

WILDLIFÉ RESEARCH

# Power of faecal pellet count and camera trapping indices to monitor mammalian herbivore activity

Naomi E. Davis<sup>A,B,\*</sup><sup>(D)</sup>, Julian Di Stefano<sup>C</sup>, Jim Whelan<sup>D</sup>, John Wright<sup>B</sup>, Lorraine Taylor<sup>B</sup>, Graeme Coulson<sup>A</sup> and Holly Sitters<sup>C</sup>

For full list of author affiliations and declarations see end of paper

\*Correspondence to: Naomi E. Davis School of BioSciences, The University of Melbourne, Vic. 3010, Australia Email: Naomi.Davis@parks.vic.gov.au

Handling Editor: Tom Sullivan

Received: 8 September 2021 Accepted: 28 February 2022 Published: 7 June 2022

Cite this: Davis NE et al. (2022) Wildlife Research 49(8), 686–697. doi:10.1071/WR21135

© 2022 The Author(s) (or their employer(s)). Published by CSIRO Publishing. This is an open access article distributed under the Creative Commons Attribution-NonCommercial-NoDerivatives 4.0 International License (CC BY-NC-ND)

**OPEN ACCESS** 

#### ABSTRACT

Context. Monitoring spatial and temporal change in relative abundance using statistically powerful designs is a critical aspect of wildlife management. Many indices of relative abundance are available, but information regarding their influence on statistical power is limited. Aims. We compared the statistical power associated with occurrence-based and frequency-based indices derived from faecal pellet counts and camera trapping to detect changes in the activity of five mammalian herbivores. Methods. We deployed camera traps and counted faecal pellets in native vegetation subjected to four management treatments in south-eastern Australia. We used simulation coupled with generalised linear mixed models to investigate the statistical power associated with a range of effect sizes for each combination of species, survey method and data type. Key results. The index derived from camera frequency data provided the greatest statistical power to detect species' responses and was the only index capable of detecting small effect sizes with high power. The occurrence index from camera trapping did not provide the same level of statistical power. Indices derived from faecal pellet frequency data also detected spatial and temporal changes in activity levels for some species, but large numbers of plots were required to detect medium to large effect sizes. High power to detect medium to large effects could be achieved using occurrence indices derived from pellet presence-absence data, but required larger sample sizes compared to the camera frequency index. Conclusions. Both camera trapping and pellet counts can be applied to simultaneously monitor the activity of multiple mammalian herbivore species with differing activity patterns, behaviour, body size and densities, in open and closed habitat. However, using frequency indices derived from camera trapping may improve management outcomes by maximising the statistical power of monitoring programs to detect changes in abundance and habitat use. Implications. Frequency indices derived from camera trapping are expected to provide the most efficient method to detect changes in abundance. Where the use of cameras is cost prohibitive, occurrence indices derived from pellet presence-absence data can be used to detect medium to large effect sizes with high power. Nonetheless, the cost-effectiveness of camera trapping will improve as equipment costs are reduced and advances in automated image recognition and processing software are made.

**Keywords:** Axis porcinus, cost-effective, Macropus giganteus, mammal, management, monitoring, Oryctolagus cuniculus, sampling methods, survey methods, Vombatus ursinus, Wallabia bicolor.

# Introduction

Efficient and reliable methods to monitor spatial and temporal changes in wildlife populations are essential to inform and evaluate management decisions (Månsson *et al.* 2011) and increasingly, these methods need to be applied in multi-species assemblages (Pfeffer *et al.* 2018). Simple indices that reflect a change in abundance can be useful for wildlife management, but choices need to be made about which of the alternative indices to use (Engeman 2005). Selection of the most appropriate index can be informed by a formal evaluation of the efficacy and efficiency of candidate methods in the context of

management objectives and resources (Garden *et al.* 2007; Månsson *et al.* 2011; Swan *et al.* 2014; Seidlitz *et al.* 2021).

Indices based on non-invasive sampling methods enable monitoring of animals that are difficult to capture or detect, occur at low densities, or occur in dense vegetation where direct counts are not feasible (Southwell 1989; O'Connell et al. 2011). Indices are commonly based on counts of animal sign (Caughley 1977) and faecal pellets are the most obvious sign of the presence of many species. The faecal pellet count method uses pellet density as an index of herbivore abundance and activity (Bailey and Putman 1981). It can also provide an indicator of spatial or temporal habitat use, because pellet density is expected to be proportional to the time spent in a habitat (Hannan and Whelan 1989). Pellet count methods have been validated against known population densities for many species, revealing both linear and non-linear relationships (Barnes 2001; Hayward et al. 2005; Forsyth et al. 2007; McCann et al. 2008; Rönnegård et al. 2008; Ariefiandy et al. 2013). Faecal pellet counts can be applied simultaneously to multiple species with differing activity patterns and behaviour (e.g. Lunney and O'Connell 1988), and are suitable for use over large areas in most habitats (Mayle et al. 1999). Compared with many other monitoring methods, pellet counts are considered to be relatively accurate (Dinerstein 1980; Bailey and Putman 1981), time efficient and cost effective (Caughley 1977), and are frequently used to derive abundance indices for research and management (McCann et al. 2008; Forsyth et al. 2012).

An alternative sampling method is remote photography using camera traps (O'Connell et al. 2011). Camera traps contain heat and motion sensors that activate a camera when animal movement is detected (O'Connell et al. 2011). Camera trap data can index changes in population abundance (Rovero and Marshall 2009; Bengsen et al. 2011; Latham et al. 2012), activity patterns (Hossain et al. 2016) and habitat use (Augustine 2004; Claridge et al. 2004). Like pellet counts, camera traps can simultaneously monitor multiple species (Wacher and Attum 2005) in many habitat types, under varied environmental conditions (Vine et al. 2009). Camera traps are easy to use and, once deployed, do not require manual operation, allowing remote sampling over extended periods (Bridges and Noss 2011). Compared to a number of traditional methods, camera traps are relatively effective at detecting mammal species (Silveira et al. 2003; De Bondi et al. 2010; Paull et al. 2012; Swan et al. 2014; Wearn and Glover-Kapfer 2019), and are increasingly used for population monitoring (Karanth et al. 2004; Nichols et al. 2011). Importantly however, camera trapping does not always outperform traditional survey methods, providing lower or similar probabilities of detection and estimates of species richness and composition for some faunal groups (Silveira et al. 2003; Gompper et al. 2006; Espartosa et al. 2011; Seidlitz et al. 2021). Camera trapping also requires a substantial investment in equipment (Silveira et al. 2003)

and time for post-processing of data (Latham *et al.* 2012) compared to traditional methods such as pellet counts.

Studies comparing camera trapping and pellet counts for mammalian herbivores suggest that indices from the two methods can be correlated (Lucherini et al. 2009; Li et al. 2014; Morgan et al. 2018a) but camera trapping can index activity in situations where pellet counts are too low to be useful (Kuijper et al. 2009), and can more accurately estimate density (Roberts 2011). For carnivores, studies have demonstrated that camera and pellet-based indices of abundance were correlated for some species (Garrote et al. 2014) but not others (Gompper et al. 2006), likely influenced by variation in factors such as densities, ranging behaviour and the ability to identify individuals (Barea-Azcón et al. 2007; Jhala et al. 2011). While these studies provide general support for agreement between camera and pellet-based indices, they are limited to a small number of species and environments.

