

Evaluating predator control using two non-invasive population metrics: a camera trap activity index and density estimation from scat genotyping

Jessica L. Keem^{A,*} , Bronwyn A. Hradsky^A , Joe Benshemesh^B, Mark Le Pla^C , Abigail Watkins^D, Andrew R. Weeks^{E,F} , Anthony van Rooyen^F , John Black^F and Darren Southwell^{A,G} 

For full list of author affiliations and declarations see end of paper

***Correspondence to:**

Jessica L. Keem
 Quantitative and Applied Ecology Group,
 School of Ecosystem and Forest Sciences,
 University of Melbourne, Parkville,
 VIC 3010, Australia
 Email: jessica.keem@unimelb.edu.au

Handling Editor:

Thomas Prowse

Received: 2 February 2022
Accepted: 27 August 2023
Published: 12 September 2023

Cite this:

Keem JL *et al.* (2024)
Wildlife Research **51**, WR23033.
 doi:[10.1071/WR23033](https://doi.org/10.1071/WR23033)

© 2024 The Author(s) (or their employer(s)). Published by CSIRO Publishing.

This is an open access article distributed under the Creative Commons Attribution-NonCommercial-NoDerivatives 4.0 International License (CC BY-NC-ND).

OPEN ACCESS

ABSTRACT

Context. Invasive mammalian predators are a threat to biodiversity and agriculture globally, yet management outcomes for lethal predator control remain difficult to monitor and evaluate. Understanding whether changes in activity indices correspond to true changes in population density will help inform effective monitoring and management programs. **Aims.** The aim of this study was to evaluate the effect of poison baiting on invasive red fox (*Vulpes vulpes*) populations using two alternative population metrics: fox activity from camera trap surveys and density estimation from scat genetic analysis. **Methods.** We conducted before–after control–impact studies in two regions of semi-arid Australia (Wimmera and Mallee) by monitoring paired non-treatment and treatment sites during unbaited and baited periods. We estimated the effects of poison baiting on: (1) a monthly fox activity index, derived from an array of 10 off-road camera traps per site; and (2) fox density. To estimate density, we collected fox scats along 14-km transects, identified individuals using polymorphic microsatellite DNA markers and fitted spatially explicit capture–recapture models. **Key results.** Fox activity remained consistently low at all sites except the Mallee non-treatment. The top-ranked models of fox activity and density contained an interaction between *treatment* and *period*, with an interactive and additive effect of *region*, respectively. However, there was little evidence that baiting reduced fox activity or density. In the unbaited period, fox densities ranged from 0.69 (95% CI: 0.47–1.0) to 1.06 (95% CI: 0.74–1.51) foxes km⁻² and were similar across regions. **Conclusions.** Camera traps have the potential to provide continuous index-based measures of fox populations but may not record sufficient observations to detect change. Indices can also be confounded by variations in animal behaviour. Scat genetic analysis is a viable option for providing direct estimates of population change at specific snapshots in time; however, this approach is considerably more expensive, and large sample sizes may be required if genotyping success is low. **Implications.** Our study presents a rare example of multiple concurrent – and non-invasive – monitoring techniques to evaluate the effectiveness of predator control. We highlight the value of rigorous study designs and high-quality density information for designing predator management and monitoring programs.

Keywords: 1080 baiting, activity index, BACI design, density estimation, genetic sampling, *Leipoo ocellata*, non-invasive sampling, semi-arid, *Vulpes vulpes*.

Introduction

Invasive mammalian predators are global drivers of biodiversity decline and extinction, as well as livestock loss (Doherty *et al.* 2016; Fleming *et al.* 2017). Such predators thrive in novel landscapes and challenge ecosystems through predation, competition, hybridisation and disease (Daniels *et al.* 2001; Norbury 2001; Harris and Macdonald 2007; Wyatt *et al.* 2008; Spencer *et al.* 2017). Reducing invasive predator populations through targeted management is therefore considered a crucial component of biodiversity conservation

and economic safeguarding (Greentree et al. 2000; Robinson et al. 2013). There is a general expectation that lethal control reduces the density of invasive predators, but there are relatively few examples where appropriate data are collected to quantify change (Clayton and Cowan 2010; Walsh et al. 2012; Doherty et al. 2016). Unless the impacts of lethal control of invasive predators are measured and evaluated, the nature and magnitude of environmental improvements arising from such investments will remain uncertain.

