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Communication

Contemplating 1,2,4-Thiadiazole-Inspired Cyclic Peptide Mimics: A Computational Investigation

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Marine derived cyclic peptides have inspired chemists for decades as the cavitand architecture can be compared with macrocyclic ligands, and hence easily conceived as mediators of metal-ion transport. Lissoclinamide 5 and ascidiacyclamide are two such cyclic peptides that have received much attention both for their metal ion complexation properties and biological activity; the metal ion binding properties of mimics of these two systems have been reported. Reported herein is a computational study aimed at evaluating the stability, and potential for copper(π) ion binding by lissoclinamide 5 mimics that substitute the naturally occurring 4-carboxy-1,3-thiazole units for novel valine- and phenylalanine-derived 1,2,4-thiadiazole units. Our results suggest that one lissoclinamide 5 mimic, 1,2,4-thiadiazole (TDA)-lissoclinamide 9, may be capable of forming a complex with one Cu^{II} ion, [Cu(9-H)(H₂O)]⁺. A complex with two Cu^{II} ions, [Cu₂(9-H)(μ -OH)]²⁺, was also considered. These results set the stage for synthetic and experimental metal binding studies.

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Cyclic peptides arising from the marine environment^[1–4] continue to attract considerable fascination prompted in part by marinebased pharmaceuticals currently being utilised in the clinic,^[5-8] which in turn is driving drug-discovery efforts,^[9-14] and substantial pursuits in understanding their chemical ecology.^[15,16] Prominent amongst these have been the azole-based cyclic peptides,^[17] with an archetypal cavitand appearance, that have led chemists to consider metal ion coordination and transport as a potential mode of biological activity and/or a role in electrontransfer processes.^[18–20] Potassium^[21] and other metal ions have been observed to form complexes with azole cyclic peptides, but the most amenable to coordination has been copper.[22-24] To explore the metal binding features of these systems further we recently introduced the concept of heteroatom-interchanged (HI) cyclic peptides.^[25,26] Lissoclinamide 5 (1)^[27–31] was investigated first as a working template, followed subsequently by ascidiacy-clamide (2), $[^{32-34]}$ which led to the synthesis of four HI-isomers (e.g. 3) of lissoclinamide 5 and one HI-isomer 4 of ascidiacyclamide (Fig. 1). Metal ion binding studies using mass spectrometry (MS), electronparamagnetic resonance (EPR), and density functional theory (DFT) revealed that the HI-lissoclinamides (e.g. 3) form complexes with one Cu^{II} ion, whereas the HI-ascidiacyclamide isomer 4 was a weaker ligand for Cu^{II},^[25,26] suggesting small changes can have a dramatic effect on co-ordination, conformation,^[35] and biological activity.^[25,35]

To enable the heteroatom-interchange concept, naturally occurring 4-carboxy-1,3-thiazole units (i.e. 5) were replaced by non-natural 5-carboxy-1,3-thiazole derivatives 6, which required new methodologies to be evaluated in pursuit of

obtaining suitable 5-carboxy-1,3-thiazole building blocks.^[25,26] Numerous synthetic methods to access 5-carboxy-1,3-thiazoles were evaluated, and in one case valine- and phenylalaninederived 1,2,4-thiadiazoles (i.e. 7 and 8) were produced unexpectedly. Given our interest and on-going efforts in the azole cyclic peptide arena we considered whether the 1,2,4-thiadiazole system would underpin next generation cyclic peptide mimic design. However, in light of the HI-isomers creating a considerable synthetic challenge, an *in silico* investigation to assist in determining a likely synthetic candidate was justified.

Reported herein is the synthesis and characterisation of novel 1,2,4-thiadiazoles 7 and 8, along with an *in silico* assessment of lissoclinamide 5 mimics 9-12 (Fig. 1) in terms of ground state stability and the propensity to coordinate Cu^{II} ions.

3,5-Disubstitiuted-1,2,4-thiadiazoles

As mentioned above, in the course of developing the synthesis of the HI-cyclic peptides (e.g. **3** and **4**),^[25,26] 5-carboxy-1,3-thiazole building blocks were required (e.g. **6**), which at the time demanded *de novo* synthetic routes to be developed. To satisfy the required functionality, and stereochemistry, any newly developed method needed to accommodate amino acid derived substrates for practicality reasons. The method reported by $Zhao^{[36]}$ was initially chosen for potential modification because it described that exposure of mono-substituted thioureas (e.g. **13**) to the in situ generated bromohydrin **14** (bromination of enol ether **15** with *N*-bromosuccinimide, NBS) afforded aminothiazoles (e.g. **16**) in good yields.^[37] Therefore, this method



Fig. 1. Lissoclinamide 5 (1) and ascidiacyclamide (2), HI-isomers 3 and 4, and thiazole building blocks 5 and 6 together with 1,2,4-thiadiazoles 7 and 8, and the corresponding four 1,2,4-thiadiazole lissoclinamide mimics 9–12.