Assessment of the ability of monitoring methods to detect changes in animal populations also requires consideration of the type of data collected (Bengsen et al. 2014). Count data are common in ecological studies (Fordyce et al. 2011) and are used in wildlife monitoring to measure spatial and temporal variation in abundance (Archaux et al. 2012). Faecal pellet indices are usually based on counts (Mayle et al. 1999) and camera indices calculated as frequencies (e.g. counts of the number of observations standardised over time; Kawanishi et al. 1999; Negrões et al. 2010), but collection of count data is resource intensive (Gompper et al. 2006; Torney et al. 2019). An alternative is to collect species detection versus non-detection (i.e. presence-absence) data, which can be used to measure the probability of occurrence at a site. Presence-absence data are widely used for monitoring (Wintle et al. 2005; Steenweg et al. 2016) and may be easier to collect than count-based data enabling improvements in efficiency (Nichols et al. 2011). For example, in the context of remote camera surveys and faecal pellet counts, deriving presence-absence data sets removes the need to process all images or count all pellets, likely saving substantial time and cost. Despite this there is little information the relative performance of count about and presence-absence data for monitoring abundance change.

To ensure monitoring programs are rigorous and to optimise return on effort and expenditure, it is essential to evaluate the capacity of monitoring methods to detect change (Smart *et al.* 2004; Field *et al.* 2007). However, little information is available to compare the relative performance of indices from faecal pellet count and camera trapping methods, based on presence–absence and count data. We conducted a field-based comparison of faecal pellet counts (both faecal accumulation and faecal standing crop) and camera trapping. Our objective was to compare the statistical power of these methods, using indices derived from both occurrence (presence–absence) and frequency (count) data, to detect spatial and temporal changes in the relative activity levels of five mammalian herbivore species that differed in body size, behaviour, activity patterns and densities.

# **Materials and methods**

#### Study area

We conducted the study on Yanakie Isthmus (38°53'S; 146°14'E), Wilsons Promontory National Park, Victoria, Australia. Indices of herbivore activity inform decisions regarding management of grazing pressure at this site as part of an adaptive management approach to the restoration of Coastal Grassy Woodland invaded by a native woody shrub, coast teatree *Leptospermum laevigatum* (Morgan *et al.* 2018*b*).

The study area consisted of 200 ha of Coastal Grassy Woodland containing dunes and inter-dune swales with an open structure. *L. laevigatum* biomass in the shrub and canopy layers had been reduced using mechanical slashing (Davis *et al.* 2016) 4 years prior to the study, and/or prescribed burning in December 2012, 4 months prior to the study. The area also contained swales with a partially open structure, in which *L. laevigatum* was regenerating following prescribed burning 4 years prior to the study.

We focussed on five mammalian herbivore species (eastern grey kangaroo *Macropus giganteus*, swamp wallaby *Wallabia bicolor*, common wombat *Vombatus ursinus*, hog deer *Axis porcinus* and European rabbit *Oryctolagus cuniculus*) that require monitoring to guide management of grazing impacts (Morgan *et al.* 2018*b*). These species differed in behaviour, activity patterns and body size (ranging from 1 kg to 45 kg; Menkhorst and Knight 2011) and densities (Davis *et al.* 2018).

# **Experimental design**

We tested camera trapping and faecal pellet counts concurrently using the same experimental design and sampling locations to maximise comparability of data collected using the two survey methods. We counted pellets in circular 3-m radius plots (n = 201) to estimate faecal standing crop (FSC; Mayle et al. 1999) from 9 April to 2 June 2013, and simultaneously cleared pellets from plots in preparation for estimation of faecal accumulation rate (FAR; Mayle et al. 1999). We conducted the FAR survey from 15 to 18 July, approximately 3 months after the mid-point of the plot clearance period, with the accumulation period for plots ranging from 80 to 97 days. We deployed cameras at a subset of the 3-m radius plots (n = 36) for the duration of the FSC and FAR survey periods (9 April-18 July). We placed cameras on the northern boundary of plots so that the detection zone encompassed most of the plot but moved cameras 5 m from the northern boundary if vegetation clipping in the detection zone was required, to ensure that clipping did not interfere with the test of pellet counts in dense regrowth.

We used both pellet counts and camera trapping to sample 12 swales (0.18–4.18 ha;  $\overline{x} = 1.74$  ha) separated by ~0.05-2 km because managers aimed to restore open vegetation and were interested in monitoring herbivore responses to management interventions at this scale. To test monitoring methods in vegetation with varied structure, which we expected would result in spatial differences in herbivore activity, we sampled four treatments (three swales per treatment): (1) slashed unburnt - swales opened by mechanical slashing alone; (2) slashed burnt 2012 - swales opened by mechanical slashing and burning; (3) unslashed burnt 2012 swales opened by burning alone in 2012; and (4) unslashed burnt 2009 - swales opened by burning alone in 2009, but which had partially closed following vegetation regeneration (Fig. 1). All swales were flat to slightly undulating. Slashed unburnt swales were predominantly open in structure, with high (60-70%) ground layer vegetation cover and small patches of dead fallen L. laevigatum branches (20-30% cover). Slashed burnt 2012 swales were similarly open in structure with less (30-50%) ground layer vegetation cover, and the remainder of the ground surface was bare. Unslashed burnt 2012 swales were largely open but dominated by dense burnt L. laevigatum stems in places, the ground was predominantly bare, and there was little (5-30% cover) ground layer vegetation. Unslashed burnt 2009 swales had dense ground (80-90%) and shrub (0.6-2 m; 40-90%) cover dominated by L. laevigatum. Vegetation cover values were estimated visually within each treatment.

To select sampling locations within each swale we applied a 10-m buffer around swale boundaries to reduce edge effects, then randomly selected locations from 20-m grid intercept points generated by geographic information system (GIS) mapping. We allocated plots to swales in approximate proportion to area: slashed unburnt (n = 50); slashed burnt 2012 (n = 59); unslashed burnt 2012 (n = 60); and unslashed burnt 2009 (n = 32). For camera traps, we divided samples equally among swales (n = 9 per treatment) to ensure that each swale was sampled in the event of camera malfunction.

# **Camera trapping**

We used Reconyx HC600 Hyperfire H.O. covert infrared cameras, which have passive infrared (PIR) capability to detect the difference between the ambient air temperature and the animal's body temperature. We used a passive survey approach (i.e. no lure), so that herbivore behaviour was not altered (Meek *et al.* 2012).

