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Fertility control to mitigate human–wildlife conflicts: a review

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Abstract. As human populations grow, conflicts with wildlife increase. Concurrently, concerns about the welfare, safety and environmental impacts of conventional lethal methods of wildlife management restrict the options available for conflict mitigation. In parallel, there is increasing interest in using fertility control to manage wildlife. The present review aimed at analysing trends in research on fertility control for wildlife, illustrating developments in fertility-control technologies and delivery methods of fertility-control agents, summarising the conclusions of empirical and theoretical studies of fertility control applied at the population level and offering criteria to guide decisions regarding the suitability of fertility control to mitigate human-wildlife conflicts. The review highlighted a growing interest in fertility control for wildlife, underpinned by increasing numbers of scientific studies. Most current practical applications of fertility control for wild mammals use injectable single-dose immunocontraceptive vaccines mainly aimed at sterilising females, although many of these vaccines are not yet commercially available. One oral avian contraceptive, nicarbazin, is commercially available in some countries. Potential new methods of remote contraceptive delivery include bacterial ghosts, virus-like particles and genetically modified transmissible and non-transmissible organisms, although none of these have yet progressed to field testing. In parallel, new species-specific delivery systems have been developed. The results of population-level studies of fertility control indicated that this approach may increase survival and affect social and spatial behaviour of treated animals, although the effects are species- and context-specific. The present studies suggested that a substantial initial effort is generally required to reduce population growth if fertility control is the sole wildlife management method. However, several empirical and field studies have demonstrated that fertility control, particularly of isolated populations, can be successfully used to limit population growth and reduce human-wildlife conflicts. In parallel, there is growing recognition of the possible synergy between fertility control and disease vaccination to optimise the maintenance of herd immunity in the management of wildlife diseases. The review provides a decision tree that can be used to determine whether fertility control should be employed to resolve specific human-wildlife conflicts. These criteria encompass public consultation, considerations about animal welfare and feasibility, evaluation of population responses, costs and sustainability.

Additional keywords: contraception, fertility inhibitor, immunocontraception, population control, wildlife management.

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Introduction

Current trends of human population growth and landscape development show that human–wildlife conflicts are increasing (Rutberg and Naugle 2008; White and Ward 2010; Gionfriddo *et al.* 2011*a*). Many of these conflicts have been traditionally managed by lethal methods. However, opposition to culling has become widespread because of concerns about welfare, human safety in urban settings and environmental impact (e.g. Beringer *et al.* 2002; Cowan and Quy 2003; Sharp and Saunders 2008; McLeod and Saunders 2014). This growing antipathy toward lethal methods places increasing constraints on wildlife management options, particularly for high-profile, iconic species (Barr *et al.* 2002; Poiani *et al.* 2002; Druce *et al.* 2011). Consequently, there has been growing interest in non-lethal

methods such as translocation and fertility control (Duka and Masters 2005; Barfield *et al.* 2006; Fagerstone *et al.* 2010).

Translocation of problem wildlife may cause stress and increase mortality, it is relatively expensive and has the potential to spread diseases and pathogens (e.g. Daszak *et al.* 2000; Massei *et al.* 2010*a*). Conversely, fertility control is increasingly advocated as a safe, humane and effective means of managing overabundant wildlife (Fagerstone *et al.* 2010; Kirkpatrick *et al.* 2011; McLaughlin and Aitken 2011). The potential market for human contraceptives and a growing public interest in alternatives to surgical sterilisation for companion animals and livestock have fostered investment in the development of novel fertility-control agents (Herbert and Trigg 2005; Naz *et al.* 2005; Massei *et al.* 2010*b*).

Early fertility-control agents lacked species-specificity, induced only transitory sterility, thus requiring repeated application, or had a limited window between the dose required to achieve sterility and the toxic or lethal dose. Other obstacles included manufacturing costs, concerns that residues might enter the human food chain and welfare issues regarding side effects (Gray and Cameron 2010; Kirkpatrick *et al.* 2011).

Several reviews on animal fertility control have been published in recent years. With the exception of the overview by Fagerstone *et al.* (2010) on issues concerning the use of reproductive inhibitors for wildlife in North America, these reviews have focussed on specific groups such as zoo species and companion animals (Asa and Porton 2005; Munson 2006; Purswell and Kolster 2006; Levy 2011; Massei and Miller 2013), on particular compounds such as immunocontraceptives (Cooper and Larsen 2006; Kirkpatrick *et al.* 2011), on selected species such as brushtail possums (*Trichosurus vulpecula*) (Ji 2009; Cross *et al.* 2011) or on groups of species such as ungulates (Patton *et al.* 2007). Here, we provide a comprehensive, critical overview of fertility control to mitigate human–wildlife conflicts, with the following aims:

- (1) to analyse trends in research on fertility control for wildlife;
- (2) to review recent developments in fertility-control technologies;
- (3) to summarise delivery methods of fertility-control agents for wildlife;
- (4) to provide a synthesis of the conclusions of empirical and theoretical studies of fertility control applied at the population level; and
- (5) to offer a framework of criteria to guide decisions regarding the suitability of fertility control to mitigate human–wildlife conflicts.

Throughout the review, 'fertility inhibitors' or 'fertilitycontrol agents' are used as a generic term to include chemicals used to block conception, or to prevent ovulation and sperm production, or that interfere with oogenesis and spermatogenesis.

Trends in research on fertility control for wildlife

We explored recent trends in wildlife fertility-control research since 1982 by searching five databases BIOSIS, CAB Abstracts, Web of Science, Zoological Records and Medline for the following keywords in the title or the abstract: immunocontraception/immunocontraceptive', 'fertility control', 'fertility inhibition/inhibitor', 'reproductive inhibition/inhibitor', 'contraception/contraceptive' and 'sterilisation'. All publications concerning empirical and theoretical studies of fertility control on wildlife species were included. Papers on laboratory animals, zoo animals and livestock were included only if they made specific references to potential wildlife applications. These searches generated data on (1) number of papers published per year, (2) type of study, including (i) laboratory and captive studies, (ii) field studies on free-living wildlife, (iii) reviews and (iv) theoretical studies based on modelling, and (3) number of gender-specific applications of fertility inhibitors, i.e. females only, males only or both.

In total, 460 papers were published between 1982 and 2010; the number of studies grew from 1-4 per year in the 1980s to an average of 27.3 per year in the past decade, with occasional peaks in numbers being due to special issues dedicated to this subject (Fig. 1). Overall, field studies followed laboratory ones with a noticeable lag of 5-6 years. Modelling studies progressively increased, whereas the number of reviews reached an asymptote with an average of 6.6 per year in the past decade. In total, 51% of papers focussed on immunocontraception, 31% on other fertility inhibitors and 18% on combinations of fertility inhibitors or on generic contraceptives. Of the 305 reported empirical and theoretical studies, 78% (n=238) were on females, 7.5% (n=23) on males and 14.5% (n=44) on both genders. The bias toward female contraception is due to (1) studies focussed on human fertility (Barfield et al. 2006), (2) the recognition that polygyny and polyandry are common among many mammal species (e.g. Garrott and Siniff 1992; Kennis et al. 2008; Huchard et al. 2012) and, thus, extremely high levels of male sterility would be required to have any effect

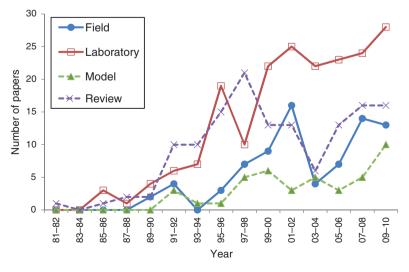


Fig. 1. Number of papers published every 2 years on fertility control for wildlife.

at the population level and (3) models demonstrating that effective control at the population level could be achieved only by rendering infertile a high proportion of females (e.g. Caughley *et al.* 1992; Hobbs *et al.* 2000; Merrill *et al.* 2006).

Fertility inhibitors for wildlife

A wildlife fertility-control agent suitable for field applications should have the following characteristics (Kirkpatrick and Turner 1991; Massei 2012; Massei and Miller 2013):

- nil or acceptable side-effects on animal physiology, welfare and behaviour,
- (2) effective when administered in a single dose,
- (3) render all or the majority of animals infertile for the duration of their potential reproductive life,
- (4) inhibit female reproduction, but ideally prevent reproduction in both sexes,
- (5) not compromise welfare by interfering with pre-existing pregnancy or lactation,
- (6) relatively inexpensive to produce and deliver,
- no bioactive residues entering any food chain associated with treated animals,
- (8) administrable through remote delivery,
- (9) species-specific, and
- (10) stable under a wide range of field conditions.

None of the fertility-control agents currently available meets all the above characteristics; however, several exhibit many of these features. The following review includes fertility inhibitors that (1) are commercially available or have been evaluated in several species, with particular focus on those used in wildlife, (2) can induce infertility for at least a year or for at least one reproductive season and (3) are primarily aimed at females, because this gender should remain the primary target for fertility control. Some examples are discussed of contraceptives that are very effective for other animals, but that cannot be regarded as suitable for wildlife.

Hormonal methods

Synthetic hormones, widely used in zoo animals and livestock, bind to endogenous hormone receptors and disrupt folliculogenesis, ovulation and egg implantation in females and impair spermatogenesis in males (Asa and Porton 2005). Those tested in wildlife include norgestomet, melengestrol acetate, levonorgestrel and quinestrol.

Norgestomet implants, used to suppress oestrus in beef cattle, inhibited reproduction in female white-tailed deer (*Odocoileus virginianus*) and black-tailed deer (*Odocoileus hemionus*) individuals for at least 1 year (Jacobsen *et al.* 1995; DeNicola *et al.* 1997).

Melengestrol acetate (MGA) with an estimated duration of efficacy of ≥ 2 years has been employed in zoos for wildlife contraception for ~20 years. MGA implants are effective on ungulates, carnivores and primates (Plotka and Seal 1989; Wood *et al.* 2001; Asa and Porton 2005; Hall-Woods *et al.* 2007). However, MGA causes uterine pathology in captive coati (*Nasua nasua*; Chittick *et al.* 2001), felids and canids (Munson 2006; Moresco *et al.* 2009) and a higher incidence of

stillbirth and infant mortality in golden lion tamarins (*Leontopithecus rosalia*; Wood *et al.* 2001).

Levonorgestrel is the active component of a multi-year implant contraceptive originally approved by the US Food and Drug Administration (FDA) for human contraception. Because of side effects such as migraine and weight changes, the implant was withdrawn from the human-contraception market in some countries (Benfield and Darney 2011). A single administration of levonorgestrel implants inhibits reproduction in wildlife species for several years, without apparent adverse side effects (Nave *et al.* 2002*a*; Middleton *et al.* 2003; Coulson *et al.* 2008; Wheaton *et al.* 2011). In addition, levonorgestrel and quinestrol have been successfully used as contraceptives for rodents such as plateau pikas (*Ochotona curzioniae*) and Mongolian gerbils (*Meriones unguiculatus*) (e.g. Liu *et al.* 2012; Fu *et al.* 2013).

Regardless of proven efficacy, the use of hormonal methods on free-ranging wildlife is still debated because of potential welfare effects of long-term exposure, environmental impact and possible transfer of steroids via food chains (Nettles 1997; DeNicola *et al.* 2000; Asa and Porton 2005).

Gonadotropin-releasing hormone (GnRH) agonists are proteins that mimic GnRH and stimulate production and release of follicle-stimulating hormone (FSH) and luteinising hormome (LH). Administration initially causes the 'flare up' effect, i.e. stimulates oestrus in females and temporarily enhances testosterone and semen production in males (Patton *et al.* 2007). Because agonists do not quickly dissociate from the GnRH receptors, the 'flare up' is followed by prolonged ovarian quiescence and infertility (Gobello 2007).

Sustained-release subcutaneous implants of GnRH agonists, such as deslorelin (Suprelorin, Virbac, Milperra, NSW, Australia), have been used to inhibit reproduction for 1-2 years in cattle and in marsupials, including tammar wallabies (Macropus eugenii), grey kangaroos (Macropus giganteus) and brushtail possums (D'Occhio et al. 2002; Herbert et al. 2005; Eymann et al. 2007). In urban brushtail possums, deslorelin implants inhibited reproduction in 80% of the females treated (Lohr et al. 2009). Deslorelin has also been shown to be effective in cats, other felids and wild dogs (Herbert and Trigg 2005; Munson 2006; Bertschinger et al. 2008). Another GnRH agonist, leuprolide, found effective in suppressing reproduction for one breeding season in wapiti (Cervus elaphus; Baker et al. 2002; Conner et al. 2007) and female mule deer (Odocoileus hemionus; Baker et al. 2004), has not been used more recently. The effectiveness of GnRH agonists depends on agonist type, release system, dose rate and duration of treatment (Gobello 2007; Patton et al. 2007). The side effects of GnRH agonists are similar to those associated with gonad removal, but are reversible and there are no known effects on lactation (Asa and Porton 2005). Because GnRH agonists can cause abortion, they should be used outside the breeding season (Asa and Porton 2005).

Immunocontraceptive vaccines

Most recent studies of fertility control for wildlife have focussed on immunocontraceptive vaccines. Immunocontraception is achieved by exposing an animal to an antigen that stimulates the animal's immune system to produce antibodies to proteins or hormones essential for reproduction (Miller and Killian 2002). As a result, immunocontraceptives can prevent ovulation, sperm production or fertilisation. Adjuvants, which are inorganic or organic chemicals, macromolecules or entire cells of specific killed bacteria, are typically used to amplify the immune response to an antigen. The factors that affect effectiveness, longevity and side effects of immunocontraceptive vaccines include species, gender, age, individual variation in immunocompetence, as well as the active immunogen, formulation, delivery system and dose and type of adjuvant (Miller *et al.* 2008*a*, 2009; Holland *et al.* 2009; Kirkpatrick *et al.* 2011; Ransom *et al.* 2011). The most studied immunocontraceptives in wildlife are zona pellucida (ZP) and GnRH vaccines (Table 1).

The ZP is a layer of glycoproteins that surrounds an ovulated egg and allows species-specific sperm recognition and binding. There are four major ZP glycoproteins, named ZP1, ZP2, ZP3 and ZP4, each with different functions in the oocyte-sperm binding process and with varying degrees of homology among mammalian species (e.g. Kitchener et al. 2009; Gupta and Bhandari 2011). These differences are partly responsible for the variable results obtained when using a particular ZP vaccine on different species and have been exploited to make ZP-based vaccines more specific (Kitchener et al. 2009; Gupta et al. 2011; Levy 2011). Porcine ZP (PZP) immunocontraceptive vaccines, derived from ZP isolated from pig ovaries, have been effective in many ungulate species, monkeys, seals, bears and marsupials, but not in rodents, cats, dogs and wild pigs (Eade et al. 2009; Kitchener et al. 2009; Kirkpatrick et al. 2009, 2011; McLaughlin and Aitken 2011; Table 1). However, recently formulated recombinant PZP3 and PZP4 vaccines, delivered in three injectable doses, caused infertility in up to 89% mice, depending on the formulation type (Gupta et al. 2013).

