Susceptibility of six tomato cultivars to the root-knot nematode, *Meloidogyne incognita*

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ABSTRACT

Six tomato cultivars Moneymaker, Beefsteak, Roma, Summertaste, Mini Roma and Small fry were tested for their susceptibility to root-knot nematodes at inoculum levels of 200, 400, 600 Juveniles (J2) per pot. All were found to be susceptible to varying degrees as egg masses were present in all with Moneymaker and Roma being the most susceptible and Mini Roma, the least susceptible. The inoculum levels had a significant effect (p<0.05) on the number of galls and plant weights. The gall numbers and plant weights was negatively correlated, with the highest gall numbers and lowest plant weights recorded at the highest inoculum level in all cultivars except in Mini Roma in which there was little variation in gall numbers and plant weights.

Keywords: Lycopersicon esculentum, Resistance, inoculum level

1 INTRODUCTION

Root-knot nematodes infect a wide range of important crop plants and are particularly damaging to vegetable crops in tropical and subtropical countries (Sikora and Fernandez 2005). There are more than 90 described species in the genus *Meloidogyne* but the four most commonly occurring species are *Meloidogyne incognita*, *M. arenaria*, *M. javanica* and *M. hapla* (Sasser and Taylor 1978; Karssen 2000; Hunt et al. 2005). The short life cycle of 6 to 8 weeks enables root-knot nematode populations to survive well in the presence of a suitable host and their populations build up to a maximum usually as crops reach maturity (Shurtleff and Averre 2000). Host plants have varying degrees of susceptibility with some plants being highly susceptible while others are less susceptible or resistant to root-knot nematodes. The highly susceptible host plants allow the juveniles to enter the roots, reach maturity and produce many eggs while the resistant plants suppress their development and thus do not allow reproduction (Sasser and Taylor 1978; Karssen and Moens 2006). In case of severe infections and lack of appropriate control measures the yield loss is high and in some cases the plants die even before reaching maturity.

Root-knot nematodes (RKN) are one of the major pathogens of tomatoes worldwide and limit fruit production (Sikora and Fernandez 2005). Plant resistance is one of the most environmentally safe and economically viable means of controlling RKN. The RKN resistant crop varieties have comparatively better crop yield than the infected susceptible crop varieties. The resistant varieties can be used as part of integrated pest management in combination with other methods such as use of chemical nematicides, organic soil amendments, heat treatment, soil solarization, and crop rotation with non hosts for controlling RKN. Several cultivars of tomatoes such as Montelle, Sun6082, Pik Red, Celebrity, Baja, Betterboy and Beefmaster have been developed in an attempt to produce RKN resistant cultivars (Milligan et al. 1998; Tisserat 2006). Tomato cultivars have varying degrees of resistance to RKN and difference in quality and quantity of fruit production. The *Mi* gene originally found in wild tomato species *Lycopersicon peruvianum* is one of the best characterized nematode resistance genes and has been genetically engineered into many commercial tomato varieties (Nono-womdim et al. 2002; Abad et al. 2003). The resistance mechanism in response to invasion by RKN involves the formation of necrotic cells at the infection site to prevent the juveniles from developing any further. However a high level of genetic variability of RKN has led to the existence of races and virulent populations which can reproduce even on plants carrying the resistance genes (Castagnone-Sereno 2006). The RKN population density also affects the yield loss and tolerance levels of the different tomato cultivars. This investigation is an attempt to identify the natural resistance of six tomato cultivars commonly available in Fiji, namely, Moneymaker, Beefsteak, Roma, Summertaste, Miniroma and Small fry, to the RKN *M. incognita* and determine the effects of varying population levels on host plant health.
Table 1. Susceptibility of tomato cultivars to varying RKN inoculum levels