We standardised camera placement and settings to minimise differential detection probabilities (O'Brien 2011) and maximise detection of target species. We mounted cameras on wooden stakes at a height of 50 cm, set horizontally with the PIR sensor aimed 6 m away at the smallest species core body zone (i.e.  $\sim$ 20 cm above the ground) (Meek *et al.* 2012). Cameras were set to high sensitivity and three images per trigger with no delay, facing south to improve



Fig. 1. Photographs demonstrating treatment variation within the study site, Yanakie Isthmus, Wilsons Promontory National Park, Victoria, Australia, 2013: (a) slashed unburnt swales; (b) slashed burnt 2012 swales; (c) unslashed burnt 2012 swales; (d) unslashed burnt 2009 swales.

image quality and prevent triggering by direct sunlight (Meek *et al.* 2012).

To allow comparisons across open and closed habitats we standardised camera detection zone size by placing cameras on flat ground with a post 6 m in front of cameras to mark the limit of the detection zone. In addition, we clipped vegetation to 15 cm above the ground using a brush-cutter within the conical shaped detection zone extending 6 m in front of the camera and  $40^{\circ}$  either side of that central line of sight.

For each day during the survey period, we recorded whether each camera detected the target species within the 6-m detection zone. For marsupials, we counted young-at-foot, but not pouch young, based on the assumption that only young-at-foot contribute directly to grazing pressure and because detectability of pouch-young would be highly variable.

#### Faecal pellet counts

We used faecal accumulation rate (FAR) counts to reduce the lag between changes in animal activity and the ability of faecal pellet counts to detect that change (Davis and Coulson 2016), and reduce bias associated with pellet decay (Campbell *et al.* 2004). In addition, we used faecal standing crop (FSC) counts, which can provide a precise index of changes in herbivore activity (Mayle *et al.* 1999).

We counted individual pellets for the three marsupial species (eastern grey kangaroo, swamp wallaby, common wombat), given their small pellet group sizes and difficulties that can arise in distinguishing groups (Southwell 1989). We counted pellet groups for hog deer, because the number of pellet groups produced daily is a more reliable index of deer density than the number of individual pellets produced (Smith 1964), and counting is more efficient for species that deposit large numbers of small pellets. Following Hickling (1986), we defined hog deer pellet groups as  $\geq 6$  pellets of the same defecation, with at least one visible above the ground litter. Rabbit pellets are also commonly deposited in groups, so we also applied this definition to rabbit pellet groups. If rabbit pellet groups could not be defined within latrines containing large deposits of pellets,

we approximated groups as consisting of *c*. 20 pellets (Davis and Coulson 2016). We counted pellet groups if >50% of pellets fell inside the plot (Mayle *et al.* 1999).

We identified pellets based on size, shape and colour (Triggs 2003). We systematically searched plots for pellets by pushing the vegetation aside, but not disturbing the litter except to look for additional deer or rabbit pellets when one was visible above the litter (Hickling 1986).

#### Statistical analyses

We used faecal pellet and camera trapping data to index spatial and temporal changes in species activity. Two activity indices were derived from each method, using occurrence (presence-absence) and frequency (count) data (Table 1). Because statistical power to detect change is inversely related to the variance of the response variable, we undertook preliminary analyses to explore the variance associated with different forms of the camera trapping frequency index, as numerous options exist (Burton et al. 2015). We selected a camera frequency index based on counts of the number of days on which each species was recorded as present as a proportion of the maximum number of survey days because it resulted in lower variance associated with outliers (large numbers of images at some plots) and overdispersion than did other options examined. Similar approaches using indices based on the number of days of detection have been applied by others (e.g. Geary et al. 2018; Nalliah et al. 2022).

We compared indices using data collected during the same sampling periods because herbivore activity patterns may change seasonally. FSC counts were conducted at the end of autumn to predominantly reflect autumn activity levels (Davis and Coulson 2016), whereas FAR counts reflected winter activity levels. We then subsampled camera data from two 14-day survey periods and compared camera and FSC data collected in autumn between 9 April and 2 June, and camera and FAR data collected in winter between 30 June and 18 July.

We used generalised linear mixed models (GLMMs) and simulated datasets to investigate the statistical power associated with a range of effect sizes for each combination of species, survey method and data type. Initially we built GLMMs for each species with treatment included as a fixed effect and swale specified as a random effect to accommodate variance associated with the spatial nestedness of the study design resulting from sampling of plots within swales (Zuur et al. 2009). We also included an individual-level random effect to correct for overdispersion where necessary (Zuur et al. 2009). Models of species' occurrence and frequency indices (Table 1) were applied with a logit link function and binomial errors, allowing us to standardise effect-size tests across all data types. The rabbit FSC frequency index model failed to converge so we excluded it from the analysis.

Power analyses were conducted in R version 3.6.3 (R Core Team 2020) using the package simr, which calculates power for GLMMs using Monte Carlo simulation (Green and MacLeod 2016). Calculations involved three steps: (1) new response-variable values were simulated 1000 times based on the GLMM; (2) the model was fitted to the simulated response data; (3) a likelihood ratio test was applied to the simulated fit. The power estimate was derived from the number of successes and failures to detect a statistically significant effect ( $P \le 0.05$ ) in step 3.

For each combination of species, survey method and data type, we adjusted the effect size of interest using the fixef function in the simr package (Green and MacLeod 2016). Retrospective observed power calculations, where the target effect size is determined by the data, can generate misleading results (Hoenig and Heisey 2001), so we considered a range of ecologically plausible effect sizes representing small (0.10), medium (0.30) and large (0.50) differences in the probability of occurrence (occurrence indices) or proportions (frequency indices) (Table 2). Greater statistical power is required to detect a given effect size at the extreme ends of the binomial distribution (for example, a difference in the

Table I.	Activity indices derived from faecal accumulation rate (FAR) counts, faecal standing crop (FSC) coun	its and camera trapping data to
index spati	al and temporal changes in species activity.	

Survey method	Data type	Description
FSC	Occurrence	Probability of occurrence based on the presence-absence of pellets (marsupials) or pellet groups (deer and rabbit) during the FSC survey period
	Frequency	Counts of the number of pellets or pellet groups during the FSC survey period as a proportion of the maximum count per species
FAR	Occurrence	Probability of occurrence based on the presence-absence of pellets (marsupials) or pellet groups (deer and rabbit) at the end of the accumulation period
	Frequency	Counts of the number of pellets or pellet groups at the end of the accumulation period as a proportion of the maximum count per species
Camera trapping	Occurrence	Probability of occurrence based on the presence-absence of each species over a 14-day period
	Frequency	Camera frequency index (counts of the number of days on which each species was recorded as present as a proportion of the maximum number of 14 days)

	Categories/scenarios
Data type	Stage I: Cameras, faecal standing crop
	Stage 2: Cameras, faecal accumulation rate
Species	Eastern grey kangaroo, swamp wallaby, common wombat, hog deer, rabbit
Response variables	Presence-absence, frequency (for cameras data, this was the number of days a species was detected out of the total number of days the camera was deployed; for FSC and FAR data, this was the number of pellets counted in a plot relative to the maximum number of pellets recorded in plot)
Effect size	Small (0.10): centre (0.55–0.45), upper end (0.90–0.80)
	Medium (0.30): centre (0.65–0.35), upper end (0.90–0.60)
	Large (0.50): centre (0.75–0.25), upper end (0.90–0.40)
Number of plots per treatment (fixed effect)	5, 10, 20, 40, 60, 80, 100, 120, 140, 160, 180, 200, 220, 240, 260, 280, 300
Total number of swales (random effect)	5, 10, 20, 40, 60, 80, 100

Table 2. Scenarios (species, method, sampling effort, data type and effect size) used for power analysis.

probability of occurrence between 0.2 and 0.1, or between 0.9 and 0.8) relative to the centre (a difference between 0.55 and 0.45), so we examined each of three effect sizes (small, medium and large) at both the centre and upper end of the range. To investigate the trade-off between sample size and power, we used the powerCurve function in simr to estimate power associated with a sample size of 0–300 plots.

