

S33-002

## Future food requirements: are improvements in photosynthesis required?

JE Sheehy

*Crop, Soil and Water Sciences Division, International Rice Research Institute (IRRI), DAPO Box 7777, Metro Manila, Philippines. j.sheehy@cgiar.org*

Keywords: rice, yield, climate change, canopy photosynthesis

### Introduction

The total amount of carbon captured in photosynthesis each year by terrestrial ecosystems is approximately  $120 \times 10^9$  t, of which half is lost in respiration (Steffen et al., 1998). Approximately  $1.6 \times 10^9$  t C are fixed by the major cereals each year and about 43% of that is in the grain. Although cereals only account for approximately 3% of total net carbon fixed, they are of enormous importance in terms of food for humans.

Solar energy captured in photosynthesis gives plants the capacity to synthesize, organize and maintain a range of structural units housing a myriad of metabolic processes. The two aspects of energy capture that are of principal interest to crop physiologists set the limits for crop growth rate. The first is the quantity of energy captured in unit time and that depends on the available irradiance and the fraction absorbed by the crop canopy. The second is the efficiency with which the absorbed energy is used for the chain of synthetic processes that culminate in harvestable yield.

While it is true that we cannot rely on demographic predictions as we rely on astronomical ones (Medawar, 1987), their importance means we cannot ignore them. Over the next 50 years it is predicted that the population of Asia will rise from 3.7 billion to 5.3 billion (UNFPA, 1999). Increases in rice production will have to be produced with less water, less fertilizer and less land (Hossain and Pingali, 1998; Tilman et al., 2001; Evans, 1998). Currently, half of the people in Southeast Asia have a calorie intake inadequate for an active life. Yields in many Asian countries have reached a plateau (Cassman, 1999) and yields in breeders trials at IRRI have not increased for 30 years (Sheehy, 2001a); indeed it has been suggested that a yield barrier has been reached (Kropff et al., 1994). How can the projected rice demands for Asia be met? Clearly, water and nitrogen use efficiencies have to increase, as does yield by more than 44%.

The phenotype of a given genotype can vary markedly according to its interaction with the environment (Mifflin, 2000). Given the genetic complexity that underlies that plasticity and that the 'same' crop is grown in geographically different regions with different climates, weather conditions and on different soil types with different histories of management, it is not surprising that in field experimentation precise repeatability, in the usual scientific sense, is the exception rather than the rule. As a result of this imprecision and the absence of universally acceptable theoretical models of crop growth, disagreements about what precisely determines both biomass and grain yield are commonplace. Why the yield potential of new rice cultivars at IRRI has remained approximately constant for more than 30 years (Sheehy, 2001a; Kropff et al., 1994) is also a matter for speculation. Nevertheless, it is simply not good enough to be optimistic that 'business as usual' will solve the problem of increasing rice yield. New and possibly radical approaches need to be explored urgently. What relevance has photosynthesis research in this context?

### *Link between photosynthesis, yield and radiation use efficiency*

For crops, the law of mass conservation can be used to link growth rate, crop photosynthesis, respiration and the loss of biomass by detachment. The product of the integral of crop growth rate and harvest index gives grain yield; it should be noted that grain yields for rice are quoted at 14% moisture content. To make progress and to derive a simple equation linking yield and canopy gross photosynthesis, Sheehy (2000) assumed a constant root weight ratio and units of  $\text{CH}_2\text{O}$ , so that the equation is

$$Y = H \left\{ (1-\beta) \int_{t_i}^{t_f} [P_g(t) - R(t) - D(t)] dt \right\} \quad (1)$$

where  $Y$  is grain yield,  $H$  is harvest index (unless otherwise stated calculated as the fraction of aboveground dry weight that is grain weight),  $\beta$  is the root weight ratio,  $P_g$  is the rate of canopy gross photosynthesis,  $R$  is the rate of shoot and root respiration,  $D$  is the loss of dry matter through detachment,  $t$  is time (usually a time step of a day is used),  $t_i$  could be the day of germination or some other suitable starting time and  $t_f$  is the day of harvest.

Equation (1) shows the link between photosynthesis and yield is mediated by several factors that could confound simple attempts to correlate photosynthesis with yield. The most obvious are differences in crop duration ( $t_f - t_i$ ), or a difference in the partitioning of assimilates to roots,  $\beta$ . Harvest index,  $H$ , can be affected by many factors, e.g. differences in susceptibility to thermally induced sterility (Satake and Yoshida, 1978; Horie, 1993) or differences in the ability to partition nitrogen to the grain (Sinclair, 1998; Sinclair and Sheehy, 1999). Nevertheless, it would be most surprising if yield improvements in modern cultivars were unaccompanied by improvements in canopy photosynthesis (Robson, 1982; Long, 1999a,b). Surrogates of photosynthesis, such as canopy air temperature depression, are proving useful in screening wheat for yield (Reynolds et al., 2000; Evans and Fischer, 1999). It is interesting to note that measurements I made in high irradiance show erect leaves in rice canopies can have temperatures  $2.5^\circ\text{C}$  cooler than prostrate leaves ( $\sim 31^\circ\text{C}$  v  $33.5^\circ\text{C}$ ) suggesting there may be a link between canopy architecture and canopy temperature. The relationship between leaf and canopy photosynthesis is presented later.