Scheme 1. Modification of Zhao's method to access 5-carboxy-1,3-thiazole building blocks, which also produced 1,2,4-thiadiazole by-products.

thioamides **17** and **18** were readily accessible, $[^{38,39]}$ and this proved to be successful giving thiazoles **20** and **19**, respectively (Scheme 1). However, a by-product was observed in both cases, but in varying amounts. When using the phenylalanine thioamide **18**, thiazole **19** was obtained in 46 % yield, in addition to 9% of 1,2,4-thiadiazole **8**, whereas the valine thioamide **17** afforded thiazole **20** and thiadiazole **7** in a higher yield of 34% (Scheme 1). The latter 1,2,4-thiadiazole structure was confirmed by X-ray crystallographic analysis (Fig. 2).

A literature search revealed that 3,5-disubstituted-1,2,4thiadiazoles have been previously synthesised from thioamides, but importantly not those derived from amino acids. Takikawa et al. reported that treatment of a range of thioamides directly with NBS afforded 1,2,4-thiadiazoles in yields ranging from 72–93 %,^[40] which provided justification as to why 1,2,4-thiadiazoles were appearing as by-products in the modified Zhao synthesis of thiazoles.

Thiadiazole Cyclic Peptide Stability and Cu^{II} Binding

Given that methodology now existed to access 1,2,4-thiadiazoles derived from amino acids, and our interest in the lissoclinamide 5 system, mimics **9–12** were conceived (Fig. 1). In these cases,



Fig. 2. *ORTEP* diagram of the X-ray crystallographic structure of 1,2,4-thiadiazole 7 (30% ellipsoid probability).

however, the 1,2,4-thiadiazoles do not have a N and C terminus to enable all peptide bonds seen in in lissoclinamide 5 to be formed. Therefore, an aspect of the design (i.e. 9-12) features two secondary amines that link both the two 1,2,4-thiadiazole units and the N-terminus of the polypeptide (i.e. left-hand fragment), of which the latter polypeptide has been conserved throughout our entire HI studies (i.e. right hand fragment).^[25,26] These structures, which break traditional cyclic peptide mimicking seen previously (i.e. all peptide bonds conserved), are of interest considering the wealth of literature available with respect to the conformations and biological activity of similar cyclic peptides.[22-27] In addition, the metal complexes of these ligands, particularly the Cu^{II} complexes, have been studied as phosphoesterase and carbonic anhydrase mimics.^[22,24,41-43] We therefore chose to explore, computationally, peptides 9-12 in terms of their structure and the potential for Cu^{II} binding.

Peptides 9-12 each exhibit eight chiral centres. To gain insight into structural effects of these chiral centres computational modelling was undertaken using a force field-based approach involving the Macrocycle Conformational Sampling algorithm of MacroModel 10.6 in conjunction with the MMFFs force field.^[44,45] The minimum energy conformers for each peptide were identified and are shown in the Supplementary Material, although it should be noted that in each case some hundreds of possible conformations were identified. The most stable conformer of each peptide displayed the peptide NH groups and the nitrogen atoms of the oxazoline and 1,2,4thiadiazoles rings pointing towards the interior of the macrocycle. This orientation of the 1,2,4-thiadiazole rings was in contrast to that seen for the 'inverted' analogues 3 and 4 where the sulfur atoms of the thiazoles were pointed towards the centre of the cyclic peptide.^[25,26] A study of the relative energies of the most stable conformers of 9-12 was performed using the B3LYP/6-31G(d) DFT method.^[46,47] The spread of energies calculated for the lowest energy conformers of 9-12 was small, of the order of 6.3 kJ mol⁻¹, with **9** representing the lowest energy structure. The conformer studies suggested that these cyclic peptides may have the capacity to bind metal ions particularly, and in contrast to 3 and 4, as the potential donor N atom atoms are pointed towards the centre of the cavity.