Early formulations of ZP vaccines were delivered as a primer shot, followed by a booster, which placed major constraints on field applications with wildlife. Initial vaccine formulations also used Freund's complete adjuvant (FCA), which raised safety concerns regarding the occurrence of false-positive tuberculosis skin tests in deer treated with vaccines containing FCA, severe injection-site reactions and potential carcinogenicity for consumers of treated animals (Kirkpatrick et al. 2011). The development of a novel, safe and effective adjuvant (AdjuVac, National Wildlife Research Center, Fort Collins, CO, USA) combined with PZP-based vaccine succeeded in rendering animals of several species infertile for several years after a single dose (Table 1). Injectable formulations of PZP vaccines, such as the proprietary liposome-containing product SpayVac (ImmunoVaccine Technologies, Inc., Halifax, NS, Canada), with controlled-release properties, have been developed that generate responses for multiple years following a single administration (Brown et al. 1997; Turner et al. 2008; Rutberg et al. 2013). Modified FCA has also been used as a safe, effective substitute for FCA (Lyda et al. 2005). Recent studies have also shown that intra-nasal delivery of four doses of mouse ZP3 result in a significant reduction of reproductive output in mice (Ma et al. 2012; Kadir et al. 2013). In parallel, several newer alternative reagents, such as purified and/or receptor-specific adjuvants (e.g. monophosphoryl lipid A, ISCOMsm CpG oligonucleotides) are being investigated for either mucosal or parenteral route of vaccine administration (Sharma and Hinds 2012).

Possible negative effects of ZP vaccines include speciesspecific ovarian pathology and multiple infertile oestrous cycles (in polyoestrous species), leading to extended breeding season, increased movements, potential late births and disruption of social hierarchy (Miller *et al.* 2000; Curtis *et al.* 2007; Kirkpatrick *et al.* 2009, 2011; Nuñez *et al.* 2009, 2010). Other studies on white-tailed deer and feral horses have reported that treatment with ZP vaccines does not affect time budget, social behaviour and body condition (Miller *et al.* 2001; Hernandez *et al.* 2006; Ransom *et al.* 2010). The incidence of ovarian pathologies was significantly reduced when purified PZP proteins were used in vaccine constructs (Gupta *et al.* 2013).

PZP was found safe to administer to pregnant or lactating females (Turner et al. 1996; Kirkpatrick and Turner 2002; Perdok et al. 2007; Delsink and Kirkpatrick 2012). Differences in the results of studies using ZP-based vaccines may reflect different formulations of native, purified or recombinant ZP vaccines, or different adjuvants and methods of extraction of PZP from pig ovaries (Munson et al. 2005; Miller et al. 2009; Kirkpatrick et al. 2011). Injection-site reactions such as abscesses are rare (~1% in various species) in animals treated with ZP vaccines, whereas granulomas (thickened tissue filled with fluid) are more common at the injection site (Kirkpatrick et al. 2009; Gray et al. 2010). In 2012, a PZP-based vaccine ZonaStat-H was registered by The Humane Society of the United States and approved by the Environment Protection Agency (EPA) as a contraceptive for population control of feral horses and feral donkeys. ZonaStat-H is not commercially available, but can be obtained from The Science and Conservation Center ZooMontana.

GnRH-based vaccines cause infertility by generating antibodies toward GnRH, thus disrupting the downstream release of hormones that stimulate ovulation and sperm production. Multi-dose GnRH-based immunocontraceptive vaccines, currently used in livestock and companion animals, are unsuitable for wildlife (reviewed in Naz et al. 2005; McLaughlin and Aitken 2011), primarily because of the impracticality of recapturing individuals to administer multiple doses. One single-dose GnRH vaccine that has seen rapid developments in wildlife applications is GonaCon (National Wildlife Research Center, Fort Collins, CO, USA), registered in the US as a contraceptive for white-tailed deer, feral horses and feral donkeys. GonaCon consists of a synthetic GnRH coupled to a mollusk protein (Miller et al. 2008a). Formulated as an injectable, single-dose immunocontraceptive, GonaCon induced infertility for several years in deer, wild boar, pigs, cats, horses and bison (Bison bison) (e.g. Miller et al. 2000; Killian et al. 2008; Massei et al. 2008, 2012; Gray et al. 2010) (Table 1). As GonaCon prevents ovulation, treated females and males do not exhibit oestrous behaviour; however, male deer showed abnormal antler development (Fagerstone et al. 2008). In the years after treatment, GnRH antibody titres decrease and fertility may be restored (Miller et al. 2008a; Massei et al. 2012). In some studies, reproductive behaviour has been observed 1-2 years before fertility returned (Killian et al. 2008).

In some species, vaccination with GonaCon causes a granuloma or a sterile abscess at the injection site. Two years after vaccination with GonaCon, 5 of 15 treated female cats had a palpable non-painful injection-site granuloma (Levy *et al.* 2011). In white-tailed deer, injection-site lesions (granulomatous

Table 1. Captive and field trials conducted with different formulations of single-dose immunocontraceptive porcine zona pellucida (PZP)- and gonadotropin-releasing hormone (GnRH)-based vaccines on females of wildlife, feral and companion animal species

The effectiveness of these vaccines to cause infertility is expressed as proportion of infertile females in the control (C) and treatment (T) groups. The different percentages listed for each study are the responses recorded for successive years after first treatment

| Species | Ν | Type of study | Antigen | Adjuvant | % infertile females | Reference |
|---|----------------------------|---------------|--|--------------------|--|----------------------------|
| White-tailed deer Odocoileus virginianus | 5 per group | Captive | GonaCon, various formulations | AdjuVac | GonaCon-KLH = 100% 60% 50% 50% 25% GonaCon-B = 100% 100% 80% 80% 80% | Miller et al. 2008a |
| White-tailed deer | T = 24 C = 13 | Field | GonaCon-KLH | AdjuVac | $T = 67\% \ 43\%$ C = 8% 17% | Gionfriddo et al. 2011a |
| White-tailed deer | T = 26 C = 13 | Field | GonaCon–KLH | AdjuVac | T = 88% 47% C = 15% 0% | Gionfriddo et al. 2009 |
| Feral horse Equus caballus | T = 15 $C = 8$ | Captive | GonaCon-KLH | AdjuVac | T = 93% 64% 57% 43% C = 25% 25% 12% 0% | Killian et al. 2008 |
| Feral horse | T = 18 $C = 31$ | Field | GonaCon-B | AdjuVac | T=61% 58% 69% C=40% 31% 14% | Gray et al. 2010 |
| Elk Cervus elaphus | T = 10 $C = 8$ | Captive | GonaCon-B | AdjuVac | T = 90% 75% 50% 25% C = 0% 0% 0% 14% | Powers et al. 2011 |
| Elk | T = 10 T = 12 C = 15 | Captive | GonaCon–KLH | AdjuVac | GonaCon–KLH (1000 µg)=92% 90% 100% GonaCon–KLH (2000 µg)=90% 100% 100% C=27% 25% 0% | Killian et al. 2009 |
| Bison Bison bison | T = 6 C = 5 | Captive | GonaCon–KLH | AdjuVac | C = 27.025700.000 T = 100% C = 0% | Miller et al. 2004 |
| Wild boar Sus scrofa | T = 12 $C = 12$ | Captive | GonaCon–KLH | AdjuVac | T = 92% infertile for at least 4-6 years C = 0% | Massei et al. 2008, 2012 |
| Feral pig Sus scrofa | T = 18 C = 3 | Captive | GonaCon-KLH | AdjuVac | T = 89% C = 0% | Killian et al. 2006 |
| Cat Felis catus | T = 15 C = 5 | Captive | GonaCon-KLH | AdjuVac | T = 93% 73% 53% 40% 27% C = 0% | Levy <i>et al.</i> 2011 |
| Fallow deer Dama dama | T = 19 C = 152 | Field | SpayVac | FCA | T = 100% 100% 100% C = 4% 3% 4% | Fraker et al. 2002 |
| White-tailed deer | T=5 per group C=84 | Captive | PZP and SpayVac, various formulations | AdjuVac or Alum | SpayVac–AdjuVac: 100% 100% 100 80% 80% IVT–PZP–AdjuVac: 100% 80% 80% 80% 80% SpayVac–Alum: 20% NWRC–PZP–AdjuVac (200 µg): 80% 0% NWRC–PZP–AdjuVac (500 µg): 100% 20% 20% 20% 0% C=0% | Miller <i>et al</i> . 2009 |
| White-tailed deer | T = 34 $C = 11$ | Field | SpayVac | AdjuVac | T = 100% 100% C = 22% | Locke et al. 2007 |
| White-tailed deer | T = 9 T = 11 C = 245 | Field | SpayVac various formulations | AdjuVac | T SpayVac aqueous: 100% 75% T SpayVac non-aqueous: 64% 75% C=22% | Rutberg et al. 2013 |
| White-tailed deer | T = 36 $C = 11$ | Field | PZP | AdjuVac | T = 100% C = 22% | Hernandez et al. 2006 |
| Feral horse | T = 12 $C = 8$ | Captive | SpayVac | AdjuVac | T = 100% 83% 83% 83% C = 25% 25% 12% 0% | Killian et al. 2008 |
| Feral horse | T = 17 $C = 21$ | Field | PZP | FCA and QS-21 | T = 95% 85% 68% 54% C = 46% 43% 49% 48% | Turner et al. 2007 |
| Feral horse | T = 14 $C = 31$ | Field | PZP | AdjuVac | T = 63% 50% 56% C = 40% 31% 14% | Gray et al. 2010 |

nodules and sterile abscesses) occurred in the deep hind-limb musculature of >85% of GonaCon-treated animals, although no evidence of limping or impaired mobility was observed in these animals (Gionfriddo *et al.* 2011*b*). GonaCon had no adverse effects on health of wild boar, white-tailed deer and prairie dogs (*Cynomys ludovicianus*; Massei *et al.* 2008, 2012; Yoder and Miller 2010; Gionfriddo *et al.* 2011*b*). In white-tailed deer, reactions at injection sites and in lymph nodes were typical responses to injection of adjuvanted vaccines formulated as water-in-oil emulsions (Miller *et al.* 2008*a*). GonaCon administered to 3–4-month-old white-tailed deer fawns did not induce contraception or prevent sexual development (Miller *et al.* 2008*b*) and, when given to pregnant bison, it did not affect pregnancy (Miller *et al.* 2004).

The gradual reversibility of the infertility effect, at least in a proportion of animals treated with ZP- and GnRH-based vaccines, is regarded as desirable in some species (Druce et al. 2011; Kirkpatrick et al. 2011). Because both vaccines are broken down when ingested, they do not enter the food chain and hence do not pose unacceptable risks to predators or human consumers even if the muscle injected with the immunocontraceptive is ingested. Both ZP and GnRH are inherently poorly immunogenic and thus must be formulated to elicit an immune response, for instance, by conjugation to larger carrier foreign proteins. More recently, recombinant technology has been used to produce antigens fused to carrier peptides. Recombinant injectable GnRH vaccines have caused a strong immune response in feral pigs (Kemp and Miller 2008; Campbell et al. 2010). Fusion protein technology has also been used to produce a plasmid-DNA vaccine encoding GnRH; injection with this vaccine caused a significant reduction in fertility in both male and female mice (Mus musculus; (Khan et al. 2008). To overcome the lack of availability of the purified native ZP glycoproteins obtained from ovaries of slaughtered pigs, porcine ZP3 and ZP4 were expressed in Escherichia coli; immunisation with these recombinant proteins significantly decreased fertility in laboratory mice and dogs (Gupta et al. 2011, 2013).

Both ZP and mammalian GnRH are highly conserved in structure and function across mammalian species (e.g. Cariño et al. 2002; Temple et al. 2003). Consequently, the development of species-specific immunocontraceptives based on ZP or GnRH will be challenging, although there is evidence of differential ZP3 specificity between marsupial and eutherian mammals (Duckworth et al. 2008). However, species-specific binding of sperm to ZP has potential for developing species-specific immunocontraceptives based on sperm-surface antigens (e.g. Moore et al. 1997; Grignard et al. 2007; Naz 2011). Recognition of sperm antigens that participate in sperm-ZP binding can be achieved using phage display techniques (Eidne et al. 2000; Naz 2005). This approach has since enabled identification of putatively pig-specific phage antigens that stimulate production of sperm-binding antibodies with potential for immunocontraception (Samoylova et al. 2012).

Other contraceptives

Several putative fertility inhibitors are still in the early phase of development. These include GnRH-toxin conjugates and cholesterol mimics. GnRH-toxin conjugates are formed by linking synthetic analogues of GnRH to cytotoxins. This enables selective targeting and mortality of cells secreting reproductive hormones, potentially leading to permanent sterility in both males and females. Because of their proteinaceous nature, these conjugates are broken down by digestion and thus do not enter the food chain. Examples include an injectable GnRH-toxin conjugate that suppressed the secretion of LH for up to 6 months in female mule deer (Baker et al. 1999) and an injectable GnRH-cytotoxin (pokeweed antiviral protein, PAP) conjugate that disrupted reproduction in adult male dogs, female rats (Rattus norvegicus) and sheep (Ovis aries) for at least 6 months (Nett et al. 2003; Ball et al. 2006). The cholesterol mimic DiazaCon can affect reproduction in birds and mammals because it inhibits production of cholesterol, which is a parent compound of male and female reproductive steroids (Fagerstone et al. 2010). Following ingestion of DiazaCon for 1-2 weeks, reproduction was suppressed for a few months in black-tailed prairie dogs, roseringed parakeets (Psittacula krameri) and monk parakeets (Myiopsitta monachus) (Nash et al. 2007; Yoder et al. 2007, 2011; Avery et al. 2008; Lambert et al. 2010). DiazaCon also reduced cholesterol in grey squirrels (Sciurus carolinensis), although the effects on reproduction were difficult to interpret because of poor breeding success in the control group (Mayle et al. 2013). DiazaCon has a relatively narrow contraceptive window before undesirable side effects on physiology and behaviour occur (Sachs and Wolfman 1965; Yoder et al. 2004, 2007). The efficacy of this compound depends on its bioaccumulation; however, its consequently relatively long elimination half-life poses potential exposure risk to predators and scavengers of treated animals. Therefore, DiazaCon seems more suited for applications to captive wildlife, seasonal breeders and localised populations experiencing little or no predation and where non-target species can be prevented from feeding on DiazaCon-treated baits (Avery et al. 2008; Fagerstone et al. 2010).

Nicarbazin (NCZ) is a bird-specific oral contraceptive widely used as a veterinary medicine to manage coccidiosis in broiler chickens. NCZ disrupts the membrane between the egg albumen and yolk, thus compromising embryo development (Jones *et al.* 1990). NCZ is registered in the USA for use with Canada geese (*Branta canadensis*; Bynum *et al.* 2007) and feral pigeons (*Columbia livia*; Fagerstone *et al.* 2008) and, in Italy, to control urban populations of feral pigeons (Ferri *et al.* 2009). Because NCZ is rapidly cleared from the body once consumption ceases, the effect on fertility is reversible and, thus, NCZ poses minimal risk to predators and scavengers of treated birds. The disadvantage is that NCZ must be fed continuously before and during egg-laying to be effective (Fagerstone *et al.* 2010). This may underlie the equivocal results reported for population-level effects in the field (Giunchi *et al.* 2007; Ferri *et al.* 2009).

Other methods currently being investigated target the mammalian ovary and aim at inducing early menopause and permanent sterility (Tran and Hinds 2013). The epoxide 4-vinylcyclohexene diepoxide (VCD) has ovarian-specific toxicity and follicle-depleting properties (Hoyer *et al.* 2001; Mayer *et al.* 2002). The administration of VCD by injection or ingestion repeatedly over a period of up to 30 days depletes the ovary of follicles leading to ovarian senescence (Mayer *et al.*

2004; Hu *et al.* 2006). Similarly, repeated oral administration of triptolide, a diterpenoid triepoxide, affects the ovarian function by causing follicular atresia (Xu and Zhao 2010; Liu *et al.* 2011). Triptolide can also compromise sperm function in males (Singla *et al.* 2013). However, reduced fertility induced by free-feeding of epoxides has yet to be demonstrated in either males or females.