It is convenient to give a definition of radiation use efficiency here. It is assumed that radiation use efficiency ( $\epsilon$ ) is the slope of the linear relationship between shoot dry weight and accumulated intercepted radiation (Monteith, 1977). The value for any day (Sheehy, 2000) can be calculated as

$$\epsilon = (dWs/dt)/I_{\text{int}}, \quad (2)$$

where  $dWs/dt$  is the daily growth rate of the shoots,  $I_{\text{int}}$  is the total amount of photosynthetically active radiation (PAR) intercepted by the crop for the same day. By substituting for crop growth rate in equation (2),  $\epsilon$  can be written

$$\epsilon = [(1-\beta) (P_g(t) - R(t) - D(t))] / I_{\text{int}}. \quad (3)$$

The average value of  $\epsilon$  for rice is  $2.2 \text{ g DW MJ}^{-1} \text{ PAR}$  and for maize is  $3.3 \text{ g DW MJ}^{-1} \text{ PAR}$  (Kiniry et al., 1989, Mitchell et al., 1998). Using a simple model, Mitchell et al. (1998) concluded that the value of  $\epsilon$  for rice would have to rise to that of maize to increase yields by 50% given a constant crop duration. The simplest of calculations using equation (3) shows that photosynthesis would account for the difference between maize and rice assuming the same dry weight, optimum nitrogen contents (Greenwood et al., 1990) and temperatures of approximately  $30^\circ\text{C}$  (Sage, 2000).

Based on the model of McCree (1970) respiration is often divided between synthetic and maintenance respiration. Differences in synthetic respiration can exist between species depending on the nature of the plant products synthesized (Penning De Vries et al., 1974). Processes such as biological nitrogen fixation require approximately 25% of daily photosynthate (Witty et al., 1983), but field-grown legumes can compensate and show no yield or quality loss when compared with nitrate-grown crops (Sheehy and McNeill, 1988). Large differences in the ratio of synthetic respiration to photosynthesis between varieties within a species are unlikely. Loomis and Amthor (1999) discussed maintenance respiration in some detail and only a brief synopsis will be given here. Variation can exist in the maintenance coefficient (Wilson, 1975; Robson, 1982), but the values obtained by McCree (1970) and Ryle et al. (1976) were broadly similar and the assumption that the maintenance coefficient is approximately constant, but strongly affected by temperature, is often made. Interestingly, Cock and Yoshida (1973) concluded that the maintenance coefficient for respiration during late grain filling was negligible. Cannell and Thornley (2000) suggested that current understanding of many plant processes was now sufficient to enable a more sophisticated approach to the modelling of respiration than that taken by McCree (1970). However, Mitchell et al. (1991) found the McCree (1970) approach useful for interpreting their field measurements of respiration.

In grain crops, senescence and detachment are largely associated with nutrient remobilization during grain filling. Sinclair and Sheehy (1999) used a simple model to show that differences between cultivars in the ability to transfer nitrogen to the grain from senescent leaves was related to canopy erectness. The more or less continuous 'throw away lifestyle' with respect to leaves of plants (Thomas, 1994a) is more likely to deliver advantages to forage crops experiencing grazing than grain crops where plant breeders have selected for high harvest index and leaf senescence during grain filling.

This leads onto the link between canopy and leaf photosynthesis. There are many models of canopy gross photosynthesis and irradiance, and the relationship is approximately hyperbolic (Cock and Yoshida, 1973; Sheehy and Peacock, 1975; Thornley and Johnson, 1990; Luo et al., 2000); the operative part of the curve is usually far from the plateau. What is common to most models is a combination of parameters describing individual leaf photosynthesis, leaf area index and light distribution. The initial slope of the relationship is a function of the quantum yield of an individual leaf at vanishing levels of PAR irradiance, light interception and the leaf transmission coefficient (Sheehy and Johnson, 1988). The asymptotic value is a function of the product of leaf area index and the maximum rate of leaf photosynthesis.