Two models for putative Cu^{II} complexes were investigated, $[Cu(9-H)(H_2O)]^+$ and $[Cu_2(\mu-OH)(9-H)]^{2+}$; here 9-H represents the cyclic peptide deprotonated at the amide. Geometries of the two complexes were optimised with M06/6-31G(d)-LANL2DZ in an implicit (SMD) methanol solvent model.^[46,48,49] For the $[Cu_2(\mu-OH)(9-H)]^{2+}$ complex, the two Cu^{II} ions exhibited different coordination geometries. In one site a five-coordinate, square based pyramid Cu^{II} ion was coordinated through the nitrogen donors of a 1,2,4-thiadiazole (2.21 Å), the (R)-5-methyl-4,5-dihydrooxazole (2.16 Å), and the deprotonated amide (1.92 Å), as well as the oxygen of the carbonyl group adjacent to the pyrrolidine (2.42 Å) and the μ-OH (1.93 Å). The coordination sphere of the second, fourcoordinate, square planar Cu^{II} site, was composed of the nitrogen donors of two secondary amines (2.24 and 2.54 Å), the nitrogen of a second 1,2,4-thiadiazole (1.92 Å), and the coordination sphere completed by the μ -OH (1.87Å). The Cu-Cu distance was 3.05 Å with the Cu-OH-Cu angle 106.7°. For the $[Cu(9-H)(H_2O)]^+$ complex the coordination sphere was modelled as comprising the deprotonated amide (1.92 Å), the nitrogen donors of 1,2,4-thiadiazole (2.17 Å) and (R)-5methyl-4,5-dihydrooxazole (2.11 Å), and a long interaction with the carbonyl oxygen adjacent to the pyrrolidine (2.57 Å) and the H_2O (2.01 Å). The proposed structures are shown in Fig. 3 (and Fig. S2, Supplementary Material).

Interesting comparisons can be drawn between the above systems and the Cu^{II} complexes of the pseudo-octapeptide H_4pat^1 , an analogue of the patellamides (Fig. 4).^[50] H_4pat^1 contains four chiral centres and both $[Cu(H_3pat^1)(H_2O)_2]^+$ and $[Cu_2(H_2pat^1)(\mu-OH)(H_2O)_2]^+)$ complexes have been characterised through MS, DFT calculations and, for the latter, X-ray crystallography.^[50] For $[Cu_2(H_2pat^1)(\mu-OH)(H_2O)_2]^+$ the Cu···Cu separation was 3.76 Å with a Cu-OH-Cu angle of 136.8°; the other metal-donor distances were comparable in $[Cu_2(\mu-OH)(9-H)]^{2+}$ and $[Cu_2(H_2pat^1)(\mu-OH)(H_2O)_2]^+$. The structure of [Cu(H₃pat¹)(H₂O)₂]⁺, calculated with DFT, proposed a five-coordinate Cu^{II} site, the Cu^{II} coordinated through the nitrogen atoms of two 1,5-dimethylimidazoles and a deprotonated amide, with two water molecules completing the coordination sphere. The DFT analysis for $[Cu_2(H_2pat^{-1})]$ $(\mu$ -OH)(H₂O)₂]⁺ and [Cu(H₃pat¹)(H₂O)₂]⁺ suggested that the former complex was 25 kJ mol⁻¹ more stable than the latter.^[50]

The question then arises - do the computations reported herein provide an indication as to whether it is possible to prepare the $[Cu(9-H)(H_2O)]^+$ and $[Cu_2(\mu-OH)(9-H)]^{2+}$ complexes? In order to provide a clue as to this question, the energy required to reorganise the macrocyclic peptide from its ground-state geometry into its coordinated geometry was calculated for the two complexes. The calculations suggested that 12.5 kJ mol⁻¹ was required for **9-**H to reorganise to accommodate the formation of $[Cu(9-H)(H_2O)]^+$ whereas for the formation of the $[Cu_2(\mu\text{-OH})(9\text{-H})]^{2+}$ complex the energy requirement for the reorganisation of the peptide was larger, of the order of 71 kJ mol⁻¹. In addition, the shorter $Cu \cdots Cu$ distance and more acute Cu-OH-Cu angle calculated for $\left[\text{Cu}_2(\mu\text{-OH})(\textbf{9-H})\right]^{2+}$ (3.05 Å, 106.7°, respectively) compared with $[Cu_2(H_2pat^1)(\mu-OH)(H_2O)_2]^+$ (3.76 Å, 136.8°, respectively)^[50] suggest that the mono-Cu^{II} complex of cyclic peptide 9 may form preferentially.