Delivery methods

Ideally, a fertility-control agent should be species-specific. In practice, this is rarely the case at present and most contraceptives can affect a variety of wildlife species. Therefore, specificity must be achieved by the delivery method.

Fertility-control agents are delivered through the parenteral and oro-nasal route or via live organisms. Parenteral delivery includes direct injection (usually intramuscular), subcutaneous implants and remote delivery systems such as bio-bullets and syringe-darts.

Subcutaneous implants that release fertility control agents into an animal over a sustained period of time have been successfully employed to induce infertility for 1-5 years in a variety of wildlife species (e.g. Plotka and Seal 1989; Nave et al. 2002a, 2002b; Coulson et al. 2008; Lohr et al. 2009). Bio-bullets are biodegradable projectiles used to administer remotely various veterinary substances (DeNicola et al. 2000). Syringe-darts, routinely employed to anaesthetise wild animals, have also been used to administer contraceptives (Aune et al. 2002). Distance-adjustable CO2-powered dart rifles have been employed to fire 2–3-mL syringe-darts at ranges of \leq 40 m into the hindquarter of large mammals (Rudolph et al. 2000; Delsink et al. 2007; Kirkpatrick et al. 2009; Rutberg et al. 2013). Such delivery systems have several advantages (Kreeger 1997), including the following: (1) they target individual animals, so specificity is assured; (2) they can administer an individually tailored dose based on a bodyweight; (3) they can deliver solid (e.g. silastic implants), semi-solid or liquid formulations; and (4) they can be used for remote delivery, to avoid the welfare and economic costs of trapping. Potential disadvantages include identification of previously vaccinated individuals, dose regulation and incomplete intra-muscular injection (DeNicola et al. 1997, 2000; Aune et al. 2002). Remote parenteral delivery of contraceptives is regarded as suitable for small or isolated groups of animals, for instance, in urban parks (DeNicola et al. 2000), on islands (Kirkpatrick et al. 2009) or in fenced wildlife reserves (Delsink et al. 2007; Delsink and Kirkpatrick 2012).

In oral delivery of antigens, a fundamental issue is the relatively high threshold of the immune system in recognising the antigen as 'foreign' before an immune response is mounted (Cross *et al.* 2011). Consequently, responses to orally delivered antigens will typically be short-lived and such vaccines are likely to require repeated applications. Miller *et al.* (1999) demonstrated the feasibility of oral vaccination of deer using a live recombinant Bacillus Calmette–Guerin (BCG) vaccine as the immunological vector of a model antigen. Vehicles for potential oro-nasal delivery of immunocontraceptives include bacterial ghost (BGs) and virus-like-particles (VLPs). BGs are bacterial-cell envelopes that have been deprived of their DNA but maintain their antigenic properties and have been engineered to be carriers of antigens (Cui *et al.* 2010). VLPs are compounds artificially

constructed to resemble viruses that are non-infectious because they do not contain any viral genetic material (Cross *et al.* 2011). Initial results of BGs as an oro-nasal delivery system for ZP-based vaccines in brushtail possums showed a significant reduction in egg-fertilisation rates (Walcher *et al.* 2008) and offspring production (Duckworth in Cross *et al.* 2011). GnRH–VLP also elicited antibodies to GnRH (Cross *et al.* 2011). VLPs used to present zona (ZP3) and spermatozoaspecific peptides to laboratory mice, generated specific antibody responses and a significant reduction in litters born (Choudhury *et al.* 2009).

Because immunocontraceptive vaccines could typically affect multiple species, species specificity must be achieved through targeted delivery methods. Examples include floating rafts to deliver baits to aquatic species (Reynolds *et al.* 2004), baits placed inside burrow systems (Delahay *et al.* 2000) and species-specific delivery devices such as the BOS (Boar-Operated System), the latter developed to deliver baits to wild boar and feral pigs (Massei *et al.* 2010*c*; Campbell *et al.* 2011).

Immunocontraceptive vaccines can also be delivered through genetically modified self-sustaining infectious vectors. These include recombinant myxoma virus for rabbits, murine cytomegalovirus in mice and feline retroviruses for feral cats (Robinson et al. 1997; Courchamp and Cornell 2000; Seamark 2001; Singleton et al. 2002; Cowan et al. 2008). The main advantages of self-sustaining infectious vectors of immunocontraception include the feasibility of large-scale applications, both in terms of number of animals and areas covered, the availability of a humane and species-specific control method with potential for a good cost-benefit outcome and the possibility of providing long-term wildlife conflict resolution (e.g. McLeod et al. 2007; Tyndale-Biscoe and Hinds 2007). Criticism of this approach raised concerns regarding its irreversibility, the difficulty of controlling the vectors once released, possible mutations of the vectors that could affect non-target species and possible development of population resistance to these vectors (e.g. Barlow 2000; Tyndale-Biscoe and Hinds 2007; Williams 2007). For these reasons, none of these systems has been approved for field studies. In specific cases, the benefits of vectored immunocontraception may overcome the potential costs. For instance, Courchamp and Cornell (2000) suggested that actively disseminating immunocontraception systems should be employed for eradicating feral cats on islands, because of humaneness, environmental safety, low cost and wide coverage of inaccessible areas of these contraceptives.

In New Zealand, attention has recently turned to speciesspecific genetically modified non-transmissible and transmissible organisms (reviewed in Cross *et al.* 2011). Among the nontransmissible organisms, good candidates are the recombinant adenoviruses and the vaccinia virus belonging to the pox-virus family, the latter being widely used in veterinary and human vaccines. Among the transmissible organisms, research has focussed on possum-specific nematode parasites (Cowan *et al.* 2006, 2008).

Fertility-control impact on wildlife populations

Fertility control is employed to reduce population size or growth or to decrease the impact of wildlife on human

| Aim | Species | Trial | Method | Results and conclusions | Reference |
|---|--|---------------------|----------------------------------|---|--|
| Evaluate effect of hormonal competence and imposed FC on population dynamics | House mouse (<i>Mus domesticus</i>) | Enclosure | Tubal ligation vs ovariectomy | No differences in effect of both methods on population size. 67% infertility, imposed at the beginning of an 18-week study, reduced population size and growth rate. Litter size of fertile females increased in the sterilised groups | Chambers et al. 1999 |
| Evaluate impact of FC on population size | Ricefield rat Rattus argentiventer | Enclosure and model | Tubal ligation, ovariectomy, | FC-induced compensatory reproduction and improved survival of juveniles did not prevent a reduction in population size if 50–75% founder females were sterilised at the beginning of the reproductive season | Jacob <i>et al</i> . 2004 |
| As above | European rabbit Oryctolagus cuniculus | Enclosure and field | Tubal ligation | FC dampened seasonal population changes but did not reduce adult abundance. Improved survival compensated for the effects of sterilising up to 80% of females | Twigg <i>et al.</i> 2000; Williams <i>et al.</i> 2007 |
| As above | White-tailed deer Odocoileus virginianus | Field and model | PZP vaccine | FC feasible, over a 4-year study, to maintain small (<200) suburban deer populations at 30–70% of carrying capacity if ~60% females were treated with vaccine | Rudolph et al. 2000 |
| As above | White-tailed deer | Field | PZP vaccine | FC induced a 7.9% population decline per year over a 6-year study, in a suburban deer population | Rutberg et al. 2004 |
| As above | White-tailed deer | Field and model | PZP vaccine | FC caused a 27–58% % decline in population size in the 5–10 years following treatment of females | Rutberg and Naugle 2008 |
| As above | Wild horse (<i>Equus caballus</i>) | Field | PZP vaccine | The effort required to achieve zero population growth decreased, as 95%, 83% and 84% of all adult mares were treated in each of the first 3 years, compared with 59% and 52% during the last 2 years. FC increased longevity and improved body condition | Turner and Kirkpatrick 2002 |

Table 2. Examples of empirical and theoretical applications of fertility control (FC) at population level in captive and free-living wildlife species FC, fertility control

(continued next page)

| Aim | Species | Trial | Method | Results and conclusions | Reference |
|--|------------------------------------|-------|--|--|-------------------------------|
| As above | Wild horse | Field | PZP vaccine | FC prevented population growth within 2 years; by Year 11, the population had declined by 22.8%. FC also increased longevity of mares | Kirkpatrick and Turne 2008 |
| As above | Wild horse | Model | PZP vaccine | FC can be used for small (<200), isolated populations to reduce population size to the target number in 5–8 years | Ballou <i>et al.</i> 2008 |
| As above | Elephant Loxodonta africana | Field | PZP vaccine | FC of all sexually mature females in a small population (73 animals) prevented population growth for the 4-year study | Delsink et al. 2007 |
| As above | Elephant | Model | Immuno- contraception | 'Rotational' FC can be used to increase the span of calving intervals, slow population growth rate and alter age structure | Druce <i>et al.</i> 2011 |
| As above | Possum Trichosurus vulpecula | Field | Tubal ligation | Immigration and increased survival rate of sterilised females compensated for effects of FC and maintained population stable. In a 4-year study, sterility rates of 50% and 80% females resulted in 60% and 74% reduction to <i>per</i> <i>capita</i> rate of recruitment | Ramsey 2005 |
| Evaluate impact of culling and FC on population size | Brandt's vole Microtus brandti | Model | Generic contraception | In a 3-year study, FC applied in autumn to 85% of the females was more effective than culling in reducing population size | Shi et al. 2002 |
| As above | Wildlife | Model | Generic contraception | FC was more effective than culling in reducing population size for medium and large-size animals | Zhang 2000 |
| As above | White-tailed deer | Model | Generic contraceptive | FC was more efficient than culling in reducing population size provided >50% females were maintained infertile | Hobbs et al. 2000 |
| As above | Elk <i>Cervus elaphus</i> | Model | Yearlong vs lifelong contraceptive | FC using lifetime contraceptives was more efficient than any other population control option | Bradford and Hobbs 2008 |
| Evaluate impact of removal and FC on population size | Feral horse | Model | Generic contraception | Compared with removal, FC resulted in smaller, less fluctuating population size | Gross 2000 |

 Table 2.
 (continued)

Table 2. (continued)

| Aim | Species | Trial | Method | Results and conclusions | Reference |
|---|---------------------------------------|-----------------|---|---|-------------------------------|
| Evaluate effects of FC on behaviour and survival | Fox Vulpes vulpes | Field | Tubal ligation | In a 3-year study, FC did not affect territorial behaviour, dispersal and survival | Saunders et al. 2002 |
| Evaluate factors affecting time to reduce a population through FC | White-tailed deer | Model | Permanent sterilisation | FC could reduce a population by 30–60% in 4–10 years if 25–50% of fertile females were captured and sterilised every year | Merrill et al. 2003 |
| Evaluate effects of immigration, stochasticity and variation in capture process on FC to manage population size | White-tailed deer | Model | Permanent sterilisation | FC was unlikely to reduce the size of an open population. In a closed population, permanent sterilisation could reduce population size if 30–45% deer were captured each year | Merrill et al. 2006 |
| Assess potential of FC to eradicate cats on islands | Feral cat Felis catus | Model | Immuno contraception | Virus-vectored and bait- delivered contraceptives were predicted to eradicate a cat population | Courchamp and Cornell 2000 |
| Evaluate impact of culling, vaccination and FC added to vaccination on rabies control | Fox Vulpes vulpes | Model | Generic contraceptive | Integrating FC with rabies vaccination was predicted to be more successful than rabies vaccination only for rabies control | Smith and Wilkinson 2003 |
| As above, on rabies and bovine tuberculosis | Fox European badger Meles meles | Model | Generic contraceptive | FC added to rabies vaccination had a similar impact of culling on population reduction and disease eradication | Smith and Cheeseman 2002 |
| Test FC to eradicate bovine tuberculosis | European badger | Model | Generic contraceptive | FC, integrated with culling was predicted to be more effective than culling alone to eradicate the disease | White <i>et al.</i> 1997; |
| Estimate effect of FC on leptospirosis and bovine tuberculosis transmission | Possum Trichosurus vulpecula | Field and model | Tubal ligation vs inhibitors that prevent oestrus | In a 3-year study, tubal ligation, that does not prevent oestrus, caused an increase of disease transmission. FC that prevents oestrus was predicted to decrease the horizontal transmission rate of diseases due to reduced contact rate between animals | Caley and Ramsey 2001 |
| Estimate effect of FC, disease vaccination and culling on bovine tuberculosis transmission | Possum | Model | Generic contraceptive | An initial cull followed by FC and oral vaccination applied every 3 years was considered as the most cost-effective strategy | Ramsey and Efford 2010 |
| Test effects of FC on possum behaviour and transmission of leptospirosis | Possum | Field | Tubal ligation vs gonadectomy | In a 5-year study, both FC types did not affect the spatial behaviour and dominance status of | Ramsey 2007 |

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| Aim | Species | Trial | Method | Results and conclusions | Reference |
|---|--------------------------------|-----------------|---|---|-----------------------------|
| | | | | females. Gonadectomy reduced the seasonal breeding range size of males. In both sexes, gonadectomy decreased the horizontal transmission rate of disease due to a reduced contact rate between animals | |
| Test FC to improve efficiency of rabies eradication in urban dog populations | Dog <i>Canis familiaris</i> | Model | Immuno- contraception | FC added to rabies vaccination was predicted to reduce the proportion of dogs to be treated with rabies vaccines and the duration of vaccination campaigns | Carroll <i>et al</i> . 2010 |
| Evaluate effect of FC on population size and rabies control | Dog | Field and model | Surgical sterilisazion | Sustained sterilisation of 62–86% dogs in 2 years led to a 34% reduction in population size. The model predicted that FC could reduce population size and in the long term lead to rabies elimination | Totton <i>et al</i> . 2010 |
| Evaluate effect of FC on population size | Cat <i>Felis catus</i> | Model | Surgical sterilisazion vs contraception | >51–60% of females must be rendered infertile every year to halt population growth | Budke and Slater 2009 |

Table 2. (continued)

interests (Table 2). Understanding how fertility control affects population dynamics and social behaviour is crucial for evaluating the effectiveness of this method and estimating the effort required for successful practical applications. Here, we summarise the evidence from theoretical and empirical studies on factors that may influence population and behavioural responses to fertility control. We also offer an overview of advantages and disadvantages of fertility control compared with other methods used to mitigate human–wildlife conflicts. Ideally, fertility control would affect only natality. In practice, fertility control may also indirectly affect physiology and survival, as well as spatial and social behaviour of treated animals. These effects depend on species-specific behavioural and life-history traits, on the type of fertility-control agents used and on the proportion of a population treated with contraceptives.

Debates about the relative efficiency of fertility control and culling have largely centred on definitions. If efficiency is defined in terms of the time taken to achieve the desired effect, then culling will always be more efficient because fertility control alone cannot generate a larger, more rapid population decline than is the natural mortality rate (Bradford and Hobbs 2008; McLeod and Saunders 2014). Conversely, fertility control might be more efficient than culling if the remaining infertile animals maintain sufficient density-dependent feedback constraints on recruitment and survival (Zhang 2000). Typically, lethal control achieves an

initial rapid reduction in population size; several models have suggested that fertility control can be used, following lethal control, to maintain density at the reduced level (e.g. White *et al.* 1997; Merrill *et al.* 2006).