One globally significant invasive mammalian predator is the European red fox (*Vulpes vulpes*; hereafter 'fox') (Lowe et al. 2000; Doherty et al. 2016). In Australia, predation by foxes is a key threat to native fauna and agriculture: foxes are implicated in the extinction of over 20 Australian native mammals, and management costs amount to more than \$16 million AUD per annum (McLeod 2004; Woinarski et al. 2015). A suite of techniques have been adopted to control foxes, including lethal poison baiting, shooting, fumigation, trapping, exclusion fencing and livestock guardian dogs (Saunders and McLeod 2007; van Bommel and Johnson 2012). Lethal poison baiting with sodium fluoroacetate (hereafter 'baiting'), commonly known as 1080, is the dominant management method for controlling fox populations in Australia (Saunders et al. 2010). However, the effects of 1080 baiting on fox population size or behaviour, and in turn the benefits to native species, are still poorly understood (Hunter et al. 2018; Manning et al. 2021).

Traditionally, the impacts of poison baiting on fox populations have largely been evaluated using index-based surrogates of fox abundance or density, such as bait-take rates, scat counts, spotlighting counts or track counts from sand pads (Sadlier et al. 2004; Reddiex et al. 2006). More recently, motion-triggered camera traps (hereafter 'camera traps') have become more affordable and therefore increasingly used to monitor the effects of fox management (e.g. Towerton et al. 2011; Benshemesh et al. 2014). Camera traps are a passive, non-invasive monitoring method that can be deployed across large spatial and temporal scales at low labour investment (Wearn and Glover-Kapfer 2019). Consequently, camera traps are potentially a cost-effective method to monitor cryptic species like the fox, which are nocturnal and capable of occupying large home ranges at low densities (Vine et al. 2009).

Camera traps are an effective tool for detecting fox presence (occupancy) if deployed for a sufficiently long period, but their value for detecting changes in fox population density is debatable. The uniform pelage of foxes prevents individual identification (Güthlin et al. 2014a; Carter et al. 2019), making it impossible to conduct mark-recapture analysis based on photographs. Methods for estimating true density from camera trap data without individual recognition are in development but remain largely unvalidated (Rowcliffe et al. 2008; Ramsey et al. 2015; Nakashima et al. 2018; Schaus et al. 2020). Instead, population changes in response to baiting are commonly inferred from relative abundance indices – such as activity – based on the photographic

trapping rate (e.g. van Hesperen et al. 2019). As with all activity indices, photographic trapping rates are a product of both population density and the observation process (Sollmann et al. 2013; Güthlin et al. 2014b; Stephens et al. 2015). This means that they are unable to distinguish changes in fox behaviour (that affect detectability) from true changes in population density.

Highly polymorphic genetic markers such as microsatellite loci provide an alternative approach for estimating fox density, by facilitating mark-recapture analysis without requiring invasive trapping and marking of individual foxes (Piggott and Taylor 2003; Le Pla et al. 2022). DNA can be obtained passively from epithelial cells shed in the environment and collected via hair-snares or scats (Sadlier et al. 2004; Immell and Anthony 2008; Ruell et al. 2009). These samples are genotyped to identify individual foxes, producing a dataset amenable to capture-recapture analysis (Royle and Young 2008; Wegge et al. 2019). Genetic techniques are becoming more cost-effective, and this approach has great potential. However, genotyping microsatellite markers can be both relatively expensive and time-intensive, and hair and scat samples often contain low quality DNA, meaning that a large quantity of samples is usually required to compensate for low genotyping success rates (Piggott 2004; Piggott et al. 2008; Berry et al. 2012).