In conclusion, this study has conceived novel cyclic peptides designed on the naturally occurring lissoclinamide 5 template, taking inspiration from preliminary synthetic studies that produced 1,2,4-thiadiazoles. Subsequently, these macrocyclic ring systems were explored in terms of determining both their ground state energies and the potential of the lowest energy conformer to bind copper(π). 1,2,4-Thiadiazole (TDA)-lissoclinamide **9** was found to have the highest likelihood of forming a mono-copper complex i.e. [Cu(**9**-H)(H₂O)]⁺, but not with two Cu^{II} ions. These combined results provide encouragement to initiate a synthetic program in this area and determine whether metal binding is observable.



Fig. 3. Calculated structures of (a) $[Cu_2(\mu-OH)(9-H)]^{2+}$ and (b) $[Cu(9-H)(H_2O)]^+$.



Fig. 4. H_4 pat¹ reported by Comba et al.^[50]

Experimental

Synthesis

General experimental procedures along with the synthetic procedures for **17–20** have been previously reported.^[25,26]

((1S,1'S)-(1,2,4-Thiadiazole-3,5-diyl)bis(2-methylpropane-1,1-diyl))dicarbamate (7) and Methyl 2-(1-[(tertbutoxycarbonyl)amino]-2-methylpropyl)thiazole-5carboxylate (**20**)

To a mixture of methyl *trans*-3-methoxyacrylate (**15**) (3.8 mL, 35.3 mmol) in water (18 mL) and dioxane (18 mL) at -10° C was added recrystallised NBS (6.9 g, 38.8 mmol). The reaction mixture was stirred at room temperature for 1 h. After cooling the reaction to 0°C, (*S*)-*N*-*tert*-(butoxycarbonyl)thiovalinamide (**17**) 8.2 g, 35.3 mmol) was added and the reaction allowed to warm to room temperature overnight. Concentrated NH₄OH (7 mL) was then added dropwise and the mixture stirred for 10 min. The resulting slurry was diluted with Et₂O (180 mL) and washed with water (3 × 360 mL). The Et₂O layer was dried with Na₂SO₄ and concentrated under vacuum. Flash chromatography (silica gel, petroleum ether/ethyl acetate 4 : 1, R_f 0.45) provided thiazole **20** as a white foam (4.6 g, 8 %), and thiadiazole **7** (1.3 g, 34 %). $\delta_{\rm H}$ (400 MHz, CDCl₃) 5.41 (d, *J* 9.0, 1H), 5.17 (d, *J* 8.3, 1H), 5.02 (m, 2H), 2.09 (m, 1H), 1.97 (m, 1H), 1.43 (s, 18H),

 $\begin{array}{l} 0.97 \ (d, J \ 6.8, \ 3H), \ 0.87 \ (d, J \ 6.9, \ 6H), \ 0.83 \ (d, J \ 6.8, \ 3H). \ \delta_C \\ (100 \ MHz, \ CDCl_3) \ 193.4, \ 176.2, \ 155.5, \ 155.3, \ 80.5, \ 79.5, \ 40.1, \\ 39.5, \ 28.3, \ 28.3, \ 24.9, \ 24.6, \ 15.4, \ 15.4, \ 11.6. \ m/z \ (ESI) \ 451.2364; \\ calcd \ for \ C_{20}H_{36}N_4O_4SNa^+ \ 451.2355. \end{array}$

((15,1'S)-(1,2,4-Thiadiazole-3,5-diyl)bis(2-benzyl-1,1diyl))dicarbamate (**8**) and Methyl 2-{1-[(tertbutoxycarbonyl)amino]-2-phenylethyl}thiazole-5carboxylate (**19**)