When the size of a population is suddenly reduced, compensatory density-dependent processes may act to return the population to its previous level (Bomford 1990; Barlow 1994; Twigg et al. 2000; Sinclair 2003; Ramsey 2005). In short-lived species, fecundity can make a greater proportional contribution than survival to population growth, and the reverse occurs in long-lived species (Sibly and Hone 2002). Compensatory population changes that may occur in response to fertility control are likely to be less pronounced than those following population reduction by lethal methods, depending on whether populations are regulated by density-dependent mortality or recruitment (Johnson and Tait 1983; Bomford 1990; Bomford and O'Brien 1997). For instance, in populations of mice and rabbits, a compensatory response in female productivity did not offset the effects of sterilisation when 60-80% of the females were made infertile (Chambers et al. 1999; Twigg and Williams 1999; Twigg et al. 2000) (Table 2).

Initial models (e.g. Hone 1992; Barlow 1994) suggested that fertility control would be most effective for small-sized r-selected species, characterised by high fertility and low survival. These conclusions have been challenged by models showing that large,

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long-lived species might be easier to manage with fertility control than are smaller, shorter-lived ones because a lower proportion of the population would need to be treated (Hone 1999), particularly if lifelong contraceptives were employed (Hobbs *et al.* 2000; Table 2).

However, in long-lived species, the benefits of using fertility control to decrease population size are accrued in the long term (Twigg *et al.* 2000; Cowan and Massei 2008; Kirkpatrick and Turner 2008). Others have suggested that contraception is likely to be better than culling for controlling species with medium to high instantaneous rates of population increase, but equivalent to culling for species with low instantaneous rates of population increase (Zhang 2000).

In some instances, fertility control might be required to reduce or halt population growth rather than to decrease population size. Druce *et al.* (2011) introduced the concept of individualbased 'rotational immunocontraception' and showed that using reversible immunocontraceptives on elephants on an individual rotational basis increased inter-calving intervals and lowered population growth to a predetermined rate.

In the context of wildlife diseases, fertility control has several specific advantages over culling. For instance, culling can increase disease transmission by disrupting social organisation and by increasing animal movements, thus leading to increased contact rates (e.g. Choisy and Rohani 2006; Carter et al. 2007; Wilkinson et al. 2009). Fertility control is likely to cause less social perturbation than is lethal control (Swinton et al. 1997; Tuvttens and Macdonald 1998). Where disease transmission has a substantial vertical component (from mother to offspring), such as brucellosis in bison, fertility control could be used to target such transmission (Miller et al. 2004). Fertility control removes the physiological costs of reproduction and lactation, which may thus enhance physical condition and improve immune function, thereby reducing susceptibility to disease. When fertility control is used as a tool for disease control, methods that prevent ovulation are likely to be more successful than those that only block fertilisation. For instance, the transmission coefficient of leptospirosis in possums was 28% higher in populations subjected to tubal ligation that leaves females cycling but unable to conceive, than in populations not subjected to fertility control (Caley and Ramsey 2001). In contrast, endocrine disruption caused by gonadectomy in possums decreased the leptospirosis transmission rate by 63-88% in sterilised female and male possums (Ramsey 2007).

Whether culling is more effective than disease vaccination for wildlife disease management will partly depend on assumptions about disease transmission, including whether the rate of transmission depends on the absolute density of susceptible individuals or the relative density of susceptible and immune individuals. Fertility control reduces the recruitment of new susceptible individuals. Thus, several models have suggested a synergistic effect of fertility control on disease vaccination that reduces the effort required to eliminate a disease from a population (Smith and Cheeseman 2002; Carroll *et al.* 2010). Complementary effects of disease vaccination and fertility control have also been suggested for the elimination of rabies from red fox (Smith and Wilkinson 2003) and free-roaming dogs (Carroll *et al.* 2010; Massei *et al.* 2010*b*; Massei 2012, 2013;

Massei and Miller 2013) and for management of bovine tuberculosis in brushtail possums (Ramsey and Efford 2010).

In terms of behaviour, fertility control might affect hierarchically structured species where dominant females suppress breeding in subordinate females. If the social status of dominant females was compromised by sterilisation, intermediate levels of sterility could lead to increased productivity (Caughley *et al.* 1992). Conversely, if dominance was maintained, irrespective of reproductive output, contraception of dominant females should lead to decreased population-level productivity.

Although more research is clearly needed in this area, the disruption of female reproductive hormonal function does not affect social behaviour in several species (e.g. Chambers et al. 1999; Poiani et al. 2002; Kirkpatrick et al. 2011; Table 2). A few studies have suggested that changes in socio-sexual behaviour involving decreased libido, decreased sexual activity and aggressiveness could lead to disruption of social structure and spacing behaviour. For instance, in female ring-tailed lemurs (Lemur catta), Crawford et al. (2011) found that medroxyprogesterone acetate significantly altered the olfactory cues that signal fertility, individual chemical 'signature' and relatedness, and suggested that treatment with this contraceptive may disrupt social interactions, kin recognition and mate choice in primates. In brushtail possums, tubally ligated but hormonally competent females showed extended breeding seasons, which attracted higher densities of males into the study area (Ji et al. 2000). The average body condition of these males was significantly poorer than that of males in control areas. Similarly, an extension of the breeding season in deer treated with PZP vaccine resulted in an increase in energy expenditure by males (Killian and Miller 2000; Curtis et al. 2002). In contrast, decreased sexual activity of both males and females has been reported in deer treated with a GnRH vaccine (Miller et al. 2000, 2009).

Physiological responses to fertility control include increased improved health, body condition and, hence, survival, possibly linked to the reduced costs of reproduction. For example, sterilised feral Soay rams showed increased food consumption and survival compared with control rams, ultimately leading to increased animal numbers and impact on the plant community (Jewell 1986). PZP-based immunocontraceptives increased lifespan and body conditions of mares (Turner and Kirkpatrick 2002; Kirkpatrick and Turner 2007), tubal ligation increased survival in rabbits (Twigg et al. 2000; Williams et al. 2007) and GonaCon improved body condition of deer (Gionfriddo et al. 2011b). Conversely, Saunders et al. (2002) observed no differences in survival, dispersal or territory size of surgically sterilised foxes compared with fertile foxes, although sterilised vixens were more likely than fertile females to share their territories with each other.

Some authors hypothesised that the use of immunocontraceptive vaccines to manage wildlife could result in the evolution of resistance through selection for individuals that remain fertile because of low or no response to vaccination (e.g. Gross 2000; Magiafoglou *et al.* 2003; Holland *et al.* 2009). Although some studies concluded that the evolution of resistance was unlikely (Magiafoglou *et al.* 2003), research programs on mammalian immunocontraception should involve measurement

of the heritability of non-response (Cooper and Larsen 2006). For instance, in brushtail possums, two sets of alleles (haplotypes) were found to associate significantly with differences in response to immunocontraceptive vaccines (Holland *et al.* 2009). The characterisation of these haplotypes offers potential to identify factors affecting non-responders.

Criteria to assess the suitability of fertility control to mitigate human-wildlife conflicts

Conflicts involving overabundant species often demand immediate solutions. If fertility control is chosen to manage overabundant wildlife, Kirkpatrick and Franck (2005) proposed a three-step approach that consisted of (1) identifying a contraceptive suitable for the species to be managed, (2) assessing whether the contraceptive could be delivered under field conditions and (3) evaluating whether the desired population effect could be achieved in the field. We suggest expanding this approach by incorporating additional elements that include public consultation, evaluation of potential animal-welfare issues, population responses, costs and sustainability. Although these suggestions are presented as a decision tree (Fig. 2), the process is not necessarily linear; for instance, modelling would contribute to several stages in the process, such as informing the product specifications for the choice of contraceptives and assessment of the necessary efficacy. Likewise, costs can be estimated at an earlier stage and recalculated, if needed, later on in the process.

Local authorities and animal-welfare organisations advocate fertility control to manage human–wildlife conflicts, particularly in urban and suburban areas (Barr *et al.* 2002; Curtis *et al.* 2008). Conversely, many hunting groups, particularly in North America, oppose the use of fertility control because of concerns that this method will replace sport hunting (Kirkpatrick 2007; Curtis *et al.* 2008; Fagerstone *et al.* 2010). These polarised views suggest that at the planning stage, comprehensive stakeholder consultation and engagement is crucial to agree on common goals as well as methods to achieve these goals to manage wildlife.

Key questions when assessing the potential of fertility control to mitigate human–wildlife conflicts are 'What is the overall proportion of the population that must be rendered infertile to achieve the target reduction in population size or to stop population growth or to achieve the desired reduction in damage?', 'What is the effort required to achieve the target population size within a certain timeframe?' and 'How frequently does the treatment need to be applied?' (Chambers *et al.* 1999; Hobbs *et al.* 2000; Bradford and Hobbs 2008). The question of the impact of fertility control on damage reduction can be complicated as population size and damage are not always linearly related and a reduction in population size is not necessarily followed by a proportional decrease in damage (Hone 1995, 2002).

Captive studies or data collected on similar species could be used to inform decisions about the type of contraceptive to be employed. If the available data confirm the potential effectiveness of the approach, the study could progress toward modelling the effects of fertility control on population dynamics (e.g. Jacob *et al.* 2008). If modelling suggests that the objectives can be

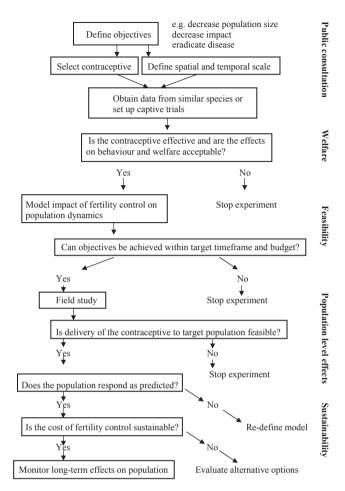


Fig. 2. Staged approach for assessing the suitability of fertility control to manage human–wildlife conflicts. This decision tree assumes that fertility control has been chosen over other options of population control.

achieved within target time scale and available budget, the project can move to a field trial.

The economic costs of reducing population densities through fertility control alone, with current delivery methods, are generally expected to be high. For instance, Rutberg (2005) estimated that the cost to render infertile a medium-largesize individual mammal varied between US\$25 and US\$500. Delsink et al. (2007) calculated that in 2005 the average cost of managing elephants through aerial vaccination with immunocontraceptives cost US\$98-110 per animal, inclusive of darts, vaccine, helicopter and veterinary assistance. The cost of capture, handling and administering contraceptives to whitetailed deer in various contexts was estimated to be in excess of US\$1000 per deer, with 75% of this cost being due to veterinary time and drugs (Boulanger et al. 2012). These costs would drop significantly if the contraceptives were delivered by trained wildlife managers and if animal capture were organised with the assistance of volunteers donating their time and skills to a project. In addition, the effort required to treat a wildlife population will be influenced by animal density, approachability of individual animals, access to private and public land, and efficacy of the contraceptive treatment (Rudolph et al. 2000).

Hobbs et al. (2000) suggested that fertility control of deer will be cost-effective, compared with culling, only where recreational hunting is not feasible and population control is carried out by employing professionals to cull deer. Comparing the costs of fertility control-based management with those of alternative control options and identifying who should bear these costs might raise awareness of the economics of current management practices among stakeholders and add a different perspective to wildlife management decision-making. This awareness would be further enhanced if the full costs, including negative environmental and welfare consequences, associated with each option were included. Once a field trial has been implemented, the effectiveness, costs and feasibility of using fertility control to manage human-wildlife conflicts can be evaluated, together with actual population responses to imposed infertility. The data collected can then be used to refine the model and to determine whether fertility control can be a sustainable approach. In addition, potential, unforeseen effects of imposing infertility must be evaluated; these include monitoring survival rate, immigration and emigration rates as well as disease transmission rates.

Conclusions

Ethical considerations regarding humane treatment of animals are shaping public attitudes toward acceptable methods of mitigating human-wildlife conflicts. The present review confirmed that the interest in fertility inhibitors for wildlife has steadily grown in the past three decades, as indicated by the trends in publications in this area. Possible reasons for the increasing trend in research and development include (1) new approaches based on advances in the understanding of the molecular mechanisms regulating mammalian fertility, (2) availability of new technologies that make practical applications for wildlife more feasible, (3) stakeholder interest in developing alternatives to culling, (4) increasing human-wildlife conflicts, (5) advances in other fields, such as contraceptives developed for livestock and companion animals, with potential for wildlife applications, (6) advances in analytical techniques used in population modelling studies and (7) internet-associated information flow raising public awareness of wildlife fertility control.

The review highlighted that several safe, effective and longlasting fertility inhibitors such as levonorgestrel, deslorelin, PZP and GnRH-based immunocontraceptives are now available to manage wildlife and that successful population control has been achieved in several field applications (Table 2). So far, most empirical and theoretical studies have focussed on ungulates, marsupials and rodents, with the main aim of reducing population size or growth, and on carnivores, with studies aimed at decreasing disease transmission. In many instances, even when reduction in population size or growth has been successful, the mitigation of the conflict that caused fertility control to be employed is inferred but not quantified. Further research is required to address whether the application of fertility control can mitigate context-specific human–wildlife conflicts.

The use of fertility control to mitigate human-wildlife conflicts might raise inappropriate expectations if its costs and benefits were not clearly examined on a case-by-case basis. A general conclusion from the results of the studies summarised in the present review is that a substantial initial effort is required if fertility control is the sole method chosen to manage overabundant populations. However, as the proportion of infertile females increases, this effort will decline and remain constant once the desired density has been achieved. In addition, the review showed that there is growing recognition of the possible synergy between fertility control and disease vaccination to optimise the maintenance of herd immunity in the management of wildlife diseases. Before fertility-control applications can be advocated as a tool to mitigate human-wildlife conflicts, there are still many aspects that must receive further attention. These aspects include the development of contraceptives for wide-scale wildlife applications, the development of species-specific, inexpensive delivery methods, field applications demonstrating population responses to imposed infertility in species with different lifehistory traits, and the evaluation of feasibility, costs and sustainability of population-management programs based on fertility control.

Because efficacy and humaneness are often the primary public concerns regarding any type of wildlife management, defining these terms, particularly in relation to other methods of population control, is crucial for any management plan to obtain and maintain public support in relation to specific, well defined objectives. Efficacy can be defined as (1) the proportion of the population rendered infertile, (2) the speed of reduction in population size or damage or (3) the eradication of a disease. Humaneness can be defined as (1) the level of stress experienced by treated animals, (2) the severity and type of side effects, (3) the proportion of animals likely to experience negative side effects following the use of a contraceptive, (4) the proportion of animals likely to suffer from capture, handling and anaesthesia associated with administering the contraceptives, or (5) as a combination of all these definitions. Comparisons of fertility control and other population-management methods often fail to account for all the costs and benefits, including welfare costs. Defining these terms and adhering to guidelines for assessing and comparing the relative humaneness of wildlife control methods is one of the main challenges for human-wildlife conflict mitigation (Sharp and Saunders 2008). In addition to the scientific challenges of exploring the effects of fertility control on individuals and wildlife populations, regulatory and legal requirement for the application of contraceptives on wildlife must be met. The fact that in different countries fertility inhibitors can be registered as pesticides, biocides or veterinary medicines, depending on the mode of action and on the target species, coupled with the significant costs of registration, present hurdles for development and use of novel products (Humphrys and Lapidge 2008).

We suggested criteria that could be used during public and stakeholders consultations to determine whether fertility control should be used to manage overabundant wildlife. This assumes that fertility control represents a rational approach to the problems posed by animal populations. However, the review highlighted how for each context and species, the use of fertility control, alone or integrated with other methods of population control, should be evaluated and compared with alternative options to mitigate conflicts between human interests and wildlife.