### Results

Species' responses to time and treatment were generally similar for the alternative activity indices (Supplementary Figs S1, S2). However, the survey effort required to detect significant effects of treatments during both autumn and winter varied between indices from different survey methods, and between indices derived from frequency and occurrence data (camera trapping compared to FSC; autumn: Fig. 2) and (camera trapping compared to FAR; winter: Fig. 3).

#### Indices based on frequency

For all species, the activity index derived from camera frequency data was able to detect treatment effects with greater power using fewer plots than the other activity indices (Figs 2, 3). For example, in considering power resulting from up to 300 plots, camera frequency data delivered the only index capable of detecting small effect sizes with high power, and fewer than 50 plots were sufficient to detect medium to large effect sizes for most species using this index.

In contrast to the activity index derived from camera frequency data, power to detect small effects using indices derived from pellet count frequency data was low (Figs 2, 3). Power associated with the FSC frequency index was consistently low, with greater than 100 plots required to detect all but large effect sizes for most species (Fig. 2). For the FAR

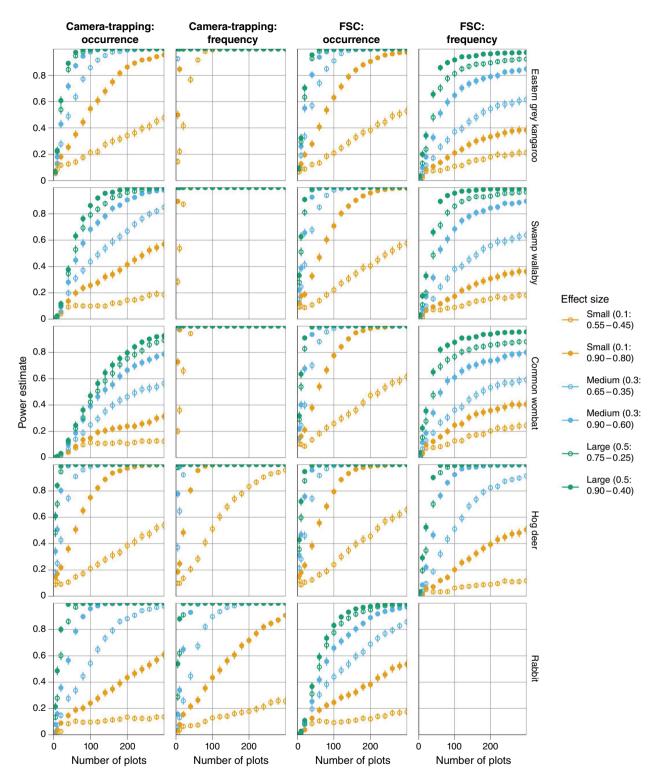
frequency index, power to detect medium to large effects varied among species (Fig. 3). FAR frequency data had high power to detect large effects in swamp wallaby and wombat activity levels (Fig. 3). However, the number of plots required to detect medium to large effect sizes for most species was similarly large (>100) to that required for the FSC frequency index, and for species such as deer, even very large sample sizes (>300 plots) were not adequate to detect large effect sizes with high power (Figs 2, 3).

#### Indices based on occurrence

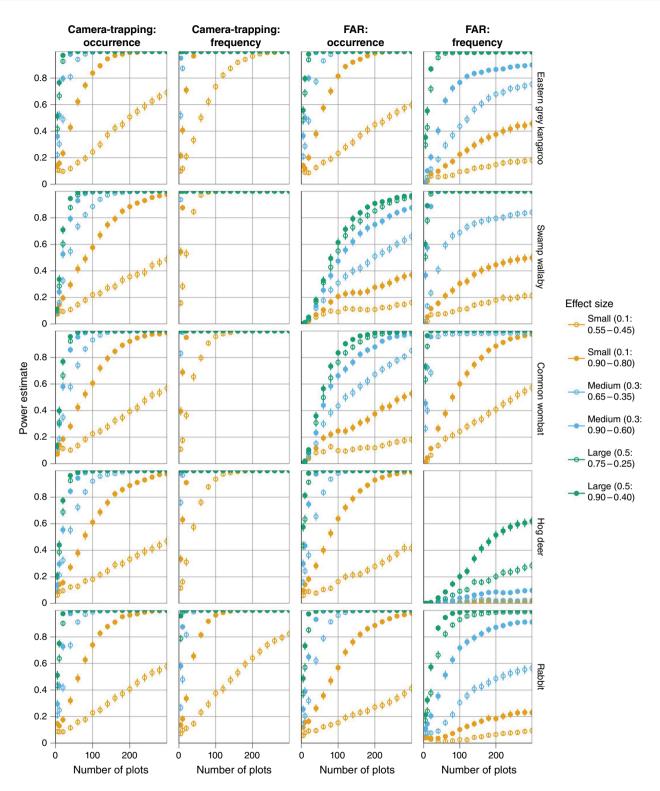
Occurrence indices from the three alternative methods performed similarly, requiring fewer plots than pellet count frequency indices, but a greater number of plots than the camera frequency index to detect small-large effects for most species (Figs 2, 3). Overall, power to detect small effects was low for indices derived from occurrence data. However, there was interspecific variation in the number of plots required to detect effects using occurrence indices, and the performance of the camera occurrence index varied over time. Medium-large effect sizes could be detected with high power for several species using occurrence indices: FSC occurrence index for all species except rabbit; and FAR and camera occurrence indices for kangaroo, deer and rabbit. Further, small effects sizes could be detected with medium to high power for some species using pellet occurrence indices if a large number of plots was used.

# Discussion

Increasingly, it is recognised that the objectives of monitoring programs can be met using indices of relative abundance (Engeman 2005; Johnson 2008), yet there is limited information on the performance of different indices to help



**Fig. 2.** Statistical power for three effect sizes (small, medium and large) at both the centre and upper end of the binomial distribution. Power was calculated using Monte Carlo simulation for combinations of species (eastern grey kangaroo, swamp wallaby, common wombat, hog deer, rabbit), data type (Faecal Standing Crop and camera activity indices derived from occupancy and frequency data collected at Wilsons Promontory National Park during autumn; 9 April–2 June 2013) and response variable modelled using generalised linear mixed models (GLMM). No output was obtained for the rabbit activity index derived from Faecal Standing Crop frequency data due to computational problems.