Farquhar et al. (1980) and Von Caemmerer (2000) have modelled leaf photosynthesis mechanistically in terms of the kinetic properties of rubisco. Those models have enabled both the quantum yield of leaf photosynthesis and its maximum rate to be described at a process level. The rate of carboxylation is the minimum of the rubisco limited rate, the rate of photosynthetic electron transport rate and the rate of triose phosphate conversion to photosynthetic end products (Farquhar and Sharkey, 1994). These equations have appealed sufficiently to ecologists to use them in modelling vegetation at a global scale in relation to climate change (Osborne et al., 2000; Woodward, 1999). Finally, in moving from measurements of leaf photosynthesis on spaced plants to crop plants, changes in leaf area (Bhagsari and Brown, 1986; Pearce et al., 1969) and leaf nitrogen content in response to competition for light can confound the unwary (Evans, 1993; Peng et al., 1995).

#### *Upper limits to canopy photosynthesis*

Sheehy (2000) used the conveniently simple model of Monteith (1965) to show that the maximum photosynthetic leaf area index of an erect canopy is approximately 6.6. The sunlit leaf area index ( $L_0$ ) reached a maximum of 3.3 and photosynthetically active shaded leaf area

index ( $L_s$ ) reached its maximum at approximately 3.0. Cock and Yoshida (1973) observed little increase in canopy photosynthesis above an LAI of 5.0 in both tall and short rice varieties; a similar result was obtained for grasses (Parsons, 1988). Sinclair and Sheehy (1999) proposed that the third class of leaves in the canopy ( $L_N$ ) were those that were close to the light compensation point and largely useful for the N they contributed to the grain ( $L_N = L - L_0 - L_s$ ; where  $L$  is the total LAI). Sheehy (2000) went on to show that the loss through senescence of  $L_s$  and  $L_N$  would reduce canopy photosynthesis by about 22-27%. In rice canopies, the distal parts of some leaves are exposed to full sunlight while the proximal parts are at the compensation point.

The maximum rate of canopy photosynthesis ( $P_{gmax}$ ) has an asymptotic value given by

$$P_{gmax} = L_0 A_{mo} + L_s A_{ms} \quad \text{as } I \rightarrow \infty, \quad (4)$$

where  $A_{mo}$  is the maximum rate of sunlit leaf photosynthesis and  $A_{ms}$  is the maximum rate of shaded leaf photosynthesis. The maximum quantum yield of the canopy at full light interception is

$$dP_g/dI_0 = \alpha(1 + \tau) \quad \text{as } I \rightarrow 0, \quad (5)$$

where  $\alpha$  is the leaf quantum yield in low irradiance and  $\tau$  is leaf transmittance. It can be seen that the limits for canopy photosynthesis are set by the properties of the individual leaves. At LAIs greater than approximately 6.0, improvements in canopy photosynthesis can only result from improvements in leaf quantum yield and/or leaf photosynthesis.

Leaf quantum yield is almost constant for all  $C_3$  plants and within subtypes of  $C_4$  plants (Ehleringer and Pearcy, 1983; Long et al., 1993); it is influenced by the relative concentrations of  $O_2$  and  $CO_2$  around rubisco and the costs of the biochemical transactions involved in carboxylation. The maximum rate of leaf photosynthesis for both  $C_3$  and  $C_4$  plants appears to vary linearly with nitrogen content per unit leaf area over a wide range of nitrogen concentrations (Evans and von Caemmerer, 2000; Peng et al., 1995). Assuming the data shown by Evans could be described by linear relationships, then maize had a gradient of approximately  $0.74 \mu\text{mol } CO_2 \text{ s}^{-1} \text{ mmol}^{-1} \text{ N}$  compared with that for  $C_3$  plants (rice and wheat) of  $0.26 \mu\text{mol } CO_2 \text{ s}^{-1} \text{ mmol}^{-1} \text{ N}$ ; a threefold difference in leaf photosynthetic nitrogen use efficiency. Sinclair and Horie (1989) showed saturation values for the photosynthesis-nitrogen relationship and that would be expected at 'luxury' N levels. Indeed, the Greenwood et al. (1990) critical nitrogen model relating nitrogen content to shoot biomass states that there is an optimum relationship for both  $C_3$  and  $C_4$  crops. It is clear that very high rates of leaf photosynthesis are possible in  $C_3$  leaves, but past some critical nitrogen content, the Greenwood relationship means such high rates are probably purchased at the cost of lower leaf area indices in crops and resulting in no overall gain in biomass (Lawlor, 1995). The issue of what sets the maximum rate of nitrogen acquisition and utilization by a crop is not a simple question to answer, but it is likely that the availability of energy will be a major part of the answer. High nitrogen fertilizer applications are a source of nitrogen pollution because of the low recovery (< 40%) in rice systems (Dobermann, 2000). An improvement in the both the rate of leaf photosynthesis and the rate per unit of leaf nitrogen is desirable if improvements in canopy photosynthesis are needed.