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References

- Asa, C. S., and Porton, I. J. (Eds) (2005). 'Wildlife Contraception: Issues, Methods and Applications.' (Johns Hopkins University Press: Baltimore, MD.)
- Aune, K., Terry, J., Kreeger, T. J., Thomas, J., and Roffe, T. J. (2002). Overview of delivery systems for the administration of vaccines to elk and bison of the greater Yellowstone area. In 'Proceedings of Brucellosis in Elk and Bison in the Greater Yellowstone Area'. (Ed. T. J. Kreeger.) pp. 66–79. (Wyoming Game and Fish Department: Cheyenne, WY.)
- Avery, M. L., Yoder, C. A., and Tillman, E. A. (2008). Diazacon inhibits reproduction in invasive monk parakeet populations. *The Journal of Wildlife Management* 72, 1449–1452. doi:10.2193/2007-391
- Baker, D. L., Nett, T. M., Hobbs, N. T., Gill, R. B., and Miller, M. M. (1999). Evaluation of GnRH-toxin conjugate as an irreversible contraceptive in female mule deer. In 'Proceedings of The Wildlife Society 6th Annual Conference'. (Ed. S. Craven.) pp.5–11. (The Wildlife Society: Bethesda, MD.)
- Baker, D. L., Wild, M. A., Conner, M. M., Ravivarapu, H. B., Dunn, R. L., and Nett, T. M. (2002). Effects of GnRH agonist (leuprolide) on reproduction and behaviour in female wapiti (*Cervus elaphus nelsoni*). *Reproduction* 60, 155–167.
- Baker, D. L., Wild, M. A., Conner, M. M., Ravivarapu, H. B., Dunn, R. L., and Nett, T. M. (2004). Gonadotropin-releasing hormone agonist: a new approach to reversible contraception in female deer. *Journal of Wildlife Diseases* 40, 713–724. doi:10.7589/0090-3558-40.4.713
- Ball, B. A., Sabeur, K., Nett, T., and Liu, I. K. M. (2006). Effects of a GnRH cytotoxin on reproductive function in peripubertal male dogs. *Theriogenology* 66, 766–774. doi:10.1016/j.theriogenology.2005.11.024
- Ballou, J. D., Traylor-Holzer, K., Turner, A., Malo, A. F., Powell, D., Maldonado, J., and Eggert, L. (2008). Simulation model for contraceptive management of the Assateague Island feral horse population using individual-based data. *Wildlife Research* 35, 502–512. doi:10.1071/WR07124
- Barfield, J. P., Nieschlag, E., and Cooper, T. G. (2006). Fertility control in wildlife: humans as a model. *Contraception* 73, 6–22. doi:10.1016/ j.contraception.2005.06.070
- Barlow, N. D. (1994). Predicting the effect of a novel vertebrate biocontrol agent: a model for viral-vectored immunocontraception of New Zealand possums. *Journal of Applied Ecology* 31, 454–462. doi:10.2307/2404442
- Barlow, N. D. (2000). The ecological challenge in immunocontraception: editor's introduction. *Journal of Applied Ecology* 37, 897–902. doi:10.1046/j.1365-2664.2000.00591.x
- Barr, J. J. F., Lurz, P. W. W., Shirley, M. D. F., and Rushton, S. P. (2002). Evaluation of immunocontraception as a publicly acceptable form of vertebrate pest species control: the introduced grey squirrel in Britain as an example. *Environmental Management* **30**, 342–351. doi:10.1007/ s00267-002-2686-7
- Benfield, N., and Darney, P. D. (2011). Contraceptive implants. In 'Contraception'. (Ed. D. Shoupe.) pp. 57–66. (Wiley-Blackwell: Oxford, UK.)
- Beringer, J., Hansen, L. P., Demand, J. A., and Sartwell, J. (2002). Efficacy of translocation to control urban deer in Missouri: costs, efficiency, and outcome. *Wildlife Society Bulletin* 30, 767–774.
- Bertschinger, H. J., de Barros Guimarães, M. A., Trigg, T. E., and Human, A. (2008). The use of deslorelin implants for the long-term contraception of lionesses and tigers. *Wildlife Research* 35, 525–530. doi:10.1071/ WR07141

- Bomford, M. (1990). 'A Role for Fertility Control in Wildlife Management? Bureau of Natural Resources Bulletin No. 7.' (Australian Government Publishing Service: Canberra.)
- Bomford, M., and O'Brien, P. (1997). Potential use of contraception for managing wildlife pests in Australia. In 'Contraception in Wildlife Management. USDA–APHIS Technical Bulletin 1853'. (Ed. T. J. Kreeger.) pp. 205–214. (USDA–APHIS: Washington, DC.)
- Boulanger, J. R., Curtis, P. D., Cooch, E. G., and DeNicola, A. J. (2012). Sterilization as an alternative deer control technique: a review. *Human–Wildlife Interactions* 6, 273–282.
- Bradford, J. B., and Hobbs, N. T. (2008). Regulating overabundant ungulate populations: an example for elk in Rocky Mountain National Park, Colorado. *Journal of Environmental Management* 86, 520–528. doi:10.1016/j.jenvman.2006.12.005
- Brown, R. G., Bowen, W. D., Eddington, J. D., Kimmins, W. C., Mezei, M., Parsons, J. L., and Pohajdak, B. (1997). Evidence for a long-lasting single administration contraceptive vaccine in wild grey seals. *Journal* of Reproductive Immunology 35, 43–51. doi:10.1016/S0165-0378(97) 00047-8
- Budke, C. M., and Slater, M. R. (2009). Utilization of matrix population models to assess a 3-year single treatment nonsurgical contraception program versus surgical sterilization in feral cat populations. *Journal* of Applied Welfare Science 12, 277–292. doi:10.1080/10888700903 163419
- Bynum, K. S., Eisemann, J. D., Weaver, G. C., Yoder, C. A., Fagerstone, K. A., and Miller, L. A. (2007). Nicarbazin OvoControl G bait reduces hatchability of eggs laid by resident Canada geese in Oregon. *The Journal of Wildlife Management* 71, 135–143. doi:10.2193/2005-603
- Caley, P., and Ramsey, D. (2001). Estimating disease transmission in wildlife, with emphasis on leptospirosis and bovine tuberculosis in possums, and effects of fertility control. *Journal of Applied Ecology* 38, 1362–1370. doi:10.1046/j.0021-8901.2001.00676.x
- Campbell, T. A., Garcia, M. R., and Miller, L. A. (2010). Immunocontraception in male feral swine treated with a recombinant gonadotropin-releasing hormone vaccine. *Journal of Swine Health and Production* 18, 118–124.
- Campbell, T. A., Long, D. B., and Massei, G. (2011). Efficacy of the boaroperated-system to deliver baits to feral swine. *Preventive Veterinary Medicine* 98, 243–249. doi:10.1016/j.prevetmed.2010.11.018
- Cariño, C., Prasad, S., Skinner, S., Dunbar, B., Chirinos, M., Schwoebel, E., Larrea, F., and Dunbar, B. (2002). Localization of species conserved zona pellucida antigens in mammalian ovaries. *Reproductive Biomedicine Online* 4, 116–126. doi:10.1016/S1472-6483(10)61928-1
- Carroll, M. J., Singer, A., Smith, G. C., Cowan, D. P., and Massei, G. (2010). The use of immunocontraception to improve rabies eradication in urban dog populations. *Wildlife Research* 37, 676–687. doi:10.1071/ WR10027
- Carter, S. P., Delahay, R. J., Smith, G. C., Macdonald, D. W., Riordan, P., Etherington, T., Pimley, E., and Cheeseman, C. L. (2007). Cullinginduced social perturbation in Eurasian badgers *Meles meles* and the management of TB in cattle: an analysis of a critical problem in applied ecology. *Proceedings. Biological Sciences* 274, 2769–2777. doi:10.1098/ rspb.2007.0998
- Caughley, C., Pech, R. P., and Grice, D. (1992). Effect of fertility control on a population's productivity. *Wildlife Research* 19, 623–627. doi:10.1071/ WR9920623
- Chambers, L. K., Singleton, G. R., and Hinds, L. A. (1999). Fertility control of wild mouse populations: the effects of hormonal competence and an imposed level of sterility. *Wildlife Research* 26, 579–591. doi:10.1071/ WR98093
- Chittick, E., Rotstein, D., Brown, T., and Wolfe, B. (2001). Pyometra and uterine adenocarcinoma in a melengestrol acetate-implanted captive coati (*Nasua nasua*). Journal of Zoo and Wildlife Medicine **32**, 245–251.

- Choisy, M., and Rohani, P. (2006). Harvesting can increase severity of wildlife disease epidemics. *Proceedings. Biological Sciences* 273, 2025–2034. doi:10.1098/rspb.2006.3554
- Choudhury, S., Kakkar, V., Suman, P., Chakrabarti, K., Vrati, S., and Gupta, S. K. (2009). Immunogenicity of zona pellucida glycoprotein-3 and spermatozoa YLP(12) peptides presented on Johnson grass mosaic virus-like particles. *Vaccine* 27, 2948–2953. doi:10.1016/j.vaccine. 2009.03.002
- Conner, M. M., Baker, D. L., Wild, M. A., Powers, J. G., Hussain, M. D., Dun, R. L., and Nett, T. M. (2007). Fertility control in free-ranging elk using gonadotropin releasing hormone agonist leuprolide: effects on reproduction, behavior, and body condition. *The Journal of Wildlife Management* 71, 2346–2356. doi:10.2193/2006-463
- Cooper, D. W., and Larsen, E. (2006). Immunocontraception of mammalian wildlife: ecological and immunogenetic issues. *Reproduction* 132, 821–828. doi:10.1530/REP-06-0037
- Coulson, G., Nave, C. D., Shaw, J., and Renfree, M. B. (2008). Long-term efficacy of levonorgestrel implants for fertility control of eastern grey kangaroos (*Macropus giganteus*). Wildlife Research 35, 520–524. doi:10.1071/WR07133
- Courchamp, F., and Cornell, S. J. (2000). Virus-vectored immunocontraception to control feral cats on islands: a mathematical model. *Journal of Applied Ecology* 37, 903–913. doi:10.1046/j.1365-2664.2000.00545.x
- Cowan, D. P., and Massei, G. (2008). Wildlife contraception, individuals and populations: how much fertility control is enough? In 'Proceedings 23rd Vertebrate Pest Conference'. (Eds R. M. Timm and M. B. Madon.) pp. 220–228. (University of California: Davis, CA.)
- Cowan, D. P., and Quy, R. J. (2003). Rodenticide use against farm rat populations: biological constraints on effectiveness and safety. In 'Conservation and Conflict: Mammals and Farming in Britain'. (Eds F. Tattersall and W. Manly.) pp. 172–185. (Linnean Society: London.)
- Cowan, D. P., Massei, G., and Mellows, R. J. B. (2006). A modeling approach to evaluating potential applications of emerging fertility control technologies in the UK. In 'Proceedings 22nd Vertebrate Pest Conference'. (Eds R. M. Timm, J. J. O'Brien.) pp. 55–62. (University of California: Davis, CA.)
- Cowan, P. E., Grant, W. N., and Ralston, M. (2008). Assessing the suitability of the parasitic nematode *Parastrongyloides trichosuri* as a vector for transmissible fertility control of brushtail possums in New Zealand – ecological and regulatory considerations. *Wildlife Research* 35, 573–577. doi:10.1071/WR07174
- Crawford, J., Boulet, M., and Drea, C. M. (2011). Smelling wrong: hormonal contraception in lemurs alters critical female odour cues. *Proceedings. Biological Sciences* 278, 122–130. doi:10.1098/rspb.2010.1203
- Cross, M. L., Zheng, T., Duckworth, J. A., and Cowan, P. E. (2011). Could recombinant technology facilitate the realisation of a fertility-control vaccine for possums? *New Zealand Journal of Zoology* 38, 91–111. doi:10.1080/03014223.2010.541468
- Cui, X., Duckworth, J. A., Lubitz, P., Molinia, F. C., Haller, C., Lubitz, W., and Cowan, P. E. (2010). Humoral immune responses in brushtail possums (*Trichosurus vulpecula*) induced by bacterial ghosts expressing possum zona pellucida 3 protein. *Vaccine* 28, 4268–4274. doi:10.1016/j.vaccine.2010.04.032
- Curtis, P. D., Pooler, R. L., Richmond, M. E., Miller, L. A., Mattfeld, G. F., and Quimby, F. W. (2002). Comparative effects of GnRH and porcine zona pellucida (PZP) immunocontraceptive vaccines for controlling reproduction in white-tailed deer (*Odocoileus virginianus*). *Reproduction* 60, 131–141.
- Curtis, P. D., Richmond, M. E., Miller, L. A., and Quimby, F. W. (2007). Pathophysiology of white-tailed deer vaccinated with porcine zona pellucida immunocontraceptive. *Vaccine* 25, 4623–4630. doi:10.1016/ j.vaccine.2007.03.033

- Curtis, P. D., Richmond, M. E., Miller, L. A., and Quimby, F. W. (2008). Physiological effects of gonadotropin-releasing hormone immunocontraception on white-tailed deer. *Human–Wildlife Conflicts* 2, 68–79.
- D'Occhio, M. J., Fordyce, G., Whyte, T. R., Jubb, T. F., Fitzpatrick, L. A., Cooper, N. J., Aspden, W. J., Bolam, M. J., and Trigg, T. E. (2002). Use of GnRH agonist implants for long-term suppression of fertility in extensively managed heifers and cows. *Animal Reproduction Science* 74, 151–162. doi:10.1016/S0378-4320(02)00189-6
- Daszak, P., Cunningham, A. A., and Hyatt, A. D. (2000). Emerging infectious diseases of wildlife – threats to biodiversity and human health. *Science* 287, 443–449. doi:10.1126/science.287.5452.443
- Delahay, R. J., Brown, J. A., Mallinson, P. J., Spyvee, P. D., Handoll, D., Rogers, L. M., and Cheeseman, C. L. (2000). The use of marked bait in studies of the territorial organization of the European badger (*Meles meles*). *Mammal Review* **30**, 73–87. doi:10.1046/j.1365-2907.2000. 00058.x
- Delsink, A. K., and Kirkpatrick, J. (Eds) (2012). 'Free-ranging African Elephant Immunocontraception.' (Trident Press: Cape Town, South Africa.)
- Delsink, A. K., van Altena, J. J., Grobler, D., Bertschinger, H., Kirkpatrick, J., and Slotow, R. (2007). Implementing immunocontraception in freeranging African elephants at Makalali Conservancy. *Journal of the South African Veterinary Association* 78, 25–30. doi:10.4102/jsava. v78i1.282
- DeNicola, A. J., Kesler, D. J., and Swihart, R. K. (1997). Dose determination and efficacy of remotely delivered norgestomet implants on contraception of white-tailed deer. *Zoo Biology* 16, 31–37. doi:10.1002/(SICI)1098-2361(1997)16:1<31::AID-ZOO5>3.0.CO;2-C
- DeNicola, A. J., VerCauteren, K. C., Curtis, P. D., and Hygnstrom, S. E. (Eds) (2000). 'Managing White-tailed Deer in Suburban Environments: a Technical Guide.' (Cornell Cooperative Extension: Ithaca, NY.)
- Druce, H. C., Mackey, R. L., and Slowtow, R. (2011). How immunocontraception can contribute to elephant management in small, enclosed reserves: Munyawana population. *PLoS ONE* 6, e27952. doi:10.1371/journal.pone.0027952
- Duckworth, J. A., Cui, X., Scobie, S., Arrow, J., and Cowan, P. E. (2008). Development of a contraceptive vaccine for the marsupial brushtail possum (*Trichosurus vulpecula*): lack of effects in mice and chickens immunised with recombinant possum ZP3 protein and a possum ZP3 antifertility epitope. *Wildlife Research* 35, 563–572. doi:10.1071/ WR07139
- Duka, T., and Masters, P. (2005). Confronting a tough issue: fertility control and translocation for over-abundant koalas on Kangaroo Island, South Australia. *Ecological Management & Restoration* 6, 172–181. doi:10.1111/j.1442-8903.2005.00234.x
- Eade, J. A., Roberston, I. D., and James, C. M. (2009). Contraceptive potential of porcine and feline zona pellucida A, B and C subunits in domestic cats. *Reproduction* 137, 913–922. doi:10.1530/REP-08-0471
- Eidne, K. A., Henery, C. C., and Aitken, R. J. (2000). Selection of peptides targeting the human sperm surface using random peptide phage display to identify ligands homologous to ZP3. *Biology of Reproduction* 63, 1396–1402. doi:10.1095/biolreprod63.5.1396
- Eymann, J., Herbert, C. A., Thomson, B. P., Trigg, T. E., Cooper, D. W., and Eckery, D. C. (2007). Effects of deslorelin implants on reproduction in the common brushtail possum (*Trichosurus vulpecula*). *Reproduction, Fertility and Development* **19**, 899–909. doi:10.1071/RD07046
- Fagerstone, K. A., Miller, L. A., Eisemann, J. D., O'Hare, J. R., and Gionfriddo, J. P. (2008). Registration of wildlife contraceptives in the United States of America, with OvoControl and GonaCon immunocontraceptive vaccines as examples. *Wildlife Research* 35, 586–592. doi:10.1071/WR07166
- Fagerstone, K. A., Miller, L. A., Killian, G. J., and Yoder, C. A. (2010). Review of issues concerning the use of reproductive inhibitors, with

particular emphasis on resolving human–wildlife conflicts in North America. *Integrative Zoology* **5**, 15–30. doi:10.1111/j.1749-4877.2010.00185.x