**Fig. 3.** Statistical power for three effect sizes (small, medium and large) at both the centre and upper end of the binomial distribution. Power was calculated using Monte Carlo simulation for combinations of species (eastern grey kangaroo, swamp wallaby, common wombat, hog deer, rabbit), data type (Faecal Accumulation Rate and camera activity indices derived from occupancy and frequency data collected at Wilsons Promontory National Park during winter; 30 June–18 July 2013) and response variable modelled using generalised linear mixed models (GLMM).

managers choose between the range of survey methods and metrics available. Our study compared the statistical power of faecal pellet count and camera trapping methods for detecting changes in the relative activity levels of mammalian herbivores using indices based on frequency and occurrence data. We demonstrated that an activity index derived from camera frequency data detected treatment effects with greater power and fewer plots compared to the other activity indices and was the only index capable of detecting small effect sizes with high power.

Our results support the work of Latham *et al.* (2012) who showed that medium to large changes in rabbit numbers could be detected using count data from small camera sample sizes and short deployment times with good statistical power. In line with others (Silveira *et al.* 2003; Balme *et al.* 2009; Roberts 2011) we, therefore, advocate camera trapping as providing high power and rigour for monitoring change in relative abundance in a range of species. Some studies have noted that a combination of methods may be required to ensure that different species are detected (Catling *et al.* 1997; Swan *et al.* 2014), yet our camera setup was effective for detecting species with different body sizes and behaviours, maximising the monitoring potential of this method (Engeman 2005).

While the camera-based frequency index gave the highest statistical power, it is important to evaluate monitoring methods relative to their cost (Perkins et al. 2013). We did not undertake a cost-benefit analysis because costs vary widely between projects due to factors such as travel times and whether paid staff or volunteers conduct field work, and because costs change over time and among regions. Nonetheless, it is known that the initial outlay required for camera trapping equipment is greater than that required for low-tech sign survey methods (Gompper et al. 2006; Lucherini et al. 2009; Garrote et al. 2014; Paton et al. 2021). This can make camera trap monitoring cost prohibitive if equipment costs cannot be spread across multiple projects, even with decreases in the cost of camera equipment and the time involved in post-processing of images (Latham et al. 2012) as camera trapping technology such as automated image recognition and processing software advance (Norouzzadeh et al. 2018; Weinstein 2018). Therefore, when cameras cannot be purchased or borrowed, but labour resources are readily available and detection of medium to large effect sizes is adequate to meet monitoring objectives, indices derived from FAR or FSC occurrence data (depending on the species) are more appropriate choices. Moreover, while detection of small effect sizes may be essential for some programs (e.g. threatened species monitoring) (Smart et al. 2004), the small effect sizes detected using camera trapping frequency indices may not be ecologically meaningful or relevant to management of many wildlife populations (Di Stefano 2003). Other indices may, therefore, provide adequate statistical power to detect species' responses with equivalent effort and lower costs.

r and any could be applied to monitor herbivore activity in open or closed habitat. Occurrence data are appealing because they are relatively simple and cost-effective to collect (De Bondi *et al.* 2010; Steenweg *et al.* 2016) compared to frequency data (Latham *et al.* 2012). Overall, we found that the performance of all occurrence indices was similar, and while their power to detect small effect sizes was low, medium-large effect sizes were detected with high power for several species.
For some species, pellet occurrence indices resulted in

greater power than the camera occurrence index, and an equivalent investment in equipment and labour may provide a similar level of power to that achieved using a camera frequency index. Further, our results showed that a large number of plots are generally required to detect medium to large effect sizes using pellet frequency indices, challenging the common preference for collecting pellet frequency data. If simple occurrence metrics can detect meaningful effect sizes, they are likely to be more cost-effective than timeconsuming counts (Latham et al. 2012). FSC counts are particularly cost-effective, as they do not require plot clearance, although they did not perform well for some species (e.g. a large number of plots were required to detect medium-large effect sizes for rabbit). Occurrence data from scat detections may be particularly useful for rare and elusive species (Seidlitz et al. 2021).

All six indices considered told a similar ecological story,

Some variation in the spatial and temporal trends detected by the different methods was evident, in line with work by Paton *et al.* (2021), suggesting that they measure slightly different aspects of activity and are subject to different biases. Nonetheless, the influence of such factors appears to be smaller than the magnitude of real change and we show that techniques that sample in different ways, at different scales, can provide comparable indices of activity, as has been found during comparisons of other techniques (e.g. camera trapping and track counts; Espartosa *et al.* 2011).

The applicability of our results to other systems and monitoring programs will depend on objectives and resourcing (Garden et al. 2007) and factors that influence the suitability, efficiency and power of monitoring using pellet counts and camera trapping, including species characteristics and population densities, survey timing, habitat type and climate (Towerton et al. 2011; Perry and Robertson 2012; Perkins et al. 2013). The experimental design, logistics and procedures involved in pellet counts and camera trapping were simple, so either could be implemented without sophisticated training, however, the ability of observers to accurately identify pellets versus images will depend on the target species. If supplementary data, such as animal behaviour observations are required, camera trapping is recommended (Silveira et al. 2003; Paull et al. 2012), but in areas of high human visitation, pellet counts may be necessary due to the risk of camera theft (Espartosa et al. 2011). Occurrence indices may not be suitable where a

species is abundant and ubiquitous due to 'oversaturation' (i.e. not enough absences) (Bengsen et al. 2014), in situations where abundance or activity are reduced without causing local extinctions (Bengsen et al. 2014), or when a species is rare (Fithian et al. 2015). On the other hand, precision of frequency indices from the FAR method is improved when levels of pellet accumulation are high (Campbell et al. 2004; Smart et al. 2004) and counts from cameras may successfully index visitation at sites where pellet counts are too low to be useful (Kuijper et al. 2009). Further, methods can be adjusted according to local conditions to improve the utility of the data. For example, when species are uncommon, camera deployment times (Latham et al. 2012) or FAR accumulation times can be increased (Campbell et al. 2004; Smart et al. 2004) to improve precision and statistical power, without additional costs.

We demonstrated that an activity index derived from camera frequency data was the most powerful wildlife monitoring method for detecting species' responses to management, providing the only index capable of detecting small effect sizes with high power. Our study was restricted to five species at a single location, yet their activity patterns, behaviour, body size and densities varied, and habitat structure varied across the four management treatments examined. Therefore, we believe our results have broader applicability, and advocate camera trapping as a powerful method that can be applied to simultaneously monitor multiple species to improve management outcomes. However, in monitoring programs where funding for cameras is inadequate, but labour resources are readily available, and detection of medium to large effect sizes is adequate to meet objectives, indices derived from pellet count occurrence data should be considered. Occurrence indices derived from FSC and FAR outperformed pellet-based frequency indices in most instances, showing that simple metrics have the potential to detect ecologically important change and improve the cost-effectiveness of monitoring. Nonetheless, with the rapid evolution of camera trapping technology, the efficiency of camera-based methods is likely to increase further as equipment costs are reduced and advances in automated image recognition and processing software are made.

# Supplementary material

Supplementary material is available online.