#### *A yield decline across 30 years and possible threats from climate change*

From seasonal observations showing positive effects of elevated  $CO_2$  on yield (Baker et al., 1992; Ziska et al., 1997), it is often concluded that projected increases in the partial pressure of atmospheric  $CO_2$  ( $pCO_2$ ) will result in yield increases. However, the yield of the best entry in breeders' trials at IRRI for the past thirty years has not increased (Sheehy, 2001a).

Furthermore, the data of Peng et al. (1999) shows that the yields for the same cultivar (IR8) grown on the IRRI farm in 1998 were about  $2.6 \text{ t ha}^{-1}$  lower than in 1966 at nitrogen inputs of approximately  $150 \text{ kg N ha}^{-1}$ . Those authors attributed the yield differences to differences in grain filling, but what caused grain filling to be lower in 1998 is obscure. Weather data collected from nearby meteorological stations for the two growing seasons are very similar and offer no explanation. Average atmospheric  $p\text{CO}_2$  increased by approximately 4.6 Pa between 1966 and 1998 (Keeling and Whorf, 1999). Peng et al. (1999) also showed that the modern cultivar IR72 outyielded IR8 in 1998, suggesting that plant breeders may have been selecting unwittingly, but successfully, for tolerance of some adverse environmental factor or factors. The adverse factors could be additional quantities of atmospheric pollutants such as  $\text{SO}_2$  or  $\text{O}_3$  (Mansfield, 1999; Unsworth and Colls, 1994; Maggs and Ashmore, 1998; Ewert and Porter, 2000) and/or toxic phenolic acids formed in soil reduction (Kirk, pers. com.). In that context, it is interesting to note that vehicle numbers increased from approximately  $310^5$  to  $3 \cdot 10^6$  between 1966 and 1998 in the Philippines. The added complication of rises in temperature that will accompany rises in  $\text{CO}_2$  have been considered by Mathews et al. (1995), Horie et al. (1995), Ziska et al. (1997) and Baker et al. (1992) and they showed that temperature effects on sterility could offset such gains. The risk of exceptional high temperature events on photosynthesis itself has not been calculated.

*Do crop physiologists really understand enough about the processes shaping yield to clearly identify the yield-limiting problems requiring solutions?*

In high yielding rice grown at IRRI, two distinct phases of shoot growth appear to be separated by the onset of flowering. Shoot growth does not follow a simple law of diminishing returns and this unusual aspect of crop ontogeny was not described by Horie (1994) or Thomas (1994b). Why growth slows down at the start of flowering and then speeds up towards maturity is again, a matter of speculation. The first phase of shoot growth is relatively simple, being concerned largely with the production of tillers, leaves and roots. That phase is characterized by increasing leaf area index and light interception, it ends with the simultaneous growth of the flag leaf, culm and the panicle within the leaf sheaths. The second phase is more complex, characterized by the emergence and growth of the panicle, flowering and grain filling. Developing spikelets act as a powerful sinks for both current and stored assimilates accumulating nearly 50% of the total biomass by maturity. Roots, for whatever reason, do not supply sufficient nitrogen to support the demand from the developing grains and the leaf area index declines as leaves die and transfer much of their nitrogen to the grains (Sinclair and Sheehy, 1999; Sheehy, 2001a). Short-term estimates of radiation use efficiency (Sheehy et al., 1998) show it to be relatively constant prior to flowering and very variable afterwards, perhaps reflecting short-term changes in the balance between photosynthesis and respiration (Equation 3). Cock and Yoshida (1971) observed that respiration decreased between flowering and maturity in rice, and in a later paper concluded that maintenance respiration in rice decreased to approximately zero towards the end of grain filling (Cock and Yoshida, 1973). Such changes in respiration could go some way to explaining the variation in  $\epsilon$  observed by Sheehy et al. (1998) between flowering and maturity.

Ten days before flowering, potential sink size in the form of juvenile spikelet number per square meter in IR72 was very large,  $106,131 \text{ spikelets m}^{-2}$ , but the number decreased by 46% between panicle emergence and flowering, resulting in  $56,979 \text{ spikelets m}^{-2}$  of which 77% were eventually filled. The failure to convert juvenile spikelets into mature spikelets and to fill them suggests resource availability was the limiting factor not sink size (Sheehy et al., 2001b). That conclusion is partly supported by the increases in spikelet number, filled grains and grain weight resulting from short-term manipulation of  $\text{CO}_2$  at different stages of panicle development (Yoshida, 1973). Because crop growth rate increases towards maturity, failure to

fill mature spikelets could mean that they were sterile by the time resources became sufficiently available to fill them, or that developing tillers became relatively stronger sinks. It is possible that during the period between the formation of the maximum number of juvenile spikelets and the emergence of the panicle bearing mature spikelets, thermally driven demands for assimilates exceeded the ability of the photosynthetic system to supply them (Baker et al., 1992).