<table>
<thead>
<tr>
<th>Cultivar</th>
<th>Inoculum level</th>
<th>Av. Gall No.</th>
<th>Av. Egg Mass</th>
<th>Av. Plant Weight</th>
<th>Av. % Wt Reduction compared to control</th>
</tr>
</thead>
<tbody>
<tr>
<td>Moneymaker</td>
<td>control</td>
<td>0</td>
<td>0</td>
<td>7.46</td>
<td>0</td>
</tr>
<tr>
<td></td>
<td>200 J2</td>
<td>16</td>
<td>10</td>
<td>6.05</td>
<td>18.9</td>
</tr>
<tr>
<td></td>
<td>400 J2</td>
<td>21</td>
<td>12</td>
<td>5.99</td>
<td>19.71</td>
</tr>
<tr>
<td></td>
<td>600 J2</td>
<td>26</td>
<td>12</td>
<td>4.81</td>
<td>35.52</td>
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<tr>
<td>Roma</td>
<td>control</td>
<td>0</td>
<td>0</td>
<td>8.67</td>
<td>0</td>
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<tr>
<td></td>
<td>200 J2</td>
<td>13</td>
<td>6</td>
<td>7.4</td>
<td>14.65</td>
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<tr>
<td></td>
<td>400 J2</td>
<td>19</td>
<td>11</td>
<td>6.66</td>
<td>23.18</td>
</tr>
<tr>
<td></td>
<td>600 J2</td>
<td>26</td>
<td>17</td>
<td>5.92</td>
<td>31.72</td>
</tr>
<tr>
<td>Summertaste</td>
<td>control</td>
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<td>0</td>
<td>7.79</td>
<td>0</td>
</tr>
<tr>
<td></td>
<td>200 J2</td>
<td>9</td>
<td>3</td>
<td>2.73</td>
<td>19.94</td>
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<tr>
<td></td>
<td>400 J2</td>
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<td>3</td>
<td>2.3</td>
<td>32.55</td>
</tr>
<tr>
<td></td>
<td>600 J2</td>
<td>22</td>
<td>8</td>
<td>2.18</td>
<td>36.07</td>
</tr>
<tr>
<td>Small Fry</td>
<td>control</td>
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<td>0</td>
<td>8.65</td>
<td>0</td>
</tr>
<tr>
<td></td>
<td>200 J2</td>
<td>11</td>
<td>5</td>
<td>7.79</td>
<td>9.94</td>
</tr>
<tr>
<td></td>
<td>400 J2</td>
<td>15</td>
<td>9</td>
<td>6.99</td>
<td>19.19</td>
</tr>
<tr>
<td></td>
<td>600 J2</td>
<td>21</td>
<td>12</td>
<td>5.9</td>
<td>31.79</td>
</tr>
<tr>
<td>Beefsteak</td>
<td>control</td>
<td>0</td>
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<td>3.09</td>
<td>0</td>
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<tr>
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<td>7</td>
<td>2.85</td>
<td>7.77</td>
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<td>15</td>
<td>8</td>
<td>2.31</td>
<td>25.24</td>
</tr>
<tr>
<td></td>
<td>600 J2</td>
<td>20</td>
<td>11</td>
<td>2.01</td>
<td>34.95</td>
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<tr>
<td>Mini Roma</td>
<td>control</td>
<td>0</td>
<td>0</td>
<td>2.56</td>
<td>0</td>
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<tr>
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<td>200 J2</td>
<td>5</td>
<td>1</td>
<td>2.33</td>
<td>8.98</td>
</tr>
<tr>
<td></td>
<td>400 J2</td>
<td>6</td>
<td>1</td>
<td>2.37</td>
<td>7.42</td>
</tr>
<tr>
<td></td>
<td>600 J2</td>
<td>6</td>
<td>2</td>
<td>2.34</td>
<td>8.59</td>
</tr>
</tbody>
</table>

2 METHODS

The root-knot nematode *Meloidogyne incognita* (Kofoid and White) was used for the experiments. The RKN culture was initiated from single egg masses and propagated on tomato cultivar, Moneymaker in the glass house at The University of the South Pacific. Egg masses from the infected host plant were hand picked and placed in distilled water kept in a BOD incubator at 28 ± 1°C to obtain the second stage juveniles (J2). The juvenile suspension was standardised to final concentration of 100 juveniles per millilitre of distilled water.

Experiments were conducted in 2 litre pots each containing 900 grams of autoclaved soil. A three week old tomato seedling was planted into each of the pots for each of the six cultivars: Moneymaker, Beefsteak, Roma, Summertaste, Mini Roma and Smallfry. The J2 inoculum was added 1 day after transplanting at the rate of 200, 400 and 600 J2 per pot using 1 ml micropipette into two wells near the roots. Four replicates were kept for each of the inoculum levels including a control without any inoculation (0 J2). The pots were arranged in a random block design and kept in the glass house and watered once every second day. The experiments ran for 8 weeks starting from the second week of December 2006. All the varieties are recommended to be grown in the summer period by the supplier.

The plants were carefully dug out at the end of the eighth week and gently washed to remove soil from the roots. The plant roots were examined under a stereoscopic microscope (Olympus Model SZ 50) and the number of galls and egg masses present were recorded. The fresh plant weights (shoot plus roots) were also recorded for each of the plant. Average values are recorded in Table 1. The results were subjected to Univariate ANOVA and trends plotted using SPSS software Version 13.0 (Figures 1, 2 and 3).

