

Age estimation of striped marlin (*Kajikia audax*) in the eastern North Pacific using otolith microincrements and fin spine sections

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Abstract. The age of striped marlin (*Kajikia audax*) in days and years (daily and yearly ages) were estimated by counts of otolith microincrements ($n = 25$) and dorsal fin spine annuli ($n = 175$) using specimens caught in the tropical eastern North Pacific between September and November 2004. Daily ages of small striped marlin (87.0–145.5 cm lower jaw–fork length) were estimated to range from 81 to 239 days, which indicates that the species in the area grow, on average, to over 100 cm within 4 months. Back-calculated hatch dates were estimated to be from March to July; this period is earlier than the known spawning season in a slightly more northern area. Approximately 20% of these year-0 striped marlin had one or more growth bands in the sectioned spine. They were thought to be false annual growth bands and excluded from yearly age estimation. The estimated yearly age of 175 individuals (87.0–228.4 cm) ranged from 0 to 5 and was dominated by ages-0 to 3 (>90%). Age composition was different among three subregions in the eastern North Pacific.

Additional keywords: billfish, Istiophoridae.

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Introduction

The age of billfishes (families Istiophoridae and Xiphiidae) has been estimated using fin spine (or ray) sections for decades (e.g. Hill *et al.* 1989; Ehrhardt 1992; Speare 2003; Chiang *et al.* 2004; Kopf *et al.* 2005). However, early growth rates (up to age-1) of billfishes estimated by otolith microincrements are known to differ to those estimated by fin spines (Prince *et al.* 1991; DeMartini *et al.* 2007; Kopf *et al.* 2011). Recent studies have used otolith microincrements to estimate the early growth of billfishes, and these results have also been used in the estimation of the position of the first-year annual growth band in sectioned fin spines (DeMartini *et al.* 2007; Kopf *et al.* 2011; Shimose *et al.* 2015). This method successfully eliminates counting false annual growth bands formed in younger ages (e.g. Kopf *et al.* 2011; Shimose *et al.* 2015). However, otolith microincrements can only be counted up to age-1 (or age-2) in most billfishes (Prince *et al.* 1991; DeMartini *et al.* 2007). Furthermore, fin spine sections are still used in the ageing of individuals older than 1 or 2 years (e.g. DeMartini *et al.* 2007; Kopf *et al.* 2011; Shimose *et al.* 2015).

The striped marlin (*Kajikia audax*) is a large istiophorid billfish species targeted by commercial and recreational fisheries in the Pacific Ocean (Nakamura 1985; Whitelaw 2003). The age and growth of striped marlin, which provide important information for stock assessment, have been estimated in several previous studies (e.g. Skillman and Yong 1976; Melo-Barrera

et al. 2003; Kopf *et al.* 2005), and a recent study provided reliable growth parameters for the western South Pacific population (Kopf *et al.* 2011). There are several genetically discrete stocks of striped marlin in the Pacific, and the western South Pacific stock is isolated from stocks in the North Pacific (McDowell and Graves 2008). Average body size of striped marlin also differs, and is smaller in the eastern North Pacific (González-Armas *et al.* 2006) than in the western South Pacific (Kopf *et al.* 2005). Therefore, the age composition and growth patterns may be different among regions and should be estimated independently. The aim of this study was to estimate the age of striped marlin in the eastern North Pacific simultaneously in days and years (daily and yearly ages). The daily age of young striped marlin was estimated by counting otolith microincrements, and the growth rate was compared with that in the western South Pacific (Kopf *et al.* 2011). Yearly age of young to adult striped marlin was estimated by counting annual growth bands in sectioned fin spines, and age composition was compared among subregions in the tropical eastern North Pacific.

Materials and methods

Sample collection

In all, 26 otolith and 175 fin spine samples of striped marlin were collected during the longline research cruise by *RV Shoyo-Maru*