## Conclusions

In this paper, I have described the link between yield and crop photosynthesis. In Asia, we cannot be laissez faire with respect to yield improvements resulting simply from increasing  $p\text{CO}_2$ . Other factors might offset such gains; Mitchell and Sheehy (2000) felt that such gains might be too small during the next 50 years. My conclusion is that to improve yields to meet the future demands of Asia for rice, using less nitrogen, water and land, large improvements in photosynthesis will have to be made soon. As a non-expert on photosynthesis, I can only make tentative suggestions on what improvements would be useful based on my reading of the literature. I am aware that Sharkey et al. (2000) suggested that sucrose-phosphate synthetase activity might also have to be enhanced in order to capture the benefits of high photosynthesis, but that is an interface problem I leave to others.

Without access to light energy plants die, so the ability to compete for light must be one measure of fitness in an evolutionary context. For spaced plants, two extreme scenarios can be imagined for the way a unit of nitrogen, captured in unit time, is utilized in different ways to gain the same quantity of photosynthate in the next unit of time. The first is building more photosynthetic apparatus by thickening the existing growing leaf, the second is expanding the area of the growing leaf and distributing the same amount of apparatus in thinner leaves. In communities of plants, the second strategy would disadvantage neighbors adopting the first strategy and it is not surprising that crop plants tend to maximize leaf area rather than the amount of photosynthesis per unit area. Given the evolutionary strategy outlined above, would it be possible to improve crop photosynthesis by suppressing the tendency of crop plants to dilute 'photosynthetic' nitrogen in leaves of high specific leaf area? The detection of neighbors and the adjustment of specific leaf area through the phytochrome system is well understood physiologically, and the molecular biology is becoming known (Smith, 2000). This topic appears ripe for functional genomics to reveal the mechanism, followed by genetic engineering to create the plant with a desired trait that could not be favored by natural selection during evolution, but could be useful in agriculture.

Hauska and Arnold (2000) suggest that the major heat loss in photosynthesis occurs in the very early steps within nanoseconds following quantum absorption. Those authors also asked why there are two light reactions when half the energy, i.e. 4 photons of red light, would suffice. I am left wondering if the quantum yield of photosynthesis in low irradiance could be doubled, thus enabling shaded leaves to photosynthesize at greater rates. Would such an improvement overload the mechanisms of photoprotection and photoinactivation (Osmond et al., 1999) at high irradiance? Horton and Murchie (2000) even suggested that rice was poorly adapted to high light conditions in the tropics.

Mitchell and Sheehy (2000) suggested that transferring the  $C_4$  syndrome to rice would provide a complete solution to how to increase yield, water and nitrogen efficiency simultaneously. Our optimism was based on the pace with which sequencing plant genomes was being made, indications of synteny (Gale and Devos, 1998) between the cereals, and the hope that comparisons made between  $C_4$  and  $C_3$  genomes would enable nature to be imitated even if we could not fully understand the processes. Data of Pamplona et al. (1990) was used to compare water use efficiencies and photosynthetic characteristics of the leaves of the  $C_4$  rice-weed *Echinochloa crus-galli* and rice cv. IR64. The ratio of  $C_4$  to  $C_3$  values show water

use efficiency was three times greater, the quantum yield at low irradiance was 50% greater and the maximum rate of leaf photosynthesis was 30% greater. Björkman (1976) suggested that relatively few genes, inherited independently, were responsible for the C<sub>4</sub> syndrome. However, Edwards and Ku (1987) pointed out that organization was perhaps most important for full expression of C<sub>4</sub>, that without proper compartmentation and interconnection of all the components the C<sub>4</sub> cycle was likely to be futile. Nevertheless, the high level of expression of C<sub>4</sub> genes observed in rice by Ku et al. (1999) is indicative of progress using biotechnological tools.

Evans and von Caemmerer (2000) properly felt that improvements in the properties of rubisco offered greater advantages and a rubisco from red algae with a high specificity for CO<sub>2</sub> has been reported (Uemura et al., 1997). However, Andrews and Lorimer (1987) pointed out that rubisco has been under intense selection for more than 3 billion years suggesting that simple structural improvements are unlikely. Edwards et al. (2001) analyzed what it takes to be C<sub>4</sub> and concluded that pCO<sub>2</sub> late in the 21<sup>st</sup> century might only have a competitive advantage in habitats characterized by high temperatures and drought. The rice yield problem is one of the first half of the century and needs a solution on an appropriate time scale: a decade.