Testing and comparing alternative methods to measure the effectiveness of invasive predator management is crucial to improving reliability in monitoring actions and hence the accuracy and precision of conservation initiatives (e.g. Lazenby et al. 2014; Wilson et al. 2017). To date, however, studies that have compared fox density estimates derived from non-invasive genotyping to activity indices have only focused on indirect signs (such as bait take, sand plots or scat counts) (e.g. Marks et al. 2009; Le Pla et al. 2022); the comparative value of camera trap data is yet to be evaluated.

Our study was conducted in association with the large-scale malleefowl Adaptive Management Predator Experiment (AMPE), which operates across southern Australia. The AMPE aims to learn about the effects of fox and cat (*Felis catus*) management on predator activity, and in turn, malleefowl (*Leipoa ocellata*) breeding activity (Benshemesh et al. 2018; Hauser et al. 2019). Approximately 20 pairs of experiment sites have been established across the malleefowl's range. Fox baiting occurs in and around treatment sites, whereas paired non-treatment sites are left unmanaged. Camera traps operate continuously at all AMPE sites to monitor background rates of predator activity. However, whether the camera data can detect reductions in fox activity associated with a reduction in density due to baiting needs evaluation.

Our study measured the effect of baiting on fox populations at two paired non-treatment/treatment sites within the AMPE. These sites are patches of remnant native vegetation in the Wimmera and Mallee regions of semi-arid Victoria, Australia. Within each region, we aimed to evaluate the effects of baiting on: (1) fox activity using data from continuously

operating camera traps; and (2) fox density using scat genotyping of microsatellite markers and spatially explicit capture–recapture analysis. We compared estimates of baiting effectiveness, based on these two different approaches while accounting for region-based differences in baiting regimes and environmental context. Comparative analysis of alternative sampling methods is crucial for designing optimal monitoring programs. Our findings help infer whether a camera trap network deployed across AMPE sites in southern Australia is likely to detect changes in fox density if fox density is affected by baiting. Our study also provides some of the first baseline estimates of fox density for semi-arid Australia, a key parameter for designing effective invasive predator management programs.

Materials and methods

Study sites

Our broad study design was replicated across the Wimmera and Mallee regions of Victoria, Australia. Within each region, we conducted a before–after control–impact (BACI) experiment, with repeated surveys through time at one non-treatment and one treatment site (Fig. 1). Baiting had previously occurred at both treatment (or ‘impact’) sites prior to initial surveying. Treatment sites in our study were managed for foxes by Parks Victoria and the Mallee Catchment Management Authority after initial surveying, respectively, with nearby non-treatment sites left unmanaged as experimental controls.

In the Wimmera region, the paired sites comprised the non-treatment site, Nurcounng Flora Reserve (36°41′39.8″S, 141°42′42.0″E), and the treatment site approximately 10 km away, the Eastern block of Little Desert National Park (36°34′50.0″S, 141°48′55.0″E) (Fig. 1a). The area of native vegetation at the non-treatment site is 5.9 km², and the treatment site has an area of 476.8 km². Both sites are bordered by pastoral and arable agricultural land. The Wimmera bioregion experiences hot, dry summers and cool winters with an average rainfall of 414 mm, falling primarily from June to August (data for Nhill, the nearest meteorological station; Bureau of Meteorology 2020a). The landscape typically supports Lowan Sands Mallee (Ecological Vegetation Class 87, *Eucalyptus* spp, *Leptospermum* spp.) (Victorian Government Department of Sustainability and Environment 2005a).

The Mallee region paired sites comprised the non-treatment site at Wandown Flora and Fauna Reserve (34°48′24.9″S, 142°59′20.9″E), and the treatment site approximately 30 km away at Annuello Flora and Fauna Reserve (34°53′38.7″S, 142°33′33.7″E) (Fig. 1b). The area of native vegetation at the non-treatment site has an area of 25.3 km², and the treatment site an area of 361.0 km². Both sites are bordered by irrigated agriculture on the northern side, and pastoral and arable agricultural land elsewhere. The Mallee bioregion also has a hot climate, with dry

summers and cool winters. Rainfall averages 328.3 mm, with a strong bias across August–October (Data for Ouyen, the nearest meteorological station; Bureau of Meteorology 2020b). Vegetation is dominated by Woorinen Sands Mallee (Ecological Vegetation Class 86, *Eucalyptus* spp, *Triodia* spp.) (Victorian Government Department of Sustainability and Environment 2005b).