- Ferri, M., Ferraresi, M., Gelati, A., Zannetti, G., Ubaldi, A., Contiero, B., and Bursi, E. (2009). Use of nicarbazine in the control of urban pigeon colonies in Italy in 1990–2007. *Annali della Facolta'di Medicina Veterinaria di Parma* 29, 91–102.
- Fraker, M. A., Brown, R. G., Gaunt, G. E., Kerr, J. A., and Pohajdak, B. (2002). Long-lasting, single-dose immunocontraception of feral fallow deer in British Columbia. *The Journal of Wildlife Management* 66, 1141–1147. doi:10.2307/3802946
- Fu, H., Zhang, J., Shi, D., and Wu, X. (2013). Effects of levonorgestrelquinestrol (EP-1) treatment on Mongolian gerbil wild populations: a case study. *Integrative Zoology* 8, 277–284. doi:10.1111/1749-4877.12018
- Garrott, R. A., and Siniff, D. B. (1992). Limitations of male-oriented contraception for controlling feral horse populations. *The Journal of Wildlife Management* 56, 456–464. doi:10.2307/3808859
- Gionfriddo, J. P., Eisemann, J. D., Sullivan, K. J., Healey, R. S., and Miller, L. A. (2009). Field test of a single-injection gonadotrophin-releasing hormone immunocontraceptive vaccine in female white-tailed deer. *Wildlife Research* 36, 177–184. doi:10.1071/WR08061
- Gionfriddo, J. P., Denicola, A. J., Miller, L. A., and Fagerstone, K. A. (2011a). Efficacy of GnRH immunocontraception of wild white-tailed deer in New Jersey. *Wildlife Society Bulletin* 35, 142–148. doi:10.1002/wsb.32
- Gionfriddo, J. P., Denicola, A. J., Miller, L. A., and Fagerstone, K. A. (2011b). Health effects of GnRH immunocontraception of wild white-tailed deer in New Jersey. *Wildlife Society Bulletin* 35, 149–160. doi:10.1002/wsb.17
- Giunchi, D., Baldaccini, N. E., Sbragia, G., and Soldatini, C. (2007). Use of pharmacological sterilisation to control feral pigeon populations. *Wildlife Research* 34, 306–318. doi:10.1071/WR06153
- Gobello, C. (2007). New GnRH analogs in canine reproduction. Animal Reproduction Science 100, 1–13. doi:10.1016/j.anireprosci.2006.08.024
- Gray, M. E., and Cameron, E. Z. (2010). Does contraceptive treatment in wildlife result in side effects? A review of quantitative and anecdotal evidence. *Reproduction (Cambridge, England)* 139, 45–55. doi:10.1530/ REP-08-0456
- Gray, M. E., Thain, D. S., Cameron, E. Z., and Miller, L. A. (2010). Multi-year fertility reduction in free-roaming feral horses with single-injection immunocontraceptive formulations. *Wildlife Research* 37, 475–481. doi:10.1071/WR09175
- Grignard, E., Cadet, R., Saez, F., Drevet, J. R., and Vernet, P. (2007). Identification of sperm antigens as a first step towards the generation of a contraceptive vaccine to decrease fossorial water vole *Arvicola terrestris sherman* proliferations. *Theriogenology* 68, 779–795. doi:10.1016/j.theriogenology.2007.06.010
- Gross, J. (2000). A dynamic simulation model for evaluating effects of removal and contraception on genetic variation and demography of Pryor Mountain wild horses. *Biological Conservation* 96, 319–330. doi:10.1016/S0006-3207(00)00078-1
- Gupta, S. K., and Bhandari, B. (2011). Acrosome reaction: relevance of zona pellucida glycoproteins. *Asian Journal of Andrology* **13**, 97–105. doi:10.1038/aja.2010.72
- Gupta, S. K., Srinivasan, V. A., Suman, P., Rajan, S., Nagendrakumar, S. B., Gupta, N., Shrestha, A., Joshi, P., and Panda, A. K. (2011). Contraceptive vaccines based on the zona pellucida glycoproteins for dogs and other wildlife population management. *American Journal of Reproductive Immunology* 66, 51–62. doi:10.1111/j.1600-0897.2011.01004.x
- Gupta, N., Chakrabarti, K., Prakash, K., Wadhwa, N., Gupta, T., and Gupta, S. K. (2013). Immunogenicity and contraceptive efficacy of *Escherichia coli*-expressed recombinant porcine zona pellucida proteins. *American Journal of Reproductive Immunology* **70**, 139–152. doi:10.1111/aji. 12095
- Hall-Woods, M. L., Bauman, K. L., Bauman, J. E., Fischer, M., Houston, E. W., and Asa, C. S. (2007). Melengestrol acetate implant contraception

in addax (*Addax nasomaculatus*) and Arabian oryx (*Oryx leucoryx*). Zoo Biology **26**, 299–310. doi:10.1002/zoo.20146

- Herbert, C. A., and Trigg, T. E. (2005). Applications of GnRH in the control and management of fertility in female animals. *Animal Reproduction Science* 88, 141–153. doi:10.1016/j.anireprosci.2005.05.007
- Herbert, C. A., Trigg, T. E., Renfree, M. B., Shaw, G., Eckery, D. C., and Cooper, D. W. (2005). Long-term effects of deslorelin implants on reproduction in the female tammar wallaby (*Macropus eugenii*). *Reproduction* **129**, 361–369. doi:10.1530/rep.1.00432
- Hernandez, S., Locke, S. L., Cook, M. W., Harveson, L. A., Davis, D. S., Lopez, R. R., Silvy, N. J., and Fraker, M. A. (2006). Effects of SpayVac[®] on urban female white-tailed deer movements. *Wildlife Society Bulletin* 34, 1430–1434. doi:10.2193/0091-7648(2006)34[1430:EOSOUF]2.0. CO;2
- Hobbs, N. T., Bowden, D. C., and Baker, D. L. (2000). Effects of fertility control on populations of ungulates: general, stage-structured models. *The Journal of Wildlife Management* 64, 473–491. doi:10.2307/3803245
- Holland, O. J., Cowan, P. E., Gleeson, D. M., Duckworth, J. A., and Chamley, L. W. (2009). MHC haplotypes and response to immunocontraceptive vaccines in the brushtail possum. *Journal of Reproductive Immunology* 82, 57–65. doi:10.1016/j.jri.2009.04.008
- Hone, J. (1992). Rate of increase and fertility control. *Journal of Applied Ecology* 29, 695–698. doi:10.2307/2404478
- Hone, J. (1995). Spatial and temporal aspects of vertebrate pest damage with emphasis on feral pigs. *Journal of Applied Ecology* **32**, 311–319. doi:10.2307/2405098
- Hone, J. (1999). On rate of increase (r): patterns of variation in Australian mammals and the implications for wildlife management. *Journal of Applied Ecology* 36, 709–718. doi:10.1046/j.1365-2664.1999.00439.x
- Hone, J. (2002). Feral pigs in Namadgi National Park, Australia: dynamics, impacts and management. *Biological Conservation* **105**, 231–242. doi:10.1016/S0006-3207(01)00185-9
- Hoyer, P. B., Devine, P. J., Hu, X., Thompson, K. E., and Sipes, I. G. (2001). Ovarian toxicity of 4-vinylcyclohexene diepoxide: a mechanistic model. *Toxicologic Pathology* 29, 91–99. doi:10.1080/019262301301 418892
- Hu, X., Roberts, J. R., Apopa, P. L., Kan, Y. W., and Ma, Q. (2006). Accelerated ovarian failure induced by 4-vinyl cyclohexene diepoxide in Nrf2 null mice. *Molecular and Cellular Biology* 26, 940–954. doi:10.1128/MCB.26.3.940-954.2006
- Huchard, E., Canale, C. I., Le Gros, C., Perret, M., Henry, P.-Y., and Kappeler, P. M. (2012). Convenience polyandry or convenience polygyny? Costly sex under female control in a promiscuous primate. *Proceedings. Biological Sciences* 279, 1371–1379. doi:10.1098/rspb.2011.1326
- Humphrys, S., and Lapidge, S. J. (2008). Delivering and registering speciestailored oral antifertility products: a review. *Wildlife Research* 35, 578–585. doi:10.1071/WR07145
- Jacob, J., Herawati, N. A., Davis, S. A., and Singleton, G. R. (2004). The impact of sterilised females on enclosed populations of ricefield rats. *The Journal of Wildlife Management* 68, 1130–1137. doi:10.2193/0022-541X (2004)068[1130:TIOSFO]2.0.CO;2
- Jacob, J., Singleton, G. R., and Hinds, L. A. (2008). Fertility control of rodent pests. Wildlife Research 35, 487–493. doi:10.1071/WR07129
- Jacobsen, N. K., Jessup, D. A., and Kesler, D. J. (1995). Contraception in captive black-tailed deer by remotely delivered norgestomet ballistic implants. *Wildlife Society Bulletin* 23, 718–722.
- Jewell, P. (1986). Survival in a feral population of primitive sheep in St Kilda, Outer Hebrides, Scotland. *National Geographic Research* 2, 402–406.
- Ji, W. H. (2009). A review of the potential of fertility control to manage brushtail possums in New Zealand. *Human–Wildlife Conflicts* 3, 20–29.
- Ji, W. H., Clout, M. N., and Sarre, S. D. (2000). Responses of male brushtail possums to sterile females: implications for biological control. *Journal of Applied Ecology* 37, 926–934. doi:10.1046/j.1365-2664.2000.00546.x

- Johnson, E., and Tait, A. J. (1983). Prospects for the chemical control of reproduction in the grey squirrel. *Mammal Review* 13, 167–172. doi:10.1111/j.1365-2907.1983.tb00278.x
- Jones, J. E., Solis, J., Hughes, B. L., Castaldo, D. J., and Toler, J. E. (1990). Production and egg-quality responses of white leghorn layers to anticoccidial agents. *Poultry Science* 69, 378–387. doi:10.3382/ ps.0690378
- Kadir, Z., Ma, X., Li, J., and Zhang, F. (2013). Granulocyte–macrophage colony-stimulating factor enhances the humoral immune responses of mouse zona pellucida 3 vaccine strategy based on DNA and protein coadministration in BALB/c mice. *Reproductive Sciences* 20, 400–407. doi:10.1177/1933719112459236
- Kemp, J., and Miller, L. A. (2008). Oral vaccination and immunocontraception of feral swine using *Brucella suis* with multimeric GnRH protein expression. In 'Proceedings of 23rd Vertebrate Pest Conference'. (Eds R. M. Timm and M. B. Madon.) pp. 250–252. (University of California: Davis, CA.)
- Kennis, J., Sluydts, V., Leirs, H., and van Hooft, W. F. P. (2008). Polyandry and polygyny in an African rodent pest species, *Mastomys natalensis*. *Mammalia* 72, 150–160. doi:10.1515/MAMM.2008.025
- Khan, M. A. H., Ogita, K., Ferro, V. A., Kumasawa, K., Tsutsui, T., and Kimura, T. (2008). Immunisation with a plasmid DNA vaccine encoding gonadotrophin releasing hormone (GnRH-I) and T-helper epitopes in saline suppresses rodent fertility. *Vaccine* 26, 1365–1374. doi:10.1016/ j.vaccine.2007.12.052
- Killian, G. J., and Miller, L. A. (2000). Behavioral observations and physiological implications for white-tailed deer treated with two different immunocontraceptives. In 'Proceedings of the 9th Wildlife Damage Management Conference'. (Eds M. C. Brittingham, J. Kays and R. McPeake.) pp. 283–291. (Pennsylvania State University: State College, PA.)
- Killian, G., Miller, L. A., Rhyan, J., and Doten, H. (2006). Immunocontraception of Florida feral swine with a single-dose GnRH vaccine. *American Journal of Reproductive Immunology* 55, 378–384. doi:10.1111/j.1600-0897.2006.00379.x
- Killian, G., Thain, D., Diehl, N. K., Rhyan, J., and Miller, L. A. (2008). Four-year contraception rates of mares treated with single-injection porcine zona pellucida and GnRH vaccines and intrauterine devices. *Wildlife Research* 35, 531–539. doi:10.1071/WR07134
- Killian, G., Kreeger, T. J., Rhyan, J., Fagerstone, K., and Miller, L. A. (2009). Observations on the use of Gonacon[™] in captive female elk (*Cervus elaphus*). Journal of Wildlife Diseases 45, 184–188. doi:10.7589/0090-3558-45.1.184
- Kirkpatrick, J. F. (2007). Measuring the effects of wildlife contraception: the argument for comparing apples with oranges. *Reproduction, Fertility and Development* 19, 548–552. doi:10.1071/RD06163
- Kirkpatrick, J. F., and Franck, K. M. (2005). Contraception in free-ranging wildlife. In 'Wildlife Contraception: Issues, Methods and Applications'. (Eds C. S. Asa and I. J.Porton.) pp. 195–221 (The Johns Hopkins University Press: Baltimore, MD.)
- Kirkpatrick, J. F., and Turner, J. W. (1991). Reversible fertility control in nondomestic animals. *Journal of Zoo and Wildlife Medicine* 22, 392–408.
- Kirkpatrick, J. F., and Turner, A. (2002). Reversibility of action and safety during pregnancy of immunization against porcine zona pellucida in wild mares (*Equus caballus*). *Reproduction (Cambridge, England)* 60, 197–202.
- Kirkpatrick, J. F., and Turner, A. (2007). Immunocontraception and increased longevity in equids. *Zoo Biology* 26, 237–244. doi:10.1002/zoo.20109
- Kirkpatrick, J. F., and Turner, A. (2008). Achieving population goals in longlived wildlife with contraception. *Wildlife Research* 35, 513–519. doi:10.1071/WR07106
- Kirkpatrick, J. F., Rowan, A., Lamberski, N., Wallace, R., Frank, K., and Lyda, R. (2009). The practical side of immunocontraception: zona

proteins and wildlife. *Journal of Reproductive Immunology* **83**, 151–157. doi:10.1016/j.jri.2009.06.257