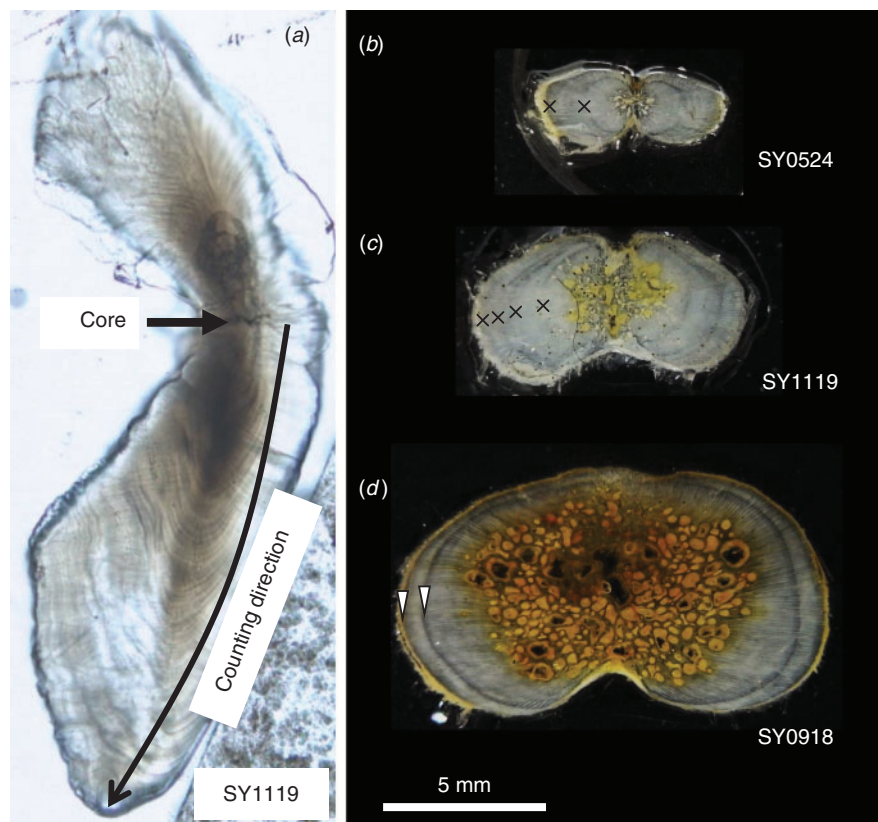


Fig. 1. Photographs of transversely sectioned sagittal otolith and dorsal fin spines of the striped marlin (*Kajikia audax*). (a, c) Sections from a 145.5-cm lower jaw–fork length (LJFL) female with 239 microincrements; (b) section from a 106.8 cm LJFL female; and (d) section from a 211.8 cm LJFL female. White arrowheads indicate assumed annual growth bands. Crosses indicate assumed false annual growth bands. Two specimen identification numbers on the photographs (SY0524 and SY1119) relate to the specimens listed in Table 1.

in the tropical areas of the eastern North Pacific Ocean (5–18°N, 103–134°W) between September and November 2004. All 26 otolith samples were from same fish used for fin spine samples. The sampling area was divided into three subregions based on previous studies (Shimose *et al.* 2010, 2013), namely an open ocean area (16–18°N, 118–134°W), a near-continental area (13–16°N, 103–107°W) and a near-equatorial area (5°N, 104–120°W). Bodyweight (BW) was recorded to the nearest 0.05 kg, and the lower jaw–fork length (LJFL) was measured to the nearest 0.1 cm for each specimen. Sex was also determined when possible through visual inspection of gonad morphology for potential use in a future study. However, sexual differences were not examined in the present study because of the insufficient sample size. The LJFL and BW of the specimens used for the otolith microincrement analysis were in the range 87.0–145.5 cm and 2.00–17.40 kg. The LJFL and BW of the specimens used for yearly age estimation based on fin spines were 87.0–228.4 cm and 2.00–78.40 kg. Much larger blue marlin (265 cm LJFL, >160 kg BW) were caught in this research cruise (Shimose *et al.* 2010, 2013), and the currently used longline gear can catch juvenile (~2.0 kg) to adult (~>100 kg) striped marlin.