To a mixture of methyl trans-3-methoxyacrylate (15) (5.29 mL, 42.4 mmol) in water (21 mL) and dioxane (21 mL) at -10°C was added recrystallised NBS (8.33 g, 46.6 mmol). The reaction mixture was stirred at room temperature for 1 h. After cooling the reaction to 0° C, (S)-N-tert-(butoxycarbonyl)thiophenylalaninamide (18) (11.90 g, 42.42 mmol) was added and the reaction allowed to warm to room temperature overnight. Concentrated NH₄OH (8 mL) was then added dropwise and the mixture stirred for 10 min. The resulting slurry was diluted with Et₂O (200 mL) and washed with water $(3 \times 400 \text{ mL})$. The Et₂O layer was dried with Na₂SO₄ and concentrated under vacuum. Flash chromatography (silica gel, petroleum ether/ethyl acetate 4:1, $R_{\rm f}$ 0.61) provided methyl 2-{1-[(tert-butoxycarbonyl)amino]-2-phenylethyl}thiazole-5-carboxylate (19) as a white foam (5.7 g, 37 %), and thiadiazole 8 (1.4 g, 9%). $\delta_{\rm H}$ (400 MHz, CDCl₃) 7.29 –7.25 (m, 2H), 7.22 – 7.13 (m, 5H), 7.11 –7.03 (m, 2H), 6.94 (m, 1H), 5.37 (m, 1H), 5.23 (s, 1H), 3.27 (m, 4H), 1.41 (s, 9H), 1.39 (s, 9H). δ_C (100 MHz, CDCl₃) 193.8, 175.7, 155.0, 154.9, 136.5, 135.5, 129.5, 129.2, 128.7, 128.2, 127.2, 126.6, 80.6, 79.6, 53.7, 53.6, 52.8, 52.7, 41.1, 41.0, 29.6, 28.2. m/z (ESI) 547.2340; calcd for C₂₈H₃₆N₄O₄SNa⁺ 547.2457.

Calculations

The conformations of cyclic peptides were investigated using the Macrocycle Conformational Sampling algorithm of *MacroModel 10.6*.^[44,45] The conformer sampling employed the MMFFs (MMFF94s) force field and GB/SA (water) solvent model. The process utilised 5000 cycles of large-scale low mode

searches performed on a set of seed structures obtained from 10000 cycles of MD simulated annealing, with eigenvectors calculated for each new global minimum, and included the 'enhanced' option for torsional sampling of amide bonds (which samples the amide C–N and C–O single bonds). The lowest-energy conformers of **9–12** obtained from the MMFFs–GB/SA forcefield-based conformer sampling were reoptimised with DFT. The DFT calculations were performed with *Gaussian*

 $16.^{[46]}$ For the study of the relative energies of **9–12** the B3LYP functional was used in conjunction with the 6-31G(d) basis set, the studies undertaken in the gas phase.^[46–49] For the Cu^{II} complexes, the M06 functional in conjunction with a mixed basis set consisting of LANL2DZ on Cu and 6-31G(d) on other atoms was used.^[49] Methanol solvent was modelled with the SMD implicit model.^[46–49]

Crystal Data for 7

C₂₀H₃₆N₄O₄S: *M* 428.59, *T* 293(2) K, Monoclinic, space group *P*2₁, *a* 11.1529(3), *b* 10.1893(2), *c* 11.1550(3) Å, β 93.620(2)°, *V* 1265.13(5) Å³, *Z* 2, F(000) 464, *D*_c 1.125 g cm⁻³, μ (Cu-Kα) 1.375 mm⁻¹, 3989 unique data, *R*₁ 0.0331 (for 3738 observed reflections *I* > 2 σ (*I*)), w*R*₂ 0.0945 (all data).

Single crystal intensity data were collected on an Oxford Diffraction Gemini S Ultra CCD diffractometer using graphite monochromatic Cu-K α radiation (λ 1.5418 Å) for 1,2,4-thiadiazole 7 operating in the ω -scan mode. Data reduction and empirical absorption corrections were performed with the CrysAlis program (Oxford Diffraction, version 171.35.11), while all other computations were performed within the WinGX suite of programs.^[51] The structure was solved by direct methods with SHELXS and refined by full matrix least-squares analysis with SHELXL97.[52] All non-H atoms were refined with anisotropic thermal parameters, and H-atoms were constrained at estimated positions using a riding model. The absolute configuration was established by anomalous dispersion effects. The atomic nomenclature is defined in Fig. 2 and drawn with ORTEP3.^[53] Crystallographic data in CIF format is available from the Cambridge Crystallographic Data Base with CCDC deposition number 1903408.

Supplementary Material

Full crystal data, atomic coordinates, bond angles, bond lengths, and copies of ¹H and ¹³C NMR data are available on the Journal's website.

Conflicts of Interest

The authors declare no conflict of interest.

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References

- J. W. Blunt, B. R. Copp, R. A. Keyzers, M. H. G. Munro, M. R. Prinsep, Nat. Prod. Rep. 2015, 32, 116. doi:10.1039/C4NP00144C
- [2] D. J. Faulkner, Nat. Prod. Rep. 2002, 19, 1.