- Kirkpatrick, J. F., Lyda, R. O., and Frank, K. M. (2011). Contraceptive vaccines for wildlife: a review. *American Journal of Reproductive Immunology* 66, 40–50. doi:10.1111/j.1600-0897.2011.01003.x
- Kitchener, A. L., Kay, D. J., Walters, B., Menkhorst, P., McCartney, C. A., Buist, J. A., Mate, K. E., and Rodger, J. C. (2009). The immune response and fertility of koalas (*Phascolarctos cinereus*) immunised with porcine zonae pellucidae or recombinant brushtail possum ZP3 protein. *Journal* of Reproductive Immunology 82, 40–47. doi:10.1016/j.jri.2009.07.001
- Kreeger, T. J. (1997). Overview of delivery systems for the administration of contraceptives to wildlife. In 'Contraception in Wildlife Management. USDA–APHIS Technical Bulletin 1853'. (Ed. T. J. Kreeger.) pp. 29–48. (USDA–APHIS: Washington, DC.)
- Lambert, M. S., Massei, G., Yoder, C. A., and Cowan, D. P. (2010). An evaluation of Diazacon as a potential contraceptive in non-native roseringed parakeets. *The Journal of Wildlife Management* 74, 573–581. doi:10.2193/2008-531
- Levy, J. K. (2011). Contraceptive vaccines for the humane control of community cat populations. *American Journal of Reproductive Immunology* 66, 63–70. doi:10.1111/j.1600-0897.2011.01005.x
- Levy, J. K., Friary, J. A., Miller, L. A., Tucker, S. J., and Fagerstone, K. A. (2011). Long-term fertility control in female cats with GonaCon[™], a GnRH immunocontraceptive. *Theriogenology* **76**, 1517–1525. doi:10.1016/j.theriogenology.2011.06.022
- Liu, J., Jiang, Z., Liu, L., Zhang, Y., Zhang, S., Xiao, J., Ma, M., and Zhang, L. (2011). Triptolide induces adverse effect on reproductive parameters of female Sprague–Dawley rats. *Drug and Chemical Toxicology* 34, 1–7. doi:10.3109/01480541003774358
- Liu, M., Qu, J., Yang, M., Wang, Z., Wang, J., Zhang, Y., and Zhang, Z. (2012). Effects of quinestrol and levonorgestrel on populations of plateau pikas, *Ochotona curzoniae*, in the Qinghai–Tibetan Plateau. *Pest Management Science* 68, 592–601. doi:10.1002/ps.2302
- Locke, S. L., Cook, M. W., Harveson, L. A., Davis, D. S., Lopez, R. R., Silvy, N. J., and Fraker, M. A. (2007). Effectiveness of SpayVac[®] on reducing white-tailed deer fertility. *Journal of Wildlife Diseases* 43, 726–730. doi:10.7589/0090-3558-43.4.726
- Lohr, C. A., Mills, H., Robertson, H., and Bencini, R. (2009). Deslorelin implants control fertility in urban brushtail possums (*Trichosurus vulpecula*) without negatively influencing their body-condition index. *Wildlife Research* 36, 324–332. doi:10.1071/WR08050
- Lyda, R. O., Hall, R., and Kirkpatrick, J. F. (2005). A comparison of Freund's complete and Freund's modified adjuvants used with a contraceptive vaccine in wild horses (*Equus caballus*). Journal of Zoo and Wildlife Medicine 36, 610–616. doi:10.1638/04104.1
- Ma, X., Li, J., and Zhang, F. (2012). Intranasal co-delivery with the mouse zona pellucida 3 and GM–CSF expressing constructs enhances humoral immune responses and contraception in mice. *Scandinavian Journal of Immunology* **76**, 521–527. doi:10.1111/j.1365-3083.2012.02765.x
- Magiafoglou, A., Schiffer, M., Hoffmann, A. A., and McKechni, S. W. (2003). Immunocontraception for population control: will resistance evolve? *Immunology and Cell Biology* 81, 152–159. doi:10.1046/ j.0818-9641.2002.01146.x
- Massei, G. (2012). Catch, inject and release: immunocontraception as alternative to culling and surgical sterilisation to control rabies in freeroaming dogs. In 'Compendium of the Office International Epizooties Global Conference on Rabies Control'. (Eds A. R. Fooks and T. Muller.) pp. 181–187. (Office International Epizooties: Paris.)
- Massei, G. (2013). Fertility control in dogs. In 'Dogs, Zoonoses and Public Health'. (Eds C. N. Macpherson, F. X. Meslin and A. I. Wandeler.) pp. 259–270. (CABI International: Wallingford, UK.)
- Massei, G., and Miller, L. A. (2013). Non-surgical fertility control for managing free-roaming dog populations: a review of products and

criteria for field applications. *Theriogenology* **80**, 829–838. doi:10.1016/j.theriogenology.2013.07.016

- Massei, G., Cowan, D. P., Coats, J., Gladwell, F., Lane, J. E., and Miller, L. A. (2008). Effect of the GnRH vaccine GonaCon[™] on the fertility, physiology and behaviour of wild boar. *Wildlife Research* 35, 540–547. doi:10.1071/WR07132
- Massei, G., Quy, R., Gurney, J., and Cowan, D. P. (2010a). Can translocations be used to manage human–wildlife conflicts? *Wildlife Research* 37, 428–439. doi:10.1071/WR08179
- Massei, G., Miller, L. A., and Killian, G. J. (2010b). Immunocontraception to control rabies in dog populations. *Human–Wildlife Interactions* 4, 155–157.
- Massei, G., Coats, J., Quy, R., Storer, K., and Cowan, D. P. (2010c). The Boar-Operated-System: a novel method to deliver baits to wild pigs. *The Journal of Wildlife Management* 74, 333–336. doi:10.2193/2008-489
- Massei, G., Cowan, D. P., Coats, J., Bellamy, F., Quy, R., Brash, M., and Miller, L. A. (2012). Long-term effects of immunocontraception on wild boar fertility, physiology and behaviour. *Wildlife Research* 39, 378–385. doi:10.1071/WR11196
- Mayer, L. P., Pearsall, N. A., Christian, P. J., Devine, P. J., Payne, C. M., McCuskey, M. K., Marion, S. L., Sipes, I. G., and Hoyer, P. B. (2002). Long-term effects of ovarian follicular depletion in rats by 4-vinylcyclohexene diepoxide. *Reproductive Toxicology* 16, 775–781. doi:10.1016/S0890-6238(02)00048-5
- Mayer, L. P., Devine, P. J., Dyer, C. A., and Hoyer, P. B. (2004). The follicledeplete mouse ovary produces androgen. *Biology of Reproduction* 71, 130–138. doi:10.1095/biolreprod.103.016113
- Mayle, B. A., Ferryman, M., Peace, A., Yoder, C. A., Miller, L., and Cowan, D. P. (2013). The use of DiazaCon[™] to limit fertility by reducing serum cholesterol in female grey squirrels, *Sciurus carolinensis. Pest Management Science* 69, 414–424. doi:10.1002/ps.3347
- McLaughlin, E. A., and Aitken, R. J. (2011). Is there a role for immunocontraception? *Molecular and Cellular Endocrinology* 335, 78–88. doi:10.1016/j.mce.2010.04.004
- McLeod, S. R., and Saunders, G. (2014). Fertility control is much less effective than lethal baiting for controlling foxes. *Ecological Modelling* 273, 1–10. doi:10.1016/j.ecolmodel.2013.10.016
- McLeod, S. R., Saunders, G., Twigg, L. E., Arthur, A. D., Ramsey, D., and Hinds, L. A. (2007). Prospects for the future: is there a role for virally vectored immunocontraception in vertebrate pest management? *Wildlife Research* 34, 555–566. doi:10.1071/WR07050
- Merrill, J. A., Cooch, E. G., and Curtis, P. D. (2003). Time to reduction: factors influencing management efficacy in sterilizing overabundant white-tailed deer. *The Journal of Wildlife Management* 67, 267–279. doi:10.2307/ 3802768
- Merrill, J. A., Cooch, E. G., and Curtis, P. D. (2006). Managing an overabundant deer population by sterilization: effects of immigration, stochasticity and the capture process. *The Journal of Wildlife Management* 70, 268–277. doi:10.2193/0022-541X(2006)70[268: MAODPB]2.0.CO;2
- Middleton, D. R., Walters, B., Menkhorst, P., and Wright, P. (2003). Fertility control in the koala, *Phascolarctos cinereus*: the impact of slowrelease implants containing levonorgestrel or oestradiol on the production of pouch young. *Wildlife Research* **30**, 207–212. doi:10.1071/WR02052
- Miller, L. A., and Killian, G. J. (2002). In search of the active PZP epitope in white-tailed deer immunocontraception. *Vaccine* 20, 2735–2742. doi:10.1016/S0264-410X(02)00195-0
- Miller, L. A., Johns, B. E., Elias, D. J., and Killian, G. K. (1999). Oral vaccination of white-tailed deer using a recombinant bacillus Calmette–Guerin vaccine expressing the *Borrelia burgdorferi* outer surface protein A: prospects for immunocontraception. *American Journal of Reproductive Immunology* **41**, 279–285. doi:10.1111/ j.1600-0897.1999.tb00439.x

- Miller, L. A., Johns, B. E., and Killian, G. J. (2000). Immunocontraception of white-tailed deer with GnRH vaccine. *American Journal of Reproductive Immunology* 44, 266–274. doi:10.1111/j.8755-8920.2000.440503.x
- Miller, L. A., Crane, K., Gaddis, S., and Killian, G. J. (2001). Porcine zona pellucida immunocontraception: long-term health effects on white-tailed deer. *The Journal of Wildlife Management* 65, 941–945. doi:10.2307/ 3803042
- Miller, L. A., Rhyan, J. C., and Drew, M. (2004). Contraception of bison by GnRH vaccine: a possible means of decreasing transmission of brucellosis in bison. *Journal of Wildlife Diseases* **40**, 725–730. doi:10.7589/0090-3558-40.4.725
- Miller, L. A., Gionfriddo, J. P., Fagerstone, K. A., Rhyan, J. C., and Killian, G. J. (2008a). The single-shot GnRH immunocontraceptive vaccine (GonaConTM) in white-tailed deer: comparison of several GnRH preparations. *American Journal of Reproductive Immunology* 60, 214–223. doi:10.1111/j.1600-0897.2008.00616.x
- Miller, L. A., Gionfriddo, J. P., Rhyan, J. C., Fagerstone, K. A., Wagner, D. C., and Killian, G. J. (2008b). GnRH immunocontraception of male and female white-tailed deer fawns. *Human–Wildlife Interactions* 2, 93–101.
- Miller, L. A., Fagerstone, K. A., Wagner, D. C., and Killian, G. J. (2009). Factors contributing to the success of a single-shot, multiyear PZP immunocontraceptive vaccine for white-tailed deer. *Human–Wildlife Conflicts* 3, 103–115.
- Moore, H. D., Jenkins, N. M., and Wong, C. (1997). Immunocontraception in rodents: a review of the development of a sperm-based immunocontraceptive vaccine for the grey squirrel (*Sciurus* carolinensis). Reproduction, Fertility and Development 9, 125–129. doi:10.1071/R96053
- Moresco, A., Munson, L., and Gardner, I. A. (2009). Naturally occurring melengestrol acetate-associated reproductive tract lesions in zoo canids. *Veterinary Pathology* 46, 1117–1128. doi:10.1354/vp.08-VP-0293-M-FL
- Munson, L. (2006). Contraception in felids. *Theriogenology* 66, 126–134. doi:10.1016/j.theriogenology.2006.03.016
- Munson, L., Harrenstien, L. A., Acton, A. E., Graham, P. A., Chassy, L. M., and Kirkpatrick, J. F. (2005). Immunologic responses and adverse reactions to Freund's-adjuvanted porcine zona pellucida immunocontraceptives in domestic cats. *Vaccine* 23, 5646–5654. doi:10.1016/ j.vaccine.2005.05.044
- Nash, P., Furcolow, C. A., Bynum, K. S., Yoder, C. A., Miller, L. A., and Johnston, J. J. (2007). 20, 25-diazacholesterol as an oral contraceptive for blacktailed prairie dog population management. *Human–Wildlife Conflicts* 1, 60–67.
- Nave, C. D., Coulson, G., Short, R. V., Poiani, A., Shaw, G., and Renfree, M. B. (2002a). Long-term fertility control in the kangaroo and the wallaby using levonorgestrel implants. *Reproduction* 60, 71–80.
- Nave, C. D., Coulson, G., Poiani, A., Shaw, G., and Renfree, M. B. (2002b). Fertility control in the eastern grey kangaroo using levonorgestrel implants. *The Journal of Wildlife Management* 66, 470–477. doi:10.2307/3803180
- Naz, R. K. (2005). Search for peptide sequences involved in human antisperm antibody-mediated male immunoinfertility by using phage display technology. *Molecular Reproduction and Development* 72, 25–30. doi:10.1002/mrd.20315
- Naz, R. K. (2011). Contraceptive vaccines: success, status, and future perspectives. *American Journal of Reproductive Immunology* 66, 2–4. doi:10.1111/j.1600-0897.2011.00999.x
- Naz, R. K., Gupta, S. K., Gupta, J. C., Vyas, H. K., and Talwar, A. G. (2005). Recent advances in contraceptive vaccine development: a mini-review. *Human Reproduction* 20, 3271–3283. doi:10.1093/humrep/dei256
- Nett, T. M., Glode, L. M., and Ball, B. A. (2003). Evaluation of GnRH conjugated to a cytotoxic agent as a reproductive sterilant in mammals. In 'Managing African elephant Populations: Act or Let Die? Proceedings of an Expert Consultation on the Control of Wild Elephant Populations'.

(Eds B. Colenbrander, J. de Gooijer, R. Paling, S. Stout, T. Stout, and T. Allen) pp. 57–58. (Faculty of Veterinary Medicine, Utrecht University: Utrecht, The Netherlands.)