Fox-baiting regime

The non-treatment sites had no history of fox management and were not managed during our study. Pre-existing management at treatment sites ceased a month prior to our study commencing in both regions: June 2019 in the Wimmera, and May 2019 in the Mallee (Fig. 2). We consider this window to be unbaited in our study design but note that fox densities may be lower in treatment sites as a result of prior management.

Both treatment sites were ground baited (Animal Control Technologies, FOXOFF[®]) following our unbaited scat surveys. Although we consulted with the land managers, the timing, extent and intensity of baiting at treatment sites was largely out of our control and dependent on the objectives and constraints of the organisations.

At the Wimmera treatment site, the Eastern and Central blocks have been pulse baited since 2010. This involved three 9-week pulses per year with fortnightly bait replacement. The unbaited period of our study was from June 2019 to September 2019. An intensified baiting regime commenced in late September 2019, and our baited survey period followed in October 2019 (Fig. 2). While pulse baiting continued in the central block, baits were laid monthly at the eastern block. Across the two blocks, 345 bait stations were operated, positioned approximately 750 m apart (Supplementary Fig. S1).

The Mallee treatment site was first baited in 2019. Baits were deployed across two pulses, one in April and May. The unbaited period of our study was from June 2019 to November 2019. The baited period of our study commenced with pulse baiting in December 2019 and continued into the next baiting event in April 2020. Baits were laid at 223 stations, spaced approximately 260 m apart. Each baiting pulse lasted for 2 weeks, and any removed baits were replaced daily.

Camera traps

All four sites were monitored with camera traps (Wimmera: Scout Guard SG560k and Ltl Acorn Ltl-5310; Mallee: Scout Guard SG560k) to estimate fox activity. The AMPE continuously operates 10 solar-powered cameras at all sites ($N = 40$; Fig. 1). These cameras have all been operating since 2015, except for the Mallee treatment site (deployed in 2019).

We obtained 12 months (June 2019–May 2020) of raw camera data from the Victorian Malleefowl Recovery Group for each site. We grouped these camera data into unbaited and baited periods, specific to each region (Fig. 2). Note, baiting commenced at the Wimmera treatment site only on