- [3] D. Skropeta, L. Wei, Nat. Prod. Rep. 2014, 31, 999. doi:10.1039/ C3NP70118B
- [4] J. W. Blunt, B. R. Copp, R. A. Keyzers, M. H. G. Munro, M. R. Prinsep, *Nat. Prod. Rep.* 2014, 31, 160. doi:10.1039/C3NP70117D
- [5] M. S. Butler, A. A. B. Robertson, M. A. Cooper, *Nat. Prod. Rep.* 2014, 31, 1612. doi:10.1039/C4NP00064A
- [6] T. L. Simmons, E. Andrianasolo, K. McPhail, P. Flatt, W. H. Gerwick, Mol. Cancer Ther. 2005, 4, 333.
- [7] S. A. Dyshlovoy, F. Honecker, *Mar. Drugs* 2015, 13, 5657. doi:10. 3390/MD13095657
- [8] M. Jaspars, D. De Pascale, J. H. Andersen, F. Reyes, A. Crawford, A. Ianora, J. Mar. Biol. Assoc. U. K. 2016, 96, 151. doi:10.1017/ S0025315415002106
- [9] V. A. Stonik, S. N. Fedorov, Mar. Drugs 2014, 12, 636. doi:10.3390/ MD12020636
- [10] Y. Zhou, Curr. Org. Chem. 2014, 18, 918. doi:10.2174/ 138527281807140515154736
- [11] P. Kiuru, M. V. D'Auria, C. D. Muller, P. Tammela, H. Vuorela, J. Yli-Kauhaluoma, *Planta Med.* 2014, 80, 1234.
- [12] L. A. Salvador-Reyes, H. Luesch, Nat. Prod. Rep. 2015, 32, 478. doi:10.1039/C4NP00104D
- [13] L. A. Salvador-Reyes, N. Engene, V. J. Paul, H. Luesch, J. Nat. Prod. 2015, 78, 486. doi:10.1021/NP500931Q
- [14] L. T. Tan, Drug Discov. Today 2013, 18, 863. doi:10.1016/J.DRUDIS. 2013.05.010
- [15] P. N. Leao, N. Engene, A. Antunes, W. H. Gerwick, V. Vasconcelos, *Nat. Prod. Rep.* 2012, 29, 372. doi:10.1039/C2NP00075J
- [16] J. Piel, Nat. Prod. Rep. 2009, 26, 338. doi:10.1039/B703499G
- [17] Z. Jin, Nat. Prod. Rep. 2006, 23, 464. doi:10.1039/B502166A
- [18] A. Bertram, G. Pattenden, Nat. Prod. Rep. 2007, 24, 18. doi:10.1039/ B612600F
- [19] J. P. Michael, G. Pattenden, Angew. Chem. Int. Ed. Engl. 1993, 32, 1.
 [Angew. Chem. 1993, 105, 1]. doi:10.1002/ANIE.199300013
- [20] Y. Shi, W. Jiang, B. N. Auckloo, B. Wu, Curr. Org. Chem. 2015, 19, 1935. doi:10.2174/1385272819666150709165210
- [21] A. L. van den Brenk, D. P. Fairlie, L. R. Gahan, G. R. Hanson, T. W. Hambley, *Inorg. Chem.* **1996**, *35*, 1095. doi:10.1021/IC9504755
- [22] P. Comba, N. Dovalil, L. R. Gahan, G. R. Hanson, M. Westphal, *Dalton Trans.* 2014, 43, 1935 and references therein. doi:10.1039/ C3DT52664J
- [23] P. Comba, A. Eisenschmidt, L. R. Gahan, D.-P. Herten, G. Nette, G. Schenk, M. Seefeld, *Chem. – Eur. J.* 2017, 23, 12264. doi:10.1002/ CHEM.201700895
- [24] P. Comba, A. Eisenschmidt, L. R. Gahan, G. R. Hanson, N. Mehrkens, M. Westphal, *Dalton Trans.* 2016, 45, 18931. doi:10.1039/ C6DT03787A
- [25] S. Xie, A. I. Savchenko, M. Kerscher, R. L. Grange, E. H. Krenske, J. R. Harmer, M. J. Bauer, N. Broit, D. J. Watters, G. M. Boyle, P. V. Bernhardt, P. G. Parsons, P. Comba, L. R. Gahan, C. M. Williams, *Eur. J. Org. Chem.* **2018**, 1465. doi:10.1002/EJOC.201701659
- [26] S. Xie, A. I. Savchenko, E. H. Krenske, R. L. Grange, L. R. Gahan, C. M. Williams, *Eur. J. Org. Chem.* 2018, 3265. doi:10.1002/EJOC. 201800449
- [27] B. M. Degnan, C. J. Hawkins, M. F. Lavin, E. J. McCaffrey, D. L. Parry, A. L. Van den Brenk, D. J. Watters, *J. Med. Chem.* **1989**, *32*, 1349. doi:10.1021/JM00126A034
- [28] C. M. Ireland, A. R. Durso, Jr, R. A. Newman, M. P. Hacker, J. Org. Chem. 1982, 47, 1807. doi:10.1021/JO00349A002
- [29] F. J. Schmitz, M. B. Ksebati, J. S. Chang, J. L. Wang, M. B. Hossain, D. Van der Helm, M. H. Engel, A. Serban, J. A. Silfer, *J. Org. Chem.* **1989**, *54*, 3463. doi:10.1021/JO00275A036
- [30] X. Fu, T. Do, F. J. Schmitz, V. Andrusevich, M. H. Engel, J. Nat. Prod. 1998, 61, 1547. doi:10.1021/NP9802872
- [31] M. A. Rashid, K. R. Gustafson, J. H. Cardellina II, M. R. Boyd, J. Nat. Prod. 1995, 58, 594. doi:10.1021/NP50118A020
- [32] Y. Hamamoto, M. Endo, M. Nakagawa, T. Nakanishi, K. Mizukawa, J. Chem. Soc. Chem. Commun. 1983, 323. doi:10.1039/C39830000323
- [33] T. Ishida, M. Inoue, Y. Hamada, S. Kato, T. Shioiri, J. Chem. Soc. Chem. Commun. 1987, 370. doi:10.1039/C39870000370