Observation of otolith microincrements

Right or left sagittal otoliths were used in the otolith microincrement analysis. In this study, transverse sections were observed under light microscopy. The otoliths were embedded in resin and transversely sectioned to ~0.5 mm before being attached to glass slides. These sections were roughly polished until the core appeared on the surface, and polished again with 1-μm abrasive powder. Polished sections were observed under an optical microscope with transmitted light, and microincrements were counted three times (in December 2017 and in June and August 2018) by one reader (T. Shimose) without previous information on the individual specimen (Fig. 1). Otolith microincrements were identified referring to Prince *et al.* (1991) and Shimose *et al.* (2015). Otolith microincrements were assumed to be formed daily as in blue marlin (*Makaira nigricans*; Prince *et al.* 1991) and striped marlin (Kopf *et al.* 2011). The precision of the three counts was quantified by the average percentage error (APE; Beamish and Fournier 1981) and the CV (Campana 2001). Many ageing studies have been conducted with CVs of <7.6, which corresponds to an APE of 5.5 (Campana 2001). The median value of the three counts was used as the daily age. Back-calculated hatch dates were compared

Table 1. Estimated age in days (daily age), ageing precision (average percentage error (APE) and CV; see text for details) and back-calculated hatch dates for 25 striped marlin *Kajikia audax* caught in the tropical eastern North Pacific
ID, identification; BW, bodyweight; F, female; LJFL, lower jaw–fork length; M, male

Number	ID number	Catch date	LJFL (cm)	BW (kg)	Sex	Daily age	APE	CV	Hatch date
1	SY0738	5 October 2004	87.1	2.65	M	81	7.26	9.44	16 July 2004
2	SY0588	1 October 2004	98.9	3.55	F	85	8.47	11.16	8 July 2004
3	SY0688	4 October 2004	87.0	2.00	F	90	4.96	7.26	6 July 2004
4	SY0447	29 September 2004	93.1	2.70	M	93	6.44	8.43	28 June 2004
5	SY0860	8 October 2004	106.0	4.30	F	94	8.39	11.94	6 July 2004
6	SY0762	5 October 2004	97.2	3.46	F	104	3.59	4.87	23 June 2004
7	SY0748	5 October 2004	100.0	3.64	M	106	3.89	5.07	21 June 2004
8	SY0795	6 October 2004	110.5	5.30	M	115	3.85	5.64	13 June 2004
9	SY0381	27 September 2004	96.1	3.65	F	119	6.87	10.04	31 May 2004
10	SY0524	30 September 2004	106.8	4.35	F	119	3.83	4.99	3 June 2004
11	SY0650	3 October 2004	106.3	5.00	M	119	2.85	3.73	6 June 2004
12	SY0885	8 October 2004	108.2	5.47	F	119	5.70	7.83	11 June 2004
13	SY0522	30 September 2004	102.4	4.20	F	120	6.99	9.30	2 June 2004
14	SY0914	19 October 2004	91.2	3.05	F	121	3.63	4.99	20 June 2004
15	SY0331	26 September 2004	94.9	3.30	F	122	2.15	2.79	27 May 2004
16	SY0488	29 September 2004	100.4	3.60	F	125	3.67	4.92	27 May 2004
17	SY0366	26 September 2004	106.5	4.35	F	128	2.56	3.35	21 May 2004
18	SY0385	27 September 2004	95.4	3.30	M	132	2.35	3.41	18 May 2004
19	SY0686	4 October 2004	110.8	4.85	M	132	1.34	1.90	25 May 2004
20	SY0423	27 September 2004	102.0	4.70	M	133	1.68	2.31	17 May 2004
21	SY0858	8 October 2004	117.6	7.80	F	139	2.53	3.36	22 May 2004
22	SY0348	26 September 2004	120.3	6.70	M	144	4.30	6.23	5 May 2004
23	SY0736-2	6 October 2004	95.0	2.75	F	145	4.69	6.26	14 May 2004
24	SY0878	8 October 2004	122.7	7.50	M	181	0.98	1.39	10 April 2004
25	SY1119	28 October 2004	145.5	17.40	F	239	4.78	6.69	3 March 2004
Mean			104.1	4.78		124	4.31	5.89	

with the known spawning season off Mexico (24°N, 110°W; González-Armas *et al.* 2006).

Observation of fin spines

Sample processing and terminology of growth bands followed Shimose *et al.* (2015). The longest spine of the first dorsal fin was boiled to remove tissues and was preserved in dry storage conditions for later analysis. The fin spines were sectioned to ~0.5 mm. The position of sectioning was at a distance of half a condyle base width up from the condyle base (Sun *et al.* 2002; Chiang *et al.* 2004). Spine sections were attached to glass slides and photographed under a microscope. The translucent zone was treated as a growth band, and the annual growth band was identified according to Speare (2003) and Kopf *et al.* (2010, 2011) to avoid the counting of false annual growth bands (Fig. 1). Assumed annual growth bands (distinct translucent growth bands) were counted once by one reader (T. Shimose). Prior to the count, all sections were carefully checked for their growth bands and compared with previous studies (Kopf *et al.* 2010, 2011). Some specimens aged <1 year by the otolith microincrement count had some translucent growth bands, and they were assumed to have been formed before 1 year of age. The diameter of these growth bands (up to ~8 mm) was considered as a criterion of non-annual growth bands. Periodicity in growth band formation was not assessed in this study because of the limited sampling season.

Results and discussion

Daily age

The sectioned otoliths of striped marlin exhibited microincrements from the core to the edge except for one individual male (141.9 cm). The daily age of the remaining 25 individual striped marlin was successfully estimated (Table 1). The mean APE and CV of three counts in 25 individuals were 4.31 and 5.89 respectively; these values were less than the average values (APE = 5.5, CV = 7.6) reported in the literature (Campana 2001), and imply that the current counts are precise. The number of otolith microincrements of the 25 individuals (87.0–145.5 cm, mean 104.1 cm) ranged from 81 to 239 (mean 124), which indicates that, on average, striped marlin in the area grow over 100 cm in 4 months (Fig. 2). This result of the age–length relationship overlapped with the formerly estimated relationship in the western South Pacific (Kopf *et al.* 2011), and the growth rate is thought to be similar between the two areas at least up to 6 months. For the one individual male for which otolith microincrements could not be counted, this was due to the low legibility after inflection of the dorsal and ventral arms of the sectioned otolith.

Otolith length increases as fish body length increase, and only otolith weight continues to increase with age after growth in body length has almost stopped (Newman *et al.* 1996). Otolith weight is linked to otolith thickness, which continues to increase

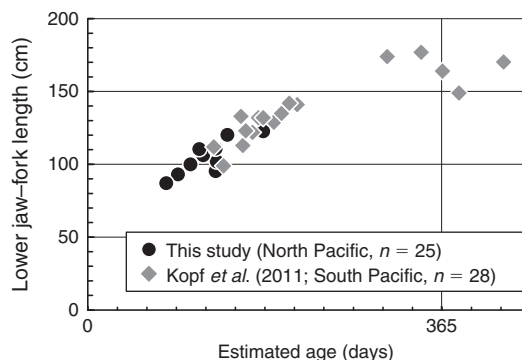


Fig. 2. Relationships between estimated age (days) and lower jaw-fork length of the striped marlin (*Kajikia audax*) in the tropical eastern North Pacific. Data for the South Pacific, from Kopf *et al.* (2011), are also plotted.

after inflection of the dorsal and ventral arms of the otolith in billfishes (Wilson *et al.* 1991). This means that the ageing of billfish for the same body size but older ages (i.e. slower growing) tends to fail more through counting their daily age from the core to the edge. This can lead to an overestimation of the growth rate because slower-growing individuals (older individuals within the same body size group) are excluded from the estimation.

The back-calculated hatch dates were between 3 March and 17 July and concentrated in May and June (Fig. 3). Maturing and mature females are known to be abundant in July and August in an area at a higher latitude off Mexico (24°N, 110°W; González-Armas *et al.* 2006) than the present study area (5–18°N, 103–134°W). Back-calculated hatch dates of the present study (May and June) are also earlier than the spawning season off Mexico. Spawning season is possibly early in the lower-latitude area, where temperature starts to increase earlier during the spring to summer transition.

Yearly age

Five of 25 observed small individuals (20%) for which daily age was estimated to be less than 365 (<1 year) had some growth bands in their sectioned fin spine (Fig. 1). Therefore, these growth bands (up to ~8 mm in diameter) were not annuli and should not be included in the count as annuli. Similar growth bands that formed before 1 year of age have also been reported in blue marlin (Shimose *et al.* 2015) and they are thought to be a common phenomenon in billfishes.

Estimated ages of the 175 striped marlin ranged from 0+ to 5+ years, with >90% of samples dominated by 0+ to 3+ years. Older striped marlin (up to 8 years of age) have been reported in the western South Pacific (Kopf *et al.* 2011), and the specimens used in the present study may not cover the full size range of striped marlin in the eastern North Pacific. However, different age compositions were observed among the three subregions (Fig. 4). In the near-continental area, a significant number of year-0 striped marlin occurred, but there were few year-0 individuals in the open ocean area. Striped marlin aged from 1+ to 3+ years were observed with high frequency in all three areas. Spawning grounds occur near the continent off Mexico (González-Armas *et al.* 2006) and in the near-continental area

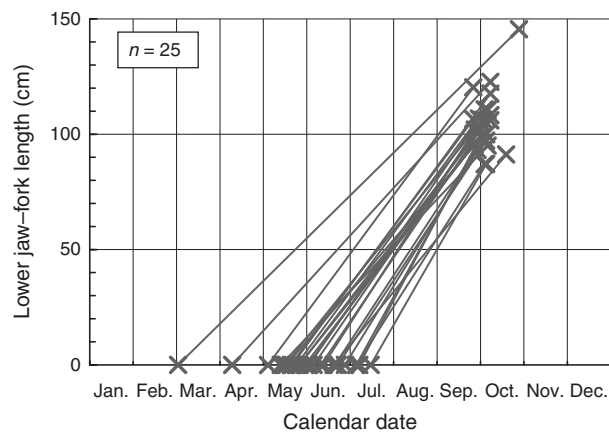


Fig. 3. Relationships between lower jaw-fork length at the catch date and back-calculated hatch date of striped marlin (*Kajikia audax*) in the tropical eastern North Pacific.

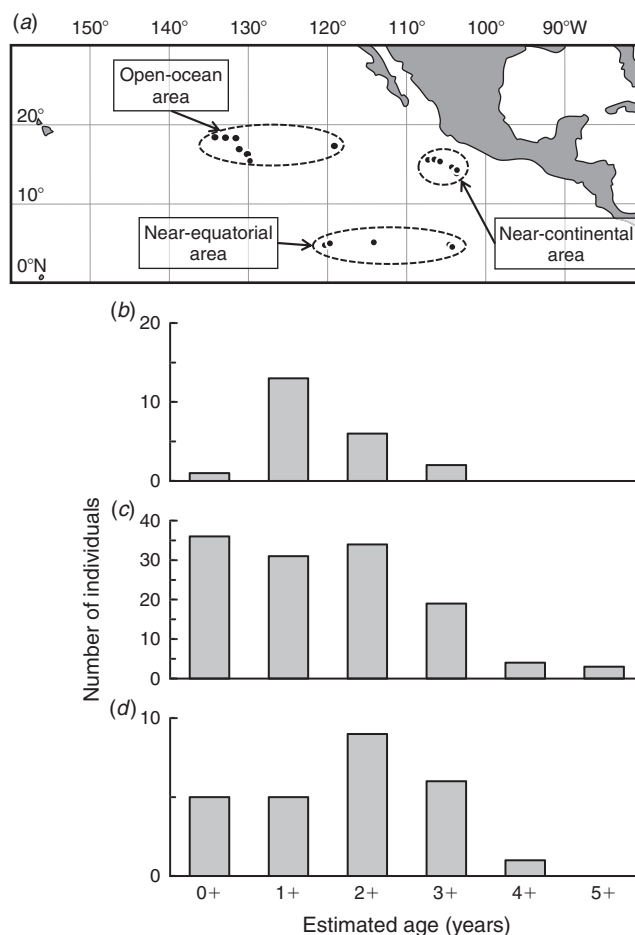


Fig. 4. Age composition of striped marlin (*Kajikia audax*) in three subregions of the tropical eastern North Pacific. (a) Map showing the location of the three subregions. (b–d) Age distribution of individuals in the open ocean area (b; $n = 22$), near-continental area (c; $n = 127$) and near-equatorial area (d; $n = 26$).

(Shimose *et al.* 2013). The age composition in the near-continental area is thought to reflect the occurrence of both juveniles and adults. Prey items are more abundant in the open ocean area than in the near-continental area (Shimose *et al.* 2010), and striped marlin start migrating to the open ocean area for feeding after reaching 1 year of age.

The results of this study will contribute to further our understanding of the life history of striped marlin relating to earlier growth and ontogenetic migration in the eastern North Pacific. Growth parameters of striped marlin were not estimated in this study because the sampling coverage was at specific points, both temporally and spatially, and the range of sizes sampled was limited due the sampling method. Further studies on age composition and growth curve estimation of striped marlin are needed based on representative samples in the eastern North Pacific.

Conflicts of interest

The authors declare that they have no conflicts of interest.

Declaration of funding

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