#### References

- Archaux F, Henry P-Y, Gimenez O (2012) When can we ignore the problem of imperfect detection in comparative studies? *Methods in Ecology and Evolution* **3**, 188–194. doi:10.1111/j.2041-210X.2011. 00142.x
- Ariefiandy A, Purwandana D, Coulson G, Forsyth DM, Jessop TS (2013) Monitoring the ungulate prey of the Komodo dragon Varanus komodoensis: distance sampling or faecal counts? Wildlife Biology 19, 126–137. doi:10.2981/11-098

- Augustine DJ (2004) Influence of cattle management on habitat selection by impala on central Kenyan rangeland. *Journal of Wildlife Management* **68**, 916–923. doi:10.2193/0022-541X(2004) 068[0916:IOCMOH]2.0.CO;2
- Bailey RE, Putman RJ (1981) Estimation of fallow deer (*Dama dama*) populations from faecal accumulation. *Journal of Applied Ecology* **18**, 697–702. doi:10.2307/2402361
- Balme GA, Hunter LTB, Slotow R (2009) Evaluating methods for counting cryptic carnivores. *Journal of Wildlife Management* 73, 433–441. doi:10.2193/2007-368
- Barea-Azcón JM, Virgós E, Ballesteros-Duperón E, Moleón M, Chirosa M (2007) Surveying carnivores at large spatial scales: a comparison of four broad-applied methods. *Biodiversity and Conservation* 16, 1213–1230. doi:10.1007/s10531-006-9114-x
- Barnes RFW (2001) How reliable are dung counts for estimating elephant numbers? *African Journal of Ecology* **39**, 1–9. doi:10.1111/j. 1365-2028.2001.00266.x
- Bengsen AJ, Leung LK-P, Lapidge SJ, Gordon IJ (2011) Using a general index approach to analyse camera-trap abundance indices. *Journal of Wildlife Management* 75, 1222–1227. doi:10.1002/jwmg.132
- Bengsen A, Robinson R, Chaffey C, Gavenlock J, Hornsby V, Hurst R, Fosdick M (2014) Camera trap surveys to evaluate pest animal control operations. *Ecological Management and Restoration* 15, 97–100. doi:10.1111/emr.12086
- Bridges AS, Noss AS (2011) Behaviour and activity patterns. In. 'Camera traps in animal ecology: Methods and analyses'. (Eds AF O'Connell, JD Nichols, KU Karanth) pp. 57–69. (Springer: London)
- Burton AC, Neilson E, Moreira D, Ladle A, Steenweg R, Fisher JT, Bayne E, Boutin S (2015) REVIEW: Wildlife camera trapping: a review and recommendations for linking surveys to ecological processes. *Journal of Applied Ecology* **52**, 675–685. doi:10.1111/1365-2664.12432
- Campbell D, Swanson GM, Sales J (2004) Comparing the precision and cost-effectiveness of faecal pellet group count methods. *Journal of Applied Ecology* **41**, 1185–1196. doi:10.1111/j.0021-8901.2004. 00964.x
- Catling PC, Burt RJ, Kooyman R (1997) A comparison of techniques used in a survey of the ground-dwelling and arboreal mammals in forests in north-eastern New South Wales. *Wildlife Research* 24, 417–432. doi:10.1071/WR96073
- Caughley G (1977) 'Analysis of vertebrate populations.' (John Wiley: New York, NY)
- Claridge AW, Mifsud G, Dawson J, Saxon MJ (2004) Use of infrared digital cameras to investigate the behaviour of cryptic species. *Wildlife Research* 31, 645–650. doi:10.1071/WR03072
- Davis NE, Coulson G (2016) Habitat-specific faecal pellet decay rates for five mammalian herbivores in south-eastern Australia. *Australian Mammalogy* 38, 105–116. doi:10.1071/AM15007
- Davis NE, Di Stefano J, Coulson G, Whelan J, Wright J (2016) Vegetation management influences habitat use by mammalian herbivores in shrub-encroached grassy woodland. Wildlife Research 43, 438–447. doi:10.1071/WR15218
- Davis NE, Gordon IR, Coulson G (2018) The influence of evolutionary history and body size on partitioning of habitat resources by mammalian herbivores in south-eastern Australia. *Australian Journal of Zoology* **65**, 226–239. doi:10.1071/ZO16075
- De Bondi N, White JG, Stevens M, Cooke R (2010) A comparison of the effectiveness of camera trapping and live trapping for sampling terrestrial small-mammal communities. *Wildlife Research* **37**, 456–465. doi:10.1071/wr10046
- Di Stefano J (2003) How much power is enough? Against the development of an arbitrary convention for statistical power calculations. *Functional Ecology* **17**, 707–709. doi:10.1046/j.1365-2435.2003. 00782.x
- Dinerstein E (1980) An ecological survey of the Royal Karnali-bardia Wildlife Reserve, Nepal. Part III: Ungulate populations. *Biological Conservation* **18**, 5–37. doi:10.1016/0006-3207(80)90063-4
- Engeman RM (2005) Indexing principles and a widely applicable paradigm for indexing animal populations. Wildlife Research 32, 203–210. doi:10.1071/WR03120
- Espartosa KD, Pinotti BT, Pardini R (2011) Performance of camera trapping and track counts for surveying large mammals in rainforest remnants. *Biodiversity and Conservation* **20**, 2815–2829. doi:10.1007/s10531-011-0110-4