- [34] T. Ishida, M. Tanaka, M. Nabae, M. Inoue, S. Kato, Y. Hamada, T. Shioiri, J. Org. Chem. 1988, 53, 107. doi:10.1021/ JO00236A022
- [35] See, for example, A. Asano, K. Minoura, T. Yamada, M. Doi, J. Pept. Sci. 2016, 22, 156. doi:10.1002/PSC.2853
- [36] R. Zhao, S. Gove, J. E. Sundeen, B. C. Chen, *Tetrahedron Lett.* 2001, 42, 2101. doi:10.1016/S0040-4039(01)00161-7
- [37] Our group has also published a synthetic route to 2-amino-1,3thiazoles. See: L. A. Baker, C. M. Williams, J. Heterocycl. Chem. 2003, 40, 353. doi:10.1002/JHET.5570400225
- [38] M. S. Kerr, J. R. de Alaniz, T. Rovis, J. Org. Chem. 2005, 70, 5725. doi:10.1021/JO050645N
- [39] E. A. Merritt, M. C. Bagley, Synthesis 2007, 22, 3535.
- [40] Y. Takikawa, K. Shimada, K. Sato, S. Sato, S. Takizawa, Bull. Chem. Soc. Jpn. 1985, 58, 995. doi:10.1246/BCSJ.58.995
- [41] P. Comba, L. R. Gahan, G. R. Hanson, M. Westphal, *Chem. Commun.* 2012, 48, 9364. doi:10.1039/C2CC34836E
- [42] P. Comba, L. R. Gahan, G. R. Hanson, M. Maeder, M. Westphal, *Dalton Trans.* 2014, 43, 3144. doi:10.1039/C3DT53135J
- [43] A. L. van den Brenk, K. A. Byriel, D. P. Fairlie, L. R. Gahan, G. R. Hanson, C. J. Hawkins, A. Jones, C. H. L. Kennard, B. Moubaraki, K. S. Murray, *Inorg. Chem.* **1994**, *33*, 3549. doi:10.1021/ IC00094A019
- [44] (a) *MacroModel, version 10.6* 2014 (Schrödinger, LLC: New York, NY).
 (b) *Maestro, version 9.9* 2014 (Schrödinger, LLC: New York, NY).
- [45] (a) T. A. Halgren, J. Comput. Chem. 1996, 17, 490. doi:10. 1002/(SICI)1096-987X(199604)17:5/6<490::AID-JCC1>3.0.CO;2-P
 (b) T. A. Halgren, J. Comput. Chem. 1996, 17, 520. doi:10. 1002/(SICI)1096-987X(199604)17:5/6<520::AID-JCC2>3.0.CO;2-W
 (c) T. A. Halgren, J. Comput. Chem. 1996, 17, 553. doi:10. 1002/(SICI)1096-987X(199604)17:5/6<553::AID-JCC3>3.0.CO;2-T
 (d) T. A. Halgren, R. B. Nachbar, J. Comput. Chem. 1996, 17, 587.
 (e) T. A. Halgren, J. Comput. Chem. 1996, 17, 616. doi:10. 1002/(SICI)1096-987X(199604)17:5/6<616::AID-JCC3>3.0.CO;2-X
 (f) T. A. Halgren, J. Comput. Chem. 1999, 20, 720. doi:10. 1002/(SICI)1096-987X(199005)20:7<720::AID-JCC7>3.0.CO;2-X
 (g) T. A. Halgren, J. Comput. Chem. 1999, 20, 730. doi:10. 1002/(SICI)1096-987X(199905)20:7<730::AID-JCC8>3.0.CO;2-T