It is clear that improvements leading to yield increases must be the result of placing an emphasis on traits that retain their importance in the phenotypes (Mifflin, 2000) and the C<sub>4</sub> syndrome, an improved rubisco, higher rates of C<sub>3</sub> canopy photosynthesis or higher efficiency of energy use in photosynthesis would satisfy that requirement. All of the aforementioned could increase yields, the first two would increase water and nitrogen use efficiency to a greater extent than the third or fourth options in a future world of increased temperatures and atmospheric pCO<sub>2</sub>. The need to improve photosynthesis is urgent. Failure to meet the rising rice demands of Asia would result in misery for hundreds of millions; the consequences of that are a matter for speculation, but are unlikely to contribute to a better world.

## References

- Andrews TJ, Lorimer GH (1987) *The Biochemistry of Plants* Vol. 10. Academic Press, Inc.
- Baker JT, Allen Jr. LH, Boote KJ (1992) *Journal of Experimental Botany* **43**, 959-964.
- Bhagsari AS, Brown RH (1986) *Crop Science* **26**, 127-132.
- Björkman O (1976) In: Burris RH and Black CC (eds) *CO<sub>2</sub> Metabolism and Plant Productivity*, 287-309. University Park Press, Baltimore.
- Cannell MGR, Thornley JHM (2000) *Annals of Botany* **85**, 45-54.
- Cassman, KG (1999) *Proceedings National Academic Science (USA)* **96**, 5952-5959.
- Cock JH, Yoshida S (1971) *Proceedings of the Crop Science Society of Japan* **41**, 226-233.
- Cock JH, Yoshida S (1973) *Soil Science Plant Nutrition* **19**, 53-59.
- Dobermann A (2000) In: Sheehy JE, Mitchell PL and Hardy B (eds) *Redesigning Rice Photosynthesis to Increase Yield*. International Rice Research Institute, Philippines.
- Edwards GE, Ku MSB (1987) In: Stumpf PK and Conn EE (eds) *The Biochemistry of Plants*, 275-325. Academic Press Inc., California.
- Edwards GE, Furbank RT, Hatch MD, Osmond CB (2001) *Plant Physiology* **125**, 1-4.
- Ehrlinger J, Pearcy RW (1983) *Plant Physiology* **73**, 555-559.
- Evans JR, Von Caemmerer S (2000) In: Sheehy JE, Mitchell PL and Hardy B (eds) *Redesigning Rice Photosynthesis to Increase Yield*. International Rice Research Institute, Philippines.
- Evans LT (1993) *Crop Evolution, Adaptation and Yield*. Cambridge University Press, Great Britain.