- Field SA, O'Connor PJ, Tyre AJ, Possingham HP (2007) Making monitoring meaningful. *Austral Ecology* **32**, 485–491. doi:10.1111/j. 1442-9993.2007.01715.x
- Fithian W, Elith J, Hastie T, Keith DA (2015) Bias correction in species distribution models: pooling survey and collection data for multiple species. *Methods in Ecology and Evolution* 6, 424–438. doi:10.1111/ 2041-210X.12242
- Fordyce JA, Gompert Z, Forister ML, Nice CC (2011) A Hierarchical Bayesian Approach to Ecological Count Data: A Flexible Tool for Ecologists. *PLoS One* **6**, e26785. doi:10.1371/journal.pone.0026785
- Forsyth DM, Barker RJ, Morriss G, Scroggie MP (2007) Modeling the relationship between fecal pellet indices and deer density. *Journal of Wildlife Management* **71**, 964–970. doi:10.2193/2005-695
- Forsyth DM, Gormley AM, Woodford L, Fitzgerald T (2012) Effects of large-scale high-severity fire on occupancy and abundances of an invasive large mammal in south-eastern Australia. Wildlife Research 39, 555–564. doi:10.1071/WR12033
- Garden JG, McAlpine CA, Possingham HP, Jones DN (2007) Using multiple survey methods to detect terrestrial reptiles and mammals: What are the most successful and cost-efficient combinations? *Wildlife Research* 34, 218–227. doi:10.1071/wr06111
- Garrote G, de Ayala RP, Tellería JL (2014) A comparison of scat counts and camera-trapping as means of assessing Iberian lynx abundance. *European Journal of Wildlife Research* **60**, 885–889. doi:10.1007/ s10344-014-0855-7
- Geary WL, Ritchie EG, Lawton JA, Healey TR, Nimmo DG (2018) Incorporating disturbance into trophic ecology: Fire history shapes mesopredator suppression by an apex predator. *Journal of Applied Ecology* 55, 1594–1603. doi:10.1111/1365-2664.13125
- Gompper ME, Kays RW, Justina CR, Lapoint SD, Bogan DA, Cryan JR (2006) A Comparison of Noninvasive Techniques to Survey Carnivore Communities in Northeastern North America. Wildlife Society Bulletin 34, 1142–1151. doi:10.2193/0091-7648(2006)34[1142:ACONTT]2. 0.CO;2
- Green P, MacLeod CJ (2016) simr: an R package for power analysis of generalised linear mixed models by simulation. *Methods in Ecology and Evolution* **7**, 493–498. doi:10.1111/2041-210X.12504
- Hannan MJ, Whelan J (1989) Deer and habitat relations in managed forests. In 'Mammals as pests'. (Ed. RJ Putman) pp. 116–127. (Chapman and Hall: New York, NY)
- Hayward MW, de Tores PJ, Dillon MJ, Fox BJ, Banks PB (2005) Using faecal pellet counts along transects to estimate quokka (*Setonix brachyurus*) population density. *Wildlife Research* **32**, 503–507. doi:10.1071/WR03046
- Hickling GJ (1986) Red deer population surveys in the Harper-Avoca catchment (1956-1983). *Forest Research Institute Bulletin* **107**, 1–10.
- Hoenig JM, Heisey DM (2001) The abuse of power: The pervasive fallacy of power calculations for data analysis. *The American Statistician* **55**, 19–24. doi:10.1198/000313001300339897
- Hossain ANM, Barlow A, Greenwood Barlow C, Lynam AJ, Chakma S, Savini T (2016) Assessing the efficacy of camera trapping as a tool for increasing detection rates of wildlife crime in tropical protected areas. *Biological Conservation* **201**, 314–319. doi:10.1016/j.biocon. 2016.07.023
- Jhala Y, Qureshi Q, Gopal R (2011) Can the abundance of tigers be assessed from their signs? *Journal of Applied Ecology* **48**, 14–24. doi:10.1111/j.1365-2664.2010.01901.x
- Johnson DH (2008) In defense of indices: The case of bird surveys. The Journal of Wildlife Management 72, 857–868. doi:10.2193/2007-294
- Karanth KU, Nichols JD, Kumar NS, Link WA, Hines JE (2004) Tigers and their prey: predicting carnivore densities from prey abundance. *Proceedings of the National Academy of Sciences* **101**, 4854–4858. doi:10.1073/pnas.0306210101
- Kawanishi K, Sahak AM, Sunquist M (1999) Preliminary analysis on abundance of large mammals at Sungai Relau, Taman Negara. *Journal of Wildlife and Parks (Malaysia)* **17**, 62–82.
- Kuijper DPJ, Cromsigt JPGM, Churski M, Adam B, Jędrzejewska B, Jędrzejewski W (2009) Do ungulates preferentially feed in forest gaps in European temperate forest? *Forest Ecology and Management* 258, 1528–1535. doi:10.1016/j.foreco.2009.07.010
- Latham ADM, Nugent G, Warburton B (2012) Evaluation of camera traps monitoring European rabbits before and after control

operations in Otago, New Zealand. Wildlife Research 39, 621-628. doi:10.1071/WR12050

- Li X, Buzzard P, Jiang X (2014) Habitat associations of four ungulates in mountain forests of southwest China, based on camera trapping and dung counts data. *Population Ecology* **56**, 251–256. doi:10.1007/s10144-013-0405-2
- Lucherini M, Reppucci JI, Luengos Vidal E (2009) A comparison of three methos to estimate variations in the relative abundance of mountain vizcachas (*Lagidium viscacia*) in the high Andes ecosystems. *Mastozoología Neotropical* **16**, 223–228.
- Lunney D, O'Connell M (1988) Habitat selection by the swamp wallaby Wallabia bicolor the red-necked wallaby Macropus rufogriseus and the common wombat Vombatus ursinus in logged burnt forest near Bega New South Wales Australia. Australian Wildlife Research 15, 695–706. doi:10.1071/WR9880695
- Månsson J, Hauser CE, Andrén H, Possingham HP (2011) Survey method choice for wildlife management: the case of moose Alces Alces in Sweden. Wildlife Biology 17, 176–190. doi:10.2981/10-052
- Mayle BA, Peace AJ, Gill RMA (1999) 'How Many Deer? A Field Guide to Estimating Deer Population Size.' (Forestry Commission: Edinburgh)
- McCann NP, Moen RA, Niemi GJ (2008) Using Pellet Counts to Estimate Snowshoe Hare Numbers in Minnesota. Journal of Wildlife Management 72, 955–958. doi:10.2193/2007-083
- Meek PD, Ballard G, Fleming P (2012) 'An Introduction to Camera Trapping for Wildlife Surveys in Australia.' (PestSmart Toolkit Publication, Invasive Animals Cooperative Research Centre: Canberra, Australia)
- Menkhorst P, Knight F (2011) 'A field guide to the mammals of Australia', 3rd edn. (Oxford University Press: South Melbourne, Victoria)
- Morgan HR, Ballard G, Fleming PJS, Reid N, Van der Ven R, Vernes K (2018*a*) Estimating macropod grazing density and defining activity patterns using camera-trap image analysis. *Wildlife Research* **45**, 706–717. doi:10.1071/WR17162
- Morgan J, Wright J, Whelan J, Clarke M, Coulson G, Lunt I, Stoner J, Varcoe T, Shannon J (2018b) What does it take to do successful adaptive management? A case study highlighting Coastal Grassy Woodland restoration at Yanakie Isthmus. *Ecological Management and Restoration* **19**, 111–123. doi:10.1111/emr.12309
- Nalliah R, Sitters H, Smith A, Di Stefano J (2022) Untangling the influences of fire, habitat and introduced predators on the endangered heath mouse. *Animal Conservation* **25**(2), 208–220. doi:10.1111/acv.12731
- Negrões N, Sarmento P, Cruz J, Eira C, Revilla E, Fonseca C, Soliman R, Tôrres NM, Furtado MM, Jácomo ATA, Silveira L (2010) Use of camera-trapping to estimate puma density and influencing factors in central Brazil. *Journal of Wildlife Management* **74**, 1195–1203. doi:10.1111/j.1937-2817.2010.tb01240.x
- Nichols JD, Karanth KU, O'Connell AF (2011). Science, Conservation, and Camera Traps. In 'Camera traps in animal ecology: Methods and analyses'. (Eds AF O'Connell, JD Nichols, KU Karanth) (Springer: London)
- Norouzzadeh MS, Nguyen A, Kosmala M, Swanson A, Palmer MS, Packer C, Clune J (2018) Automatically identifying, counting, and describing wild animals in camera-trap images with deep learning. *Proceedings of the National Academy of Sciences* **115**, E5716–E5725. doi:10.1073/pnas.1719367115
- O'Brien TG (2011) Abundance, density and relative abundance: A conceptual framework. In 'Camera traps in animal ecology: Methods and analyses'. (Eds AF O'Connell, JD Nichols, KU Karanth) pp. 71–96. (Springer: London)
- O'Connell AF, Nichols JD, Karanth KU (2011) Preface. In 'Camera traps in animal ecology: Methods and analyses'. (Eds AF O'Connell, JD Nichols, KU Karanth) (Springer: London)
- Paton AJ, Buettel JC, Brook BW (2021) Evaluating scat surveys as a tool for population and community assessments. *Wildlife Research* **49**(3), 206–214. doi:10.1071/WR21056
- Paull DJ, Claridge AW, Cunningham RB (2012) Effective detection methods for medium-sized grounddwelling mammals: a comparison between infrared digital cameras and hair tunnels. *Wildlife Research* 39, 546–553. doi:10.1071/WR12034