3 RESULTS

All the six tomato cultivars were susceptible to root-knot nematodes to varying degrees. Egg masses were recorded in all six cultivars thus indicating that none of the tested cultivars was resistant to RKN. A direct relationship was observed between the gall numbers and the inoculation level for all cultivars except for Mini Roma which had relatively little variation in gall number and plant weight with increasing inoculum levels (Figure 1). The plant weights in all cultivars except Mini Roma decreased as compared to the controls with increasing inoculum levels (Figure 2). The greatest percentage reduction in plant weight compared to the control was observed for the cultivar Summertaste while Mini Roma had the least variation in the percentage plant weight loss (Figure 3). Mini Roma was found to be the least susceptible cultivar while Moneymaker and Roma were found to be the most susceptible cultivar having the highest gall numbers. The inoculum level was found to have significant effect (p<0.05) on the number of galls and plant weight.
4 DISCUSSION

The susceptibility of a plant to RKN depends on the ability of RKN juveniles to penetrate the roots of the plant and cause the formation of giant cells which appears as knots (galls) on the roots (Chen and Dickson 2004). The juveniles feed and moult twice before developing into the adult stage (Siddiqi 2000). The adult female RKN stays inside the giant cells and continues to feed and produces egg mass in a gelatinous matrix protruding out of the root gall. The egg masses give rise to infective juveniles (J2) which may infect other uninfected roots of the same plant or migrate and infect the nearby plants. In case of a plant resistant to RKN, the juveniles are either unable to penetrate the roots, or die after penetration or are unable to complete their development, or females are unable to reproduce. The Mi gene confers resistance by localized tissue necrosis around the region where the juveniles penetrate, thus juveniles are unable to establish feeding sites resulting in their death or migration out of the roots (Milligan et al. 1998; López-Pérez et al. 2006).

The presence of root galls along with egg mass on all six varieties of tomato plants indicates that none of the varieties is resistant to root-knot nematodes. However significant differences in the number of galls present on each of the six varieties indicate different levels of susceptibility. The level of susceptibility is controlled by the presence of resistance genes such as Mi gene and genetic background of the tomato cultivar (Castagnone-Sereno 2006; Jacquet et al. 2005). The homozygous or heterozygous state of the Mi locus has been found to affect
the degree of resistance to RKN, with the cultivars having the heterozygous form of the Mi gene being more susceptible than the homozygous cultivars (Jacquet et al. 2005). The variation in the susceptibility to RKN in the six tomato cultivars screened is likely to be due to the genetic differences between the cultivars and thus explains the variation in gall numbers and egg masses. Moneymaker and Roma were found to be the most susceptible as greatest number of juveniles penetrated and completed their development to maturity as shown by the high gall numbers and egg masses present. Mini Roma was the least susceptible variety as only a limited number of juveniles were able to penetrate, develop to maturity and lay egg masses.

Increasing inoculum levels led to an increase in the number of galls with greater reduction in plant weight for all varieties except for Mini Roma. This shows that when the inoculum levels are high, greater number of juveniles are able to infect the plant roots which results in reduced nutrient and water uptake by the roots and consequently poor plant growth (Karssen and Moens 2006). In Mini Roma, even higher inoculum (600 J2) could not establish a larger population, indicating the presence of some genetic resistance and consequently insignificant decrease in plant weight with increasing inoculum levels. Further trials with higher inoculum levels or planting in heavily infested soil would give a more conclusive result about the susceptibility of each cultivar and the effect of RKN on yield quantity and quality.

Figure 3. Percentage plant weight loss vs. inoculum level

<table>
<thead>
<tr>
<th>Variety</th>
<th>Inoculum Level</th>
</tr>
</thead>
<tbody>
<tr>
<td>1. Moneymaker</td>
<td>600 J2</td>
</tr>
<tr>
<td>2. Roma</td>
<td>600 J2</td>
</tr>
<tr>
<td>3. Summertaste</td>
<td>600 J2</td>
</tr>
<tr>
<td>4. Smallfry</td>
<td>600 J2</td>
</tr>
<tr>
<td>5. Beefsteak</td>
<td>600 J2</td>
</tr>
<tr>
<td>6. Mini Roma</td>
<td>600 J2</td>
</tr>
</tbody>
</table>

The variability in pathogenicity is also dependent on the genetic variability of RKN population and species composition but since the populations used for inoculation were raised from a single egg mass under the same conditions and host plants it is likely to have had minimal effect in this investigation. There is sophisticated interaction between the host plant and root-knot nematodes and a number of studies have found resistance breaking pathotypes of RKN that are able to parasitize even RKN resistant plants (Jacquet et al. 2005; Abad et al. 2003; Baicheva et al. 2002) which is a major limiting factor in using plant resistance as a means for controlling root-knot nematodes. However, identification and use of RKN resistant and tolerant varieties can still be a viable means of minimising loss caused by RKN. Another factor which needs to be taken into consideration in any further investigation is the quality and quantity of fruit production of the resistant, less susceptible and highly susceptible varieties because at times the resistant varieties do not produce fruit with the desirable taste and quality (Lo´pez-Pe´rez et al. 2006). Grafting of desired varieties on the roots of the less susceptible and resistant varieties is an alternative but requires technical knowledge and has additional costs associated with getting the grafted plants to the farmers. The susceptibility of the different tomato varieties has important implications on the yield and economic returns thus information on susceptibility to RKN can be useful to farmers while selecting the variety for planting on RKN infested fields.
REFERENCES
Tisserat, N. 2006. Root knot nematode of tomato. Fact sheets tomato. Extension plant pathology –Kansas State University, Manhattan.

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