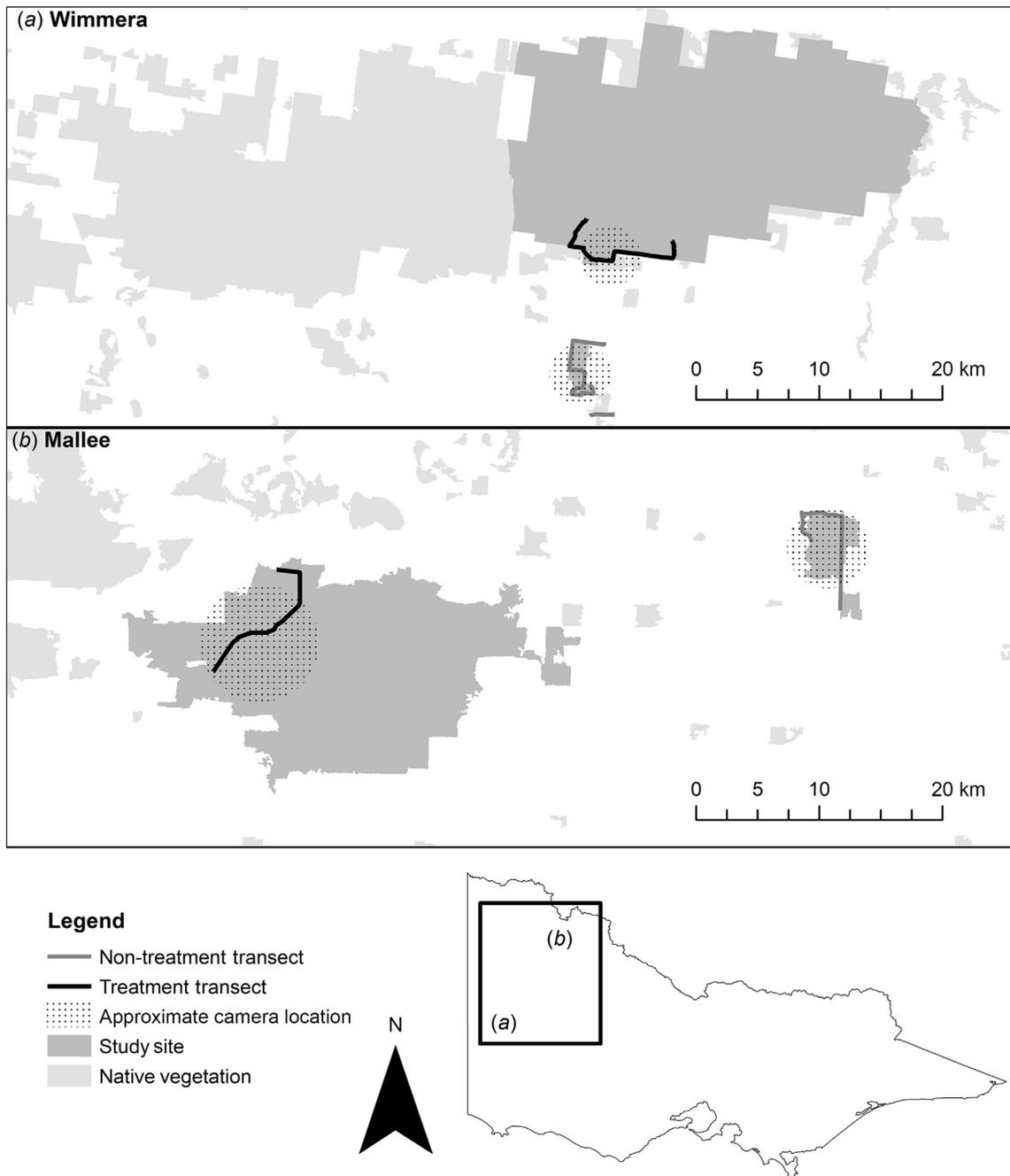


Fig. 1. Location of the four study sites in (a) the Wimmera region and (b) the Mallee region in Victoria, Australia, used for camera trap surveys and scat collection. Baiting was conducted at treatment sites (black transect), whereas the non-treatment sites (grey transect) remained unbaited. Approximate camera sites are spread within the areas represented by black stippling, close to transects; precise locations are not shown for security reasons.

the last day of September, therefore we treated this month as unbaited.

The camera setup followed the approach described by van Hespén (2015) and van Hespén *et al.* (2019). Cameras were 1 m above ground in an off-track, grid formation, and spaced 0.8 km apart (Fig. 1). They were programmed to capture one photograph with a 5-min interval between sequential triggers

to minimise repeat captures, and were not fitted with a lure to avoid altering predator behaviour (Benshemesh *et al.* 2014). Although camera spacing was not independent relative to fox movements, the design captured the ‘background’ rate of fox activity (therefore predation opportunities) at sites and accounted for potential spatial and temporal heterogeneities in fox activity (van Hespén *et al.* 2019).