- Perkins GC, Kutt AS, Vanderduys EP, Perry JJ (2013) Evaluating the costs and sampling adequacy of a vertebrate monitoring program. *Australian Zoologist* **36**, 373–380. doi:10.7882/AZ.2013.003
- Perry ME, Robertson AW (2012) Cleared and uncleared pellet plots as indices of brown hare density. *New Zealand Journal of Ecology* **36**, 157–163.
- Pfeffer SE, Spitzer R, Allen AM, Hofmeester TR, Ericsson G, Widemo F, Singh NJ, Cromsigt JPGM (2018) Pictures or pellets? Comparing camera trapping and dung counts as methods for estimating population densities of ungulates. *Remote Sensing in Ecology and Conservation* **4**, 173–183. doi:10.1002/rse2.67
- R Core Team (2020) R: A language and environment for statistical computing. R Foundation for Statistical Computing, Vienna, Austria. Available at https://www.R-project.org/
- Roberts NJ (2011) Investigation into survey techniques of large mammals: surveyor competence and camera-trapping vs. transect-sampling. *Bioscience Horizons* **4**, 40–49. doi:10.1093/biohorizons/hzr006
- Rönnegård L, Sand H, Andrén H, Månsson J, Pehrson Å (2008) Evaluation of four methods used to estimate population density of moose Alces alces. Wildlife Biology 14, 358–371. doi:10.2981/0909-6396(2008)14[358:EOFMUT]2.0.CO;2
- Rovero F, Marshall AR (2009) Camera trapping photographic rate as an index of density in forest ungulates. *Journal of Applied Ecology* **46**, 1011–1017. doi:10.1111/j.1365-2664.2009.01705.x
- Seidlitz A, Bryant KA, Armstrong NJ, Calver MC, Wayne AF (2021) Sign surveys can be more efficient and cost effective than driven transects and camera trapping: a comparison of detection methods for a small elusive mammal, the numbat (*Myrmecobius fasciatus*). *Wildlife Research* **48**, 491–500. doi:10.1071/WR20020
- Silveira L, Jácomo ATA, Diniz-Filho JAF (2003) Camera trap, line transect census and track surveys: a comparative evaluation. *Biological Conservation* **114**, 351–355. doi:10.1016/S0006-3207(03)00063-6
- Smart JCR, Ward AI, White PCI (2004) Monitoring woodland deer populations in the UK: an imprecise science. *Mammal Review* 34, 99–114. doi:10.1046/j.0305-1838.2003.00026.x
- Smith AD (1964) Defecation rates of mule deer. Journal of Wildlife Management 28, 435–444. doi:10.2307/3798195
- Southwell C (1989) Techniques for monitoring the abundance of kangaroo and wallaby populations. In 'Kangaroos, Wallabies and

Rat-kangaroos, Vol. 2'. (Eds G Grigg, P Jarman, I Hume) pp. 659–693. (Surrey Beatty and Sons Pty Limited: Chipping Norton)

- Steenweg R, Whittington J, Hebblewhite M, Forshner A, Johnston B, Petersen D, Shepherd B, Lukacs PM (2016) Camera-based occupancy monitoring at large scales: power to detect trends in grizzly bears across the Canadian Rockies. *Biological Conservation* 201, 192–200. doi:10.1016/j.biocon.2016.06.020
- Swan M, Di Stefano J, Christie F, Steel E, York A (2014) Detecting mammals in heterogeneous landscapes: implications for biodiversity monitoring and management. *Biodiversity and Conservation* 23, 343–355. doi:10.1007/s10531-013-0604-3
- Torney CJ, Lloyd-Jones DJ, Chevallier M, Moyer DC, Maliti HT, Mwita M, Kohi EM, Hopcraft GC (2019) A comparison of deep learning and citizen science techniques for counting wildlife in aerial survey images. *Methods in Ecology and Evolution* **10**, 779–787. doi:10.1111/2041-210X.13165
- Towerton AL, Penman TD, Kavanagh RP, Dickman CR (2011) Detecting pest and prey responses to fox control across the landscape using remote cameras. *Wildlife Research* **38**, 208–220. doi:10.1071/WR10213
- Triggs B (2003) 'Tracks, Scats and Other Traces; a field guide to Australian mammals.' (Oxford University Press: Melbourne)
- Vine SJ, Crowther MS, Lapidge SJ, Dickman CR, Mooney N, Piggott MP, English AW (2009) Comparison of methods to detect rare and cryptic species: a case study using the red fox (*Vulpes vulpes*). *Wildlife Research* **36**, 436–446. doi:10.1071/WR08069
- Wacher T, Attum O (2005) Preliminary investigation into the presence and distribution of small carnivores in the Empty Quarter of Saudi Arabia through the use of a camera trap. *Mammalia* **69**, 81–84. doi:10.1515/mamm.2005.007
- Wearn OR, Glover-Kapfer P (2019) Snaphappy: camera traps are an effective sampling tool when compared with alternative methods. *Royal Society Open Science* **6**, 181748. doi:10.1098/rsos.181748
- Weinstein BG (2018) A computer vision for animal ecology. Journal of Animal Ecology 87, 533–545. doi:10.1111/1365-2656.12780
- Wintle BA, Elith J, Potts JM (2005) Fauna habitat modelling and mapping: A review and case study in the Lower Hunter Central Coast region of NSW. *Austral Ecology* **30**, 719–738. doi:10.1111/j. 1442-9993.2005.01514.x
- Zuur AF, Ieno EN, Walker NJ, Saveliev AA, Smith GM (2009) 'Mixed Effects Models and Extensions in Ecology with R.' (Springer: New York)

Data availability. The data that support this study will be shared upon reasonable request to the corresponding author.

Conflicts of interest. The authors declare no conflicts of interest.

Declaration of funding. This research was funded by Parks Victoria.

Acknowledgements. We thank Emily Green (Parks Victoria) for assistance with image processing and Steven Shelley (Parks Victoria) for assistance with GIS mapping.

#### Author affiliations

<sup>A</sup>School of BioSciences, The University of Melbourne, Vic. 3010, Australia.

<sup>B</sup>Parks Victoria, Environment and Science Division, Level 10, 535 Bourke Street, Melbourne, Vic. 3000, Australia.

 $^{
m C}$ School of Ecosystem and Forest Sciences, University of Melbourne, 4 Water Street, Creswick, Vic. 3363, Australia.

<sup>D</sup>Parks Victoria, Eastern Region, Meeniyan-Promontory Road, Yanakie, Vic. 3960, Australia.