- [46] M. J. Frisch, G. W. Trucks, H. B. Schlegel, G. E. Scuseria, M. A. Robb, J. R. Cheeseman, G. Scalmani, V. Barone, G. A. Petersson, H. Nakatsuji, X. Li, M. Caricato, A. V. Marenich, J. Bloino, B. G. Janesko, R. Gomperts, B. Mennucci, H. P. Hratchian, J. V. Ortiz, A. F. Izmaylov, J. L. Sonnenberg, D. Williams-Young, F. Ding, F. Lipparini, F. Egidi, J. Goings, B. Peng, A. Petrone, T. Henderson, D. Ranasinghe, V. G. Zakrzewski, J. Gao, N. Rega, G. Zheng, W. Liang, M. Hada, M. Ehara, K. Toyota, R. Fukuda, J. Hasegawa, M. Ishida, T. Nakajima, Y. Honda, O. Kitao, H. Nakai, T. Vreven, K. Throssell, J. A. Montgomery, Jr, J. E. Peralta, F. Ogliaro, M. J. Bearpark, J. J. Heyd, E. N. Brothers, K. N. Kudin, V. N. Staroverov, T. A. Keith, R. Kobayashi, J. Normand, K. Raghavachari, A. P. Rendell, J. C. Burant, S. S. Iyengar, J. Tomasi, M. Cossi, J. M. Millam, M. Klene, C. Adamo, R. Cammi, J. W. Ochterski, R. L. Martin, K. Morokuma, O. Farkas, J. B. Foresman, D. J. Fox, Gaussian 16, Revision D.01 2016 (Gaussian, Inc.: Wallingford, CT).
- [47] (a) A. D. Becke, *J. Chem. Phys.* 1993, *98*, 5648. doi:10.1063/1.464913
 (b) C. Lee, W. Yang, R. G. Parr, *Phys. Rev. B Condens. Matter* 1988, *37*, 785. doi:10.1103/PHYSREVB.37.785
 (c) S. H. Vosko, L. Wilk, M. Nusair, *Can. J. Phys.* 1980, *58*, 1200. doi:10.1139/P80-159
 (d) P. J. Stephens, F. J. Devlin, C. F. Chabalowski, M. J. Frisch, *J. Phys. Chem.* 1994, *98*, 11623. doi:10.1021/J100096A001
- [48] Y. Zhao, D. G. Truhlar, J. Chem. Phys. 2006, 125, 194101. doi:10. 1063/1.2370993
- [49] (a) Y. Zhao, D. G. Truhlar, *Theor. Chem. Acc.* 2008, *120*, 215. doi:10. 1007/S00214-007-0310-X
 (b) A. V. Marenich, C. J. Cramer, D. G. Truhlar, *J. Phys. Chem. B* 2009, *113*, 6378. doi:10.1021/JP810292N
- [50] P. Comba, N. Dovalil, G. R. Hanson, G. Linti, *Inorg. Chem.* 2011, 50, 5165. doi:10.1021/IC2004694
- [51] L. J. Farrugia, J. Appl. Cryst. 1999, 32, 837. doi:10.1107/ S0021889899006020
- [52] G. M. Sheldrick, Acta Crystallogr. Sect. A 2008, 64, 112. doi:10.1107/ S0108767307043930
- [53] L. J. Farrugia, J. Appl. Cryst. 1997, 30, 565. doi:10.1107/ S0021889897003117