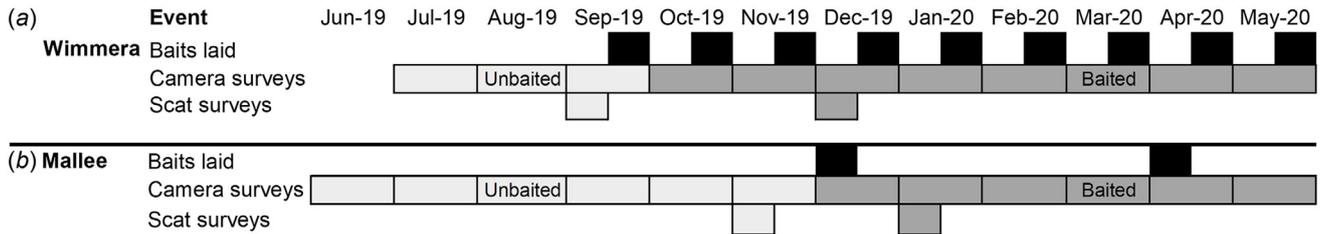


Fig. 2. Timeline of study events at the (a) Wimmera and (b) Mallee region. Poison baits were laid (black) at the treatment sites. For each region, the months before the first baiting event were the unbaited period (light grey), and those after baiting were the baited period (dark grey). Note, baiting commenced at the Wimmera treatment site only on the last days of September, therefore we treated this month as unbaited.

Scat collection

We established a 14-km scat collection transect at each site, with a minimum of 8-km displacement between transect start and end (Fig. 1). Transects were based along roads adjacent to the camera traps at each site. This placement ensured that we maximised the number of scats collected (because roads are regions of high fox activity in mixed-agricultural landscapes; Hradsky *et al.* 2017) and that we surveyed a similar area as the camera traps. Transect length was approximately double the length of average fox home range estimates (95% kernel) from the nearest analogous bioregion (Carter *et al.* 2012). This ensured that the sampling area was likely to be greater than the range of any individual, an important model assumption for spatially explicit capture–recapture analysis (Efford and Boulanger 2019). Where feasible, transects were continuous for logistical reasons; otherwise, we split transects to preferentially survey along connected native vegetation (Fig. 1a).

We surveyed each transect across two primary survey sessions per region, one in the unbaited period and another in the baited period (Fig. 2). We cleared each transect of fox scats 2 days prior to a survey session so that only fresh scats would be collected. We aimed to collect scats across four secondary survey occasions (survey days) per survey session, alternating between the non-treatment and treatment transects over eight consecutive days. This ensured that scats were less than 48 h old and reduced the likelihood of DNA degradation (Piggott 2004). In total, there were eight survey sessions: two regions × two sites × two periods.

The survey sessions in the unbaited periods were conducted in full in both regions: Wimmera (September 2019) and Mallee (November 2019). For the baited period in the Wimmera region, one survey occasion at the non-treatment site was lost due to a Total Fire Ban day (December 2019). In the Mallee, the first survey occasion in the baited period was delayed until 7 days after scats were cleared due to a Bushfire State of Emergency, and we were only able to complete two and three survey occasions at the non-treatment and treatment site, respectively (January 2020).

Transects were walked simultaneously by two observers to collect fox scats from the middle of the road and up to 2 m

either side. At all times, one observer was experienced in scat identification and the other had been trained for at least 1 day prior. We identified the species based on visual and odour specifications (Triggs 2004) and associated signs (Moseby *et al.* 2012). One disposable glove was used per scat to avoid cross-contamination of DNA. Scats were placed into individual zip-lock bags and marked with date, site, unique identifier and collector according to Piggott and Taylor (2003). We preserved scats frozen at -18°C immediately upon returning from collection (Piggott and Taylor 2003).

Analytic approach

We used a model comparison approach to evaluate support for an effect of baiting on fox activity (derived from camera records) and fox density (derived from scat genotypes).

We hypothesised that three main effects could influence fox activity and density: *treatment* (non-treatment or treatment site); *period* (unbaited or baited); and *region* (Wimmera or Mallee). We derived a set of candidate models for each response variable (activity rate and density), based on single, additive and interactive combinations of these effects, and a null model. In each case, support for models that included an interaction effect between *treatment* and *period* would be considered evidence that baiting affected fox activity or density. The *region* covariate was included to represent differences in the landscape context and baiting regime between the regions.