- Evans LT (1998) In: Buxton DR, Shibles R, Forsberg RA, Blad BL, Say KH, Paulsen GM and Wilson RF (eds) *International Crop Science I*. Crop Science Society of America, USA.
- Evans LT, Fischer RA (1999) *Crop Science* **39**, 1544-1551.
- Ewert F, Porter JR (2000) *Global Change Biology* **6**, 735-750.
- Farquhar GD, von Caemmerer S, Berry JA (1980) *Planta* **149**, 78-90.
- Farquhar GD, Sharkey TD (1994) In: Boote KJ, Bennett JM, Sinclair TR and Paulsen GM (eds) *Physiology and Determination of Crop Yield*, 153-180. Madison, Wisconsin, USA.
- Gale MD, Devos KM (1998) *Science* **282**, 656.
- Greenwood DJ, Lemaire G, Goose G, Cruz P, Draycott A, Neeteson JJ (1990) *Annals of Botany* **66**, 425-436.
- Hauska G, Arnold M (2000) In: Yunus M, Pathre U and Mohanty P (eds) *Probing Photosynthesis-Mechanisms, regulation and adaptation*, 108-126. Taylor & Francis Inc., London.
- Horie T (1993) *Journal of Agricultural Meteorology* **48**, 567-574.
- Horie T (1994) In: Boote KJ, Bennett JM, Sinclair TR and Paulsen GM (eds) *Physiology and Determination of Crop Yield*, 153-180. Madison, Wisconsin, USA.
- Horie T, Kropff MJ, Centeno HG, Nakagawa H, Nakano J, Kim HY, Ohnishi M (1995) In: Peng S, Ingram KT, Neue HU and Ziska LH (eds) *Climate Change and Rice*. Springer-Verlag Berlin Heidelberg.
- Horton P, Murchie EH (2000) In: Sheehy JE, Mitchell PL and Hardy B (eds) *Redesigning Rice Photosynthesis to Increase Yield*. International Rice Research Institute, Philippines.
- Hossain M, Pingali PL (1998) In: Pingali PL and Hossain M (eds) *Impact of Rice Research*, 1-26. International Rice Research Institute, Philippines.
- Keeling CD, Whorf TP (1999) *Trends: A Compendium of Data on Global Change*. Carbon Dioxide Information Analysis Center, Oak Ridge National Laboratory, U.S. Department of Energy, Oak Ridge, Tenn, U.S.A.
- Kiniry JR, Jones CA, O'Toole JC, Blanchet R, Cabelguenne M, Spanel DA (1989) *Field Crops Research* **20**, 51-64.
- Kropff MJ, Cassman KG, Peng S, Matthews RB, Setter TL (1994) In: Cassman KG (ed) *Breaking the Yield Barrier*. Proceedings of a Workshop on Rice Yield Potential in Favorable Environments, International Rice Research Institute, Philippines.
- Ku MSB, Agarie S, Nomura M, Fukayama H, Tsuchida H, Ono K, Toki S, Miyao M, Matsuoka M (1999) *Nature Biotechnology* **17**, 76-80.
- Lawlor DW (1995) *Journal of Experimental Botany* **46**, 1449-1461.
- Long SP, Postl WF, Bolhár-Nordenkampf HR (1993) *Planta* **189**, 226-234.
- Long SP (1999a) In: Press MC, Scholes JD and Barker MG (eds) *Physiological Plant Ecology*, 263-282. Blackwell Science Ltd., Oxford.
- Long SP (1999b) In: Sage RF and Monson RK (eds) *C<sub>4</sub> Plant Biology*. London (UK), Academic Press.
- Loomis RS, Amthor JS (1999) *Crop Science* **39**, 1584-1596.
- Luo Y, Hui D, Coleman JS, Johnson DW, Sims DA (2000) *Agricultural and Forest Meteorology* **100**, 35-48.
- Maggs R, Ashmore MR (1998) *Environmental Pollution* **103**, 159-170.
- Mansfield TA (1999) In: Press MC, Scholes JD and Barker MG (eds) *Physiological Plant Ecology*, 219-240. Blackwell Science Ltd., Oxford.
- Matthews RB, Kropff MJ, Bachelet D, van Laar HH (1995) *Modeling the impact of climate change on rice production in Asia*, Wallingford (UK): CAB International, in association with the International Rice Research Institute, Manila, Philippines.

- McCree KJ (1970) In: Setlik I (ed) Prediction and measurement of photosynthetic productivity, 221-229. Proc. IBP/PP Tech. Mtg. Trebon, Czechoslovakia. Pudoc, Wageningen, the Netherlands.
- Medawar P (1987) Pluto's Republic. Oxford University Press, Oxford.
- Miflin B (2000) *Journal of Experimental Botany* **51**, 1-8.
- Mitchell PL, Sheehy JE, Woodward FI (1998) IRRI Discussion Paper Series No. 32. Manila, Philippines.
- Mitchell PL, Sheehy JE (2000) In: Sheehy JE, Mitchell PL and Hardy B (eds) Redesigning Rice Photosynthesis to Increase Yield. International Rice Research Institute, Philippines.
- Mitchell RA, Lawlor DW, Young AT (1991) *Annals of Botany* **67**, 7-16.
- Monteith JL (1965) *Annals of Botany* **29**, 17-37.
- Monteith JL (1977) *Philosophical Transactions of the Royal Society of London* **B281**, 277-294.
- Osborne CP, Mitchell PL, Sheehy JE, Woodward FI (2000) *Global Change Biology* **6**, 445-458.
- Osmond CB, Anderson JM, Ball MC, Egerton JJG (1999) In: Press MC, Scholes JD and Barker MG (eds) Physiological Plant Ecology, 1-24. Blackwell Science Ltd., Oxford.
- Pamplona, RR, Dingkuhn M, Ampong-Nyarko K, Moll CJ and DeDatta SK (1990) Paper at 21<sup>st</sup> Annual Convention of Pest Control of the Philippines, May 6-10, 1990, IRRI Library.
- Parsons AJ (1988) In: Jones MB and Lazenby A (eds) The Grass Crop, 129-177. Chapman and Hall Ltd., New York.
- Pearce RB, Carlson GE, Barnes DK, Hart RH, Hanson CH (1969) *Crop Science* **9**, 423-426.
- Peng S, Cassman KG, Kropff MJ (1995) *Crop Science* **35**, 1627-1630.
- Peng S, Cassman KG, Virmani SS, Sheehy JE, Khush GS (1999) *Crop Science* **39**, 1552-1559.
- Penning de Vries FWT, Brunsting AHM, Van Laar HH (1974) *Journal of Theoretical Biology* **45**, 339-377.
- Reynolds MP, van Ginkel M, Ribaut JM (2000) *Journal of Experimental Botany* **51**, 459-473.
- Robson MJ (1982) *Annals of Botany* **49**, 321-329.
- Ryle GJA, Cobby JM, Powell CE (1976) *Annals of Botany* **40**, 571-586.
- Sage RF (2000) In: Sheehy JE, Mitchell PL and Hardy B (eds) Redesigning Rice Photosynthesis to Increase Yield. International Rice Research Institute, Philippines.
- Satake T, Yoshida S (1978) *Japanese Journal of Crop Science* **47**, 6-17.
- Sharkey TD, Laporte MM, Kruger EL (2000) In: Sheehy JE, Mitchell PL and Hardy B (eds) Redesigning Rice Photosynthesis to Increase Yield. International Rice Research Institute, Philippines.
- Sheehy JE, Peacock JM (1975) *Journal of Experimental Botany* **26**, 679-691.
- Sheehy JE, Johnson IR (1988) In: Jones MB and Lazenby A (eds) The Grass Crop, 243-275. Chapman and Hall Ltd., London.
- Sheehy JE, McNeill A (1988) In: O'Gara F, Manian S and Drevon JJ (eds) Physiological Limitations and the Genetic Improvement of Symbiotic Nitrogen Fixation, 87-96. Kluwer Academic Publishers, London.
- Sheehy JE, Mitchell PL, Tsukaguchi T, Dionora J, Ferrer A, Torres R (1998) *Japanese Journal of Crop Science* **67**, extra issue 2.
- Sheehy JE (2000) In: Sheehy JE, Mitchell PL and Hardy B (eds) Redesigning Rice Photosynthesis to Increase Yield. International Rice Research Institute, Philippines.
- Sheehy JE (2001a) In: Nösberger J, Geiger HH and Struik PC (eds) Crop Science: Progress and Prospects. Proceedings of the 3<sup>rd</sup> International Crop Science Congress, Hamburg, Germany. CAB International (In Press).
- Sheehy JE, Dionora MJA, Mitchell PL (2001b) *Field Crops Research* **71**, 77-85.