- Nettles, V. F. (1997). Potential consequences and problems with wildlife contraceptives. *Reproduction, Fertility and Development* 9, 137–143. doi:10.1071/R96054
- Nuñez, C. M. V., Adelman, J. S., Mason, C., and Rubenstein, D. I. (2009). Immunocontraception decreases group fidelity in a feral horse population during the non-breeding season. *Applied Animal Behaviour Science* **117**, 74–83. doi:10.1016/j.applanim.2008.12.001
- Nuñez, C. M. V., Adelman, J. S., and Rubenstein, D. I. (2010). Immunocontraception in wildwild horses (*Equus caballus*) extends reproductive cycling beyond the normal breeding season. *PLoS ONE* 5, e13635. doi:10.1371/journal.pone.0013635
- Patton, M. L., Jochle, W., and Penfold, L. M. (2007). Review of contraception in ungulate species. Zoo Biology 26, 311–326. doi:10.1002/zoo.20154
- Perdok, A. A., de Boer, W. F., and Stout, T. A. E. (2007). Prospects for managing African elephant population growth by immunocontraception: a review. *Pachyderm* 42, 97–107.
- Plotka, E. D., and Seal, U. S. (1989). Fertility control in female white-tailed deer. *Journal of Wildlife Diseases* 25, 643–646. doi:10.7589/0090-3558-25.4.643
- Poiani, A., Coulson, G., Salamon, D., Holland, S., and Nave, C. D. (2002). Fertility control of eastern grey kangaroos: do levonorgestrel implants affect behavior? *The Journal of Wildlife Management* 66, 59–66. doi:10.2307/3802871
- Powers, J. G., Baker, D. L., Davis, T. L., Conner, M. M., Lothridge, A. H., and Nett, T. M. (2011). Effects of gonadotropin-releasing hormone immunization on reproductive function and behavior in captive female Rocky Mountain elk (*Cervus elaphus nelsoni*). *Biology of Reproduction* 85, 1152–1160. doi:10.1095/biolreprod.110.088237
- Purswell, B. J., and Kolster, K. A. (2006). Immunocontraception in companion animals. *Theriogenology* 66, 510–513. doi:10.1016/ j.theriogenology.2006.04.018
- Ramsey, D. (2005). Population dynamics of brushtail possums subject to fertility control. *Journal of Applied Ecology* **42**, 348–360. doi:10.1111/ j.1365-2664.2005.01006.x
- Ramsey, D. (2007). Effects of fertility control on behavior and disease transmission in brushtail possums. *The Journal of Wildlife Management* 71, 109–116. doi:10.2193/2005-699
- Ramsey, D., and Efford, M. G. (2010). The effect of fertility control on the transmission of bovine tuberculosis in wild brushtail possums. *Journal of Applied Ecology* 47, 911–919. doi:10.1111/j.1365-2664.2010.01839.x
- Ransom, J. I., Cade, B. S., and Hobbs, N. T. (2010). Influences of immunocontraception on time budgets, social behavior, and body condition in feral horses. *Applied Animal Behaviour Science* 124, 51–60. doi:10.1016/j.applanim.2010.01.015
- Ransom, J. I., Roelle, J. E., Cade, B. S., Coates-Markle, L., and Kane, A. J. (2011). Foaling rates in feral horses treated with the immunocontraceptive porcine zona pellucida. *Wildlife Society Bulletin* 35, 343–352. doi:10.1002/wsb.66
- Reynolds, J. C., Short, M. J., and Leigh, R. J. (2004). Development of population control strategies for mink *Mustela vison*, using floating rafts as monitors and trap sites. *Biological Conservation* **120**, 533–543. doi:10.1016/j.biocon.2004.03.026
- Robinson, A. J., Jackson, R., Kerr, P., Merchant, J., Parer, I., and Pech, R. (1997). Progress towards using recombinant myxoma virus as a vector for fertility control in rabbits. *Reproduction, Fertility and Development* 9, 77–83. doi:10.1071/R96067
- Rudolph, B. A., Porter, W. F., and Underwood, H. B. (2000). Evaluating immunocontraception for managing suburban white-tailed deer in Irondequoit, New York. *The Journal of Wildlife Management* 64, 463–473. doi:10.2307/3803244

- Rutberg, A. T. (2005). Deer contraception: what we know and what we don't. In 'Humane Wildlife Solutions: the Role of Immunocontraception'. (Ed. A. T. Rutberg.) pp. 23–42. (Humane Society Press: Washington, DC.)
- Rutberg, A. T., and Naugle, R. E. (2008). Population effects of immunocontraception in white-tailed deer (*Odocoileus virginianus*). *Wildlife Research* 35, 494–501. doi:10.1071/WR07128
- Rutberg, A. T., Naugle, R. E., Thiele, L. A., and Liu, I. K. M. (2004). Effects of immunocontraception on a suburban population of white-tailed deer *Odocoileus virginianus. Biological Conservation* **116**, 243–250. doi:10.1016/S0006-3207(03)00195-2
- Rutberg, A. T., Naugle, R. E., Turner, J. W., Fraker, M. A., and Flanagan, D. R. (2013). Field testing of single-administration porcine zona pellucida contraceptive vaccines in white-tailed deer (*Odocoileus virginianus*). *Wildlife Research* 40, 281–288. doi:10.1071/WR12117
- Sachs, B. A., and Wolfman, L. (1965). 20, 25-diazacholestenol dihydrochloride: inhibition of cholesterol biosynthesis in hyperlipemic subjects. *Archives of Internal Medicine* **116**, 366–372. doi:10.1001/ archinte.1965.03870030046009
- Samoylova, T. I., Cochran, A. M., Samoylov, A. M., Schemera, B., Breiteneicher, A. H., Ditchkoff, S. S., Petrenko, V. A., and Cox, N. R. (2012). Phage display allows identification of zona pellucida-binding peptides with species-specific properties: novel approach for development of contraceptive vaccines for wildlife. *Journal of Biotechnology* 162, 311–318. doi:10.1016/j.jbiotec.2012.10.006
- Saunders, G., Mcilroy, J., Berghout, M., Kay, B., Gifford, E., Perry, R., and Van De Ven, R. (2002). The effects of induced sterility on the territorial behaviour and survival of foxes. *Journal of Applied Ecology* **39**, 56–66. doi:10.1046/j.1365-2664.2002.00696.x
- Seamark, R. F. (2001). Biotech prospects for the control of introduced mammals in Australia. *Reproduction, Fertility and Development* 13, 705–711. doi:10.1071/RD01073
- Sharma, S., and Hinds, L. A. (2012). Formulation and delivery of vaccines: ongoing challenges for animal management. *Journal of Pharmacy and Bioallied Sciences* 4, 258–266. doi:10.4103/0975-7406.103231
- Sharp, T., and Saunders, G. (2008). 'A Model for Assessing the Relative Humaneness of Pest Animal Control Methods.' (Australian Government Department of Agriculture, Fisheries and Forestry: Canberra.)
- Shi, D. Z., Wan, X. R., Davis, S. A., Pech, R. P., and Zhang, Z. B. (2002). Simulation of lethal control and fertility control in a demographic model for Brandt's vole *Microtus brandti. Journal of Applied Ecology* 39, 337–348. doi:10.1046/j.1365-2664.2002.00716.x
- Sibly, R. M., and Hone, J. (2002). Population growth rate and its determinants: an overview. *Philosophical Transactions of the Royal Society of London Series Biological Sciences* 357, 1153–1170. doi:10.1098/rstb.2002.1117
- Sinclair, A. R. E. (2003). Mammal population regulation, keystone processes and ecosystem dynamics. *Philosophical Transactions of the Royal Society London Series B* 358, 1729–1740. doi:10.1098/rstb.2003.1359
- Singla, N., Kaur, G., Babbar, B. K., and Sandhu, B. S. (2013). Potential of triptolide in reproductive management of the house rat, *Rattus rattus*. *Integrative Zoology* 8, 260–276. doi:10.1111/1749-4877.12013
- Singleton, G. R., Farroway, L. N., Chambers, L. K., Lawson, M. A., Smith, A. L., and Hinds, L. A. (2002). Ecological basis for fertility control in the house mouse (*Mus domesticus*) using immunocontraceptive vaccines. *Reproduction* **60**, 31–39.
- Smith, G. C., and Cheeseman, C. L. (2002). A mathematical model for the control of diseases in wildlife populations: culling, vaccination and fertility control. *Ecological Modelling* **150**, 45–53. doi:10.1016/S0304-3800(01)00471-9
- Smith, G. C., and Wilkinson, D. (2003). Modeling control of rabies outbreaks in red fox populations to evaluate culling, vaccination, and vaccination combined with fertility control. *Journal of Wildlife Diseases* 39, 278–286. doi:10.7589/0090-3558-39.2.278

- Swinton, J., Tuyttens, F., MacDonald, D., Nokes, D. J., Cheeseman, C. L., and Clifton-Hadley, R. (1997). A comparison of fertility control and lethal control of bovine tuberculosis in badgers: the impact of perturbation induced transmission. *Philosophical Transactions of the Royal Society* of London. Series B, Biological Sciences 352, 619–631. doi:10.1098/ rstb.1997.0042
- Temple, J. L., Millar, R. P., and Rissman, E. F. (2003). An evolutionarily conserved form of gonadotropin-releasing hormone coordinates energy and reproductive behavior. *Endocrinology* 144, 13–19. doi:10.1210/ en.2002-220883
- Totton, S. C., Wandeler, A. I., Zinsstag, J., Bauch, C. T., Ribble, C. S., Rosatte, R. C., and McEwen, S. A. (2010). Stray dog population demographics in Jodhpur, India following a population control/rabies vaccination program. *Preventive Veterinary Medicine* 97, 51–57. doi:10.1016/j.prevetmed. 2010.07.009
- Tran, T. T., and Hinds, L. A. (2013). Fertility control of rodent pests: a review of the inhibitory effects of plant extracts on ovarian function. *Pest Management Science* 69, 342–354. doi:10.1002/ps.3354
- Turner, A., and Kirkpatrick, J. F. (2002). Effects of immunocontraception on population, longevity and body condition in wild mares *Equus caballus*. *Reproduction* **60**(Suppl.), 187–195.
- Turner, J. W., Liu, I. K. M., and Kirkpatrick, J. F. (1996). Remotely delivered immunocontraception in free-roaming feral burros (*Equus asinus*). *Journal of Reproduction and Fertility* **107**, 31–35. doi:10.1530/ jrf.0.1070031
- Turner, J. W., Flanagan, D. R., Rutberg, A. T., and Kirkpatrick, J. F. (2007). Immunocontraception in wild horses: one inoculation provides two years of infertility. *The Journal of Wildlife Management* **71**, 662–667. doi:10.2193/2005-779
- Turner, J. W., Rutberg, A. T., Naugle, R. E., Kaur, M. A., Flanagan, D. R., Bertschinger, H. J., and Liu, I. K. M. (2008). Controlled-release components of PZP contraceptive vaccine extend duration of infertility. *Wildlife Research* 35, 555–562. doi:10.1071/WR07159
- Tuyttens, F. A. M., and Macdonald, D. W. (1998). Sterilization as an alternative strategy to control wildlife diseases: bovine tuberculosis in European badgers as a case study. *Biodiversity and Conservation* 7, 705–723. doi:10.1023/A:1008830418123
- Twigg, L. E., and Williams, C. K. (1999). Fertility control of overabundant species; can it work for feral rabbits? *Ecology Letters* 2, 281–285. doi:10.1046/j.1461-0248.1999.00085.x
- Twigg, L. E., Lowe, T. J., Martin, G. R., Wheeler, A. G., Gray, G. S., Griffin, S. L., O'Reilly, C. M., Robinson, D. J., and Hubach, P. H. (2000). Effects of surgically imposed sterility on free-ranging rabbit populations. *Journal of Applied Ecology* **37**, 16–39. doi:10.1046/ j.1365-2664.2000.00471.x
- Tyndale-Biscoe, H., and Hinds, L. A. (2007). Introduction virally vectored immunocontraception in Australia. *Wildlife Research* 34, 507–510. doi:10.1071/WRv34n7_IN
- Walcher, P., Cui, X., Arrow, J. A., Scobie, S., Molinia, F. C., Cowan, P. E., Lubitz, W., and Duckworth, J. A. (2008). Bacterial ghosts as a delivery system for zona pellucida-2 fertility control vaccines for brushtail possums (*Trichosurus vulpecula*). *Vaccine* 26, 6832–6838. doi:10.1016/j.vaccine. 2008.09.088

- Wheaton, C. J., Savage, A., Shukla, A., Neiffer, D., Qu, W., Sun, Y., and Lasley, B. L. (2011). The use of long acting subcutaneous levonorgestrel (LNG) gel depot as an effective contraceptive option for cotton-top tamarins (*Saguinus oedipus*). *Zoo Biology* **30**, 498–522. doi:10.1002/ zoo.20354
- White, P. C. L., and Ward, A. I. (2010). Interdisciplinary approaches for the management of existing and emerging human–wildlife conflicts. *Wildlife Research* 37, 623–629. doi:10.1071/WR10191
- White, P. C. L., Lewis, A. J. G., and Harris, S. (1997). Fertility control as a means of controlling bovine tuberculosis in badger (*Meles meles*) populations in south-west England: predictions from a spatial stochastic simulation model. *Proceedings. Biological Sciences* 264, 1737–1747. doi:10.1098/rspb.1997.0241
- Wilkinson, D., Bennett, R., McFarlane, I., Rushton, S., Shirley, M., and Smith, G. C. (2009). Cost–benefit analysis model of badger (*Meles meles*) culling to reduce cattle herd tuberculosis breakdowns in Britain, with particular reference to badger perturbation. *Journal of Wildlife Diseases* 45, 1062–1088. doi:10.7589/0090-3558-45.4.1062
- Williams, C. K. (2007). Assessment of the risk of inadvertently exporting from Australia a genetically modified immunocontraceptive virus in live mice (*Mus musculus domesticus*). Wildlife Research 34, 540–554. doi:10.1071/WR05028
- Williams, C. K., Davey, C. C., Moore, R. J., Hinds, L. A., Silvers, L. E., Kerr, P. J., French, N., Hood, G. M., Pech, R. P., and Krebs, C. J. (2007). Population responses to sterility imposed on female European rabbits. *Journal of Applied Ecology* 44, 291–301. doi:10.1111/j.1365-2664.2006. 01264.x
- Wood, C., Ballou, J. D., and Houle, C. S. (2001). Restoration of reproductive potential following expiration or removal of melengestrol acetate contraceptive implants in golden lion tamarins (*Leontopithecus rosalia*). Journal of Zoo and Wildlife Medicine **32**, 417–425.
- Xu, C.-K., and Zhao, Y.-H. (2010). Apoptosis of rat's ovarian follicle cells induced by triptolide *in vivo*. *African Journal of Pharmacy and Pharmacology* 4, 422–430.
- Yoder, C. A., and Miller, L. A. (2010). Effect of GonaCon[™] vaccine on black-tailed prairie dogs: immune response and health effects. *Vaccine* 29, 233–239. doi:10.1016/j.vaccine.2010.10.055
- Yoder, C., Andelt, W., Miller, L., Johnston, J., and Goodall, M. (2004). Effectiveness of twenty, twenty-five diazacholesterol, avian gonadotropin-releasing hormone, and chicken riboflavin carrier protein for inhibiting reproduction in *Coturnix* quail. *Poultry Science* 83, 234–244. doi:10.1093/ps/83.2.234
- Yoder, C. A., Avery, M. L., Keacher, K. L., and Tillman, E. A. (2007). Use of DiazaCon[™] as a reproductive inhibitor for monk parakeets (*Myiopsitta monachus*). Wildlife Research 34, 8–13. doi:10.1071/WR06069
- Yoder, C. A., Mayle, B. A., Furcolow, C. A., Cowan, D. P., and Fagerstone, K. A. (2011). Feeding of grey squirrels (*Sciurus carolinensis*) with the contraceptive agent DiazaCon[™]: effect on cholesterol, hematology, and blood chemistry. *Integrative Zoology* 6, 409–419. doi:10.1111/j.1749-4877.2011.00247.x
- Zhang, Z. (2000). Mathematical models of wildlife management by contraception. *Ecological Modelling* 132, 105–113. doi:10.1016/ S0304-3800(00)00308-2