We ranked candidate models using Akaike's information criterion adjusted for small sample size (AICc) (Burnham and Anderson 2002). We regarded the 'best' model to be the one with the lowest AICc value but considered those within two AICc as equally plausible. To measure precision around our estimates, we calculated the percentage relative standard error (RSE) as the standard error of the coefficient divided by the estimate; an RSE > 20% was assumed to indicate low precision and, therefore, low statistical power to detect a change in the population size (Williams *et al.* 2002; Palmer *et al.* 2018; Efford and Boulanger 2019). All statistical analyses were performed in R v4.2.1 (R Core Team 2022).

Activity analysis

We identified fox captures from camera traps using FastStone software (Benshemesh et al. 2018; FastStone 2020). To minimise non-independent captures of the same individual, we classed repeat captures within a 30-min time frame as a single capture event using the ‘CamTrapR’ package (Niedballa et al. 2016). We then extracted the total number of independent fox captures per site for each camera-month. We assigned months into unbaited and baited periods (as detailed above; Fig. 2).

We modelled fox activity as the mean fox capture rate (number of captures per month) using generalised linear mixed effect models (GLMMs). To account for excess zeros in monthly fox capture rates (Blasco-Moreno et al. 2019), we assessed the fit of four alternative distributions (Poisson, negative binomial, zero-inflated Poisson and zero-inflated negative binomial) using simulation for dispersion and zero-inflation with ‘DHARMA’ (Hartig 2020). *P*-values suggested a zero-inflated negative binomial provided the best fit to the data, given by:

$$P_{c,m} \sim \text{ZINB}(\eta_{c,m}, \alpha) \tag{1}$$

where $P_{c,m}$ is the number of captures taken by camera c in month m , $\eta_{c,m}$ is the mean capture rate and α is the variance of the distribution.

We fitted models using maximum likelihood methods, using the ‘glmmTMB’ package (Magnusson et al. 2021). We derived a set of 12 candidate models, based on combinations of the fixed effects and a null model (Table 1). To decide whether we needed to account for resampling at the same camera trap points each month, we evaluated our most complex fixed-effects model (*treatment* × *period* × *region*), with or

without inclusion of camera trap point identity (1–10) as a random effect. We then used the structure of the most strongly supported model to fit the 12 candidate fixed effects models. We assessed competitive models with a combination of model weights, 95% confidence intervals of interaction terms and R^2 values (calculated as per Nakagawa and Schielzeth (2013)).

DNA extraction and species identification

Scats were removed from the freezer prior to DNA extraction and allowed to thaw before swabbing the outer surface of the scat repeatedly with a Copan dry swab (155CIS, Copan). Swabs were then placed in lysis buffer (500 µL ATL and 40 µL Proteinase K) in a 1.5-mL tube, with the swab tip snapped so that the tube lid could close. The tube was vortexed thoroughly, incubated at 56°C for 2 h before the sample was centrifuged at 6000g to pellet any debris. We then removed 200 µL of supernatant for extraction following the DNeasy spin column protocol with final elution in 100 µL buffer AE. Initially, we screened extracted DNA to confirm presence of fox DNA using the vv-CytB fox-specific primers in a qPCR assay described in Berry et al. (2012) on a Roche LightCycler 480 II system. A TaqMan Exogenous Internal Positive Control (VIC probe) was run for each sample to test for the presence of PCR inhibitors. Samples that were inhibited were then diluted 1/10 and rescreened with the qPCR assay.

All samples that were deemed positive for fox DNA (including diluted samples) were then genotyped by microsatellite loci and a single sex-specific marker to identify individuals and their sex. Microsatellite loci genotyped were V142, V374, V402, V602 (Wandeler and Funk 2006), C2054 (Francisco et al. 1996), DB1, DB3 and DB4 (Holmes et al. 1995), and

Table 1. Generalised linear mixed models of fox captures (C) in the Wimmera and Mallee regions, Victoria, Australia, fitted across 11 and 12 months, respectively.

