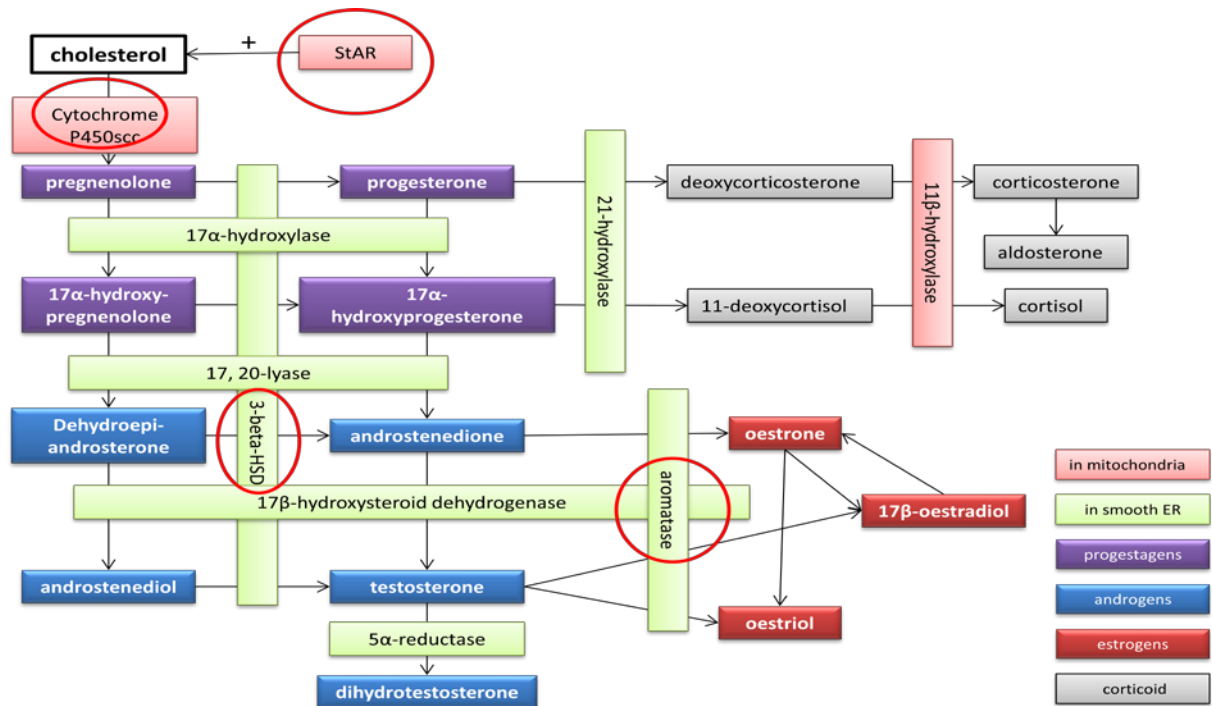


Fig. S2. Overview of steroidogenesis (based on Lavoie and King 2009). The enzymes discussed in the paper are encircled in red.