- Sinclair TR, Horie T (1989) *Crop Science* **29**, 90-98.
- Sinclair T (1998) *Crop Science* **38**, 638-643.
- Sinclair TR, Sheehy JE (1999) *Science* **283**, 1456-1457.
- Smith H (2000) *Nature* **407**, 585-591.
- Steffen W, Noble I, Canadell J, Apps M, Schulze ED, Field CB, Ghazi A, Gifford R, Heimann M, Houghton R, Kabat P, Körner C, Lambin E, Linder S, Mooney HA, Murdiyarso D, Post WM, Prentice C, Raupach MR, Schimel DS, Shvidenko A and Valentini R (1998) *Science* **280**, 1393-1394.
- Thomas H (1994a) In: Monteith, J. L., Scott, R.K. and Unsworth, M.H. (eds.) *Resource Capture by Crops*. Nottingham University Press, Leicestershire.
- Thomas JF (1994b) In: Boote KJ, Bennett JM, Sinclair TR and Paulsen GM (eds) *Physiology and Determination of Crop Yield*, 153-180. Madison, Wisconsin, USA.
- Thornley JHM, Johnson IA (1990) *Plant and Crop Modelling*. Oxford University Press, Oxford.
- Tilman D, Fargione J, Wolff B, D' Antonio C, Dobson A, Howarth R, Schindler D, Schlesinger WH, Simberloff D, Swackhamer D (2001) *Science* **292**, 281-284.
- Uemura K, Anwaruzzaman, Miyachi S and Yokota, A (1997) *Biochemical and Biophysical Research Communications* **2333**, 568-571.
- UNFPA (1999) *The State of World Population*, <http://www.unfpa.org/swp/1999.chapter2fig5.htm>, United Nations Population Fund, New York, USA.
- Unsworth MH, Colls JJ (1994) In: Monteith JL, Scott RK and Unsworth MH (eds) *Resource Capture by Crops*, 189-209. Nottingham University Press, Nottingham.
- von Caemmerer S (2000) *Biochemical models of leaf photosynthesis*, CSIRO Publishing, Australia.
- Wilson D (1975) *Annals of Applied Biology* **80**, 323-338.
- Witty JF, Minchin FR, Sheehy JE (1983) *Journal of Experimental Botany* **34**, 951.
- Woodward FI (1999) In: Press MC, Scholes JD and Barker MG (eds) *Physiological Plant Ecology*, 443-455. Blackwell Science Ltd., Oxford.
- Yoshida S (1973) *Soil Science and Plant Nutrition* **19**, 311-316.
- Ziska LH, Namuco O, Moya T, Quilang J (1997) *Agronomy Journal* **89**, 45-53.