Rank	Model	K	LogLk	AICc	ΔAIC _c	AICcwt	R ² m	R ² c
1	$C \sim (t \times p \times r)$	11	-502.10	1026.94	0.00	0.94	0.23	0.52
2	$C \sim (t + p + r)$	7	-510.40	1035.10	8.16	0.02	0.21	0.52
3	$C \sim (t \times r)$	7	-510.86	1036.02	9.08	0.01	0.22	0.50
4	$C \sim (t + r)$	6	-512.13	1036.48	9.54	0.01	0.21	0.52
5	$C \sim (t \times p)$	7	-511.18	1036.68	9.74	0.01	0.04	0.50
6	$C \sim (p + r)$	6	-512.24	1036.71	9.77	0.01	0.17	0.54
7	$C \sim (p \times r)$	7	-511.65	1037.61	10.67	0.00	0.17	0.54
8	$C \sim (r)$	5	-513.88	1037.92	10.98	0.00	0.17	0.53
9	$C \sim (t + p)$	6	-516.78	1045.79	18.85	0.00	0.04	0.50
10	$C \sim (p)$	5	-517.87	1045.91	18.97	0.00	0.00	0.51
11	$C \sim (.)$	4	-519.85	1047.82	20.88	0.00	0.00	0.51
12	$C \sim (t)$	5	-518.87	1047.91	20.97	0.00	0.03	0.50

Zero-inflated negative binominal models are defined with fox captures (C) as constant (.), or varying according to treatment (t), period (p) and region (r). Number of parameters (K), LogLk, ΔAIC_c and AICcwt follow Burnham and Anderson (2002); marginal R² (R²m) and conditional R² (R²c) follow Nakagawa and Schielzeth (2013).

the sex-determining marker was vvSRY (Berry *et al.* 2007). All markers were tagged with a unique fluorescent label during PCR using the method outlined in Blacket *et al.* (2012). Reactions included 5- μ L Qiagen multiplex mix (Qiagen, Chadstone, Victoria, Australia), 4 μ L of primer mix (0.2 μ M of each primer) and 2 μ L of template DNA. PCR conditions consisted of an initial 15-min denaturing step at 94°C, followed by 40 cycles of 94°C for 30 s, 59°C for 90 s and 72°C for 60 s, with a final extension step of 60°C for 30 min. All PCRs were undertaken on an Eppendorf Mastercycler S gradient (Eppendorf). All samples were genotyped in duplicate to avoid issues with allelic dropout.

After PCR, genotyping was undertaken using an Applied Biosystems 3730 capillary analyser, with product lengths determined relative to a GS500LIZ_3730 size standard. GeneMapper ver. 4.0 (Applied Biosystems) was used to score allele sizes, with all profiles examined manually. Individual fox haplotypes (multi-locus genotypes) were then identified with GenAlEx v6.5 (Peakall and Smouse 2012), as in Le Pla *et al.* (2022). If replicates matched at a locus, we assumed this to be the correct genotype, whereas if replicate genotypes differed at a locus, we then manually re-examined each histogram output to determine if there was an error in scoring, and if still conflicting, we excluded the locus from analysis. Individuals with six or more microsatellite loci scored across both replicate samples were included in all subsequent analyses to limit errors when identifying individuals (Waits *et al.* 2001).

Spatially explicit capture–recapture analysis

We estimated fox density using spatially explicit capture–recapture (SECR) analysis (Borchers and Efford 2008). SECR models the probability of detecting an animal as a declining function of distance from its range centre at an unknown location, and distance to the nearest trap location (Royle and Young 2008). Resulting models produce estimates of population density unbiased by edge effects and incomplete detection (Borchers and Efford 2008).

We fitted multisession SECR models to the data from the eight survey sessions. The multisession formulation was adopted because data aggregation can improve the precision of parameter estimations by sharing detection information across sessions (Efford *et al.* 2009) and regions (Morin *et al.* 2018; Proffitt *et al.* 2020).

Each 14-km transect was discretised into 200-m segments to create conceptual traps with scats assigned to the nearest segment (Efford 2011; Lonsinger *et al.* 2018; Le Pla *et al.* 2022). Each trap was considered a proximity detector to allow multiple detections per individual per occasion (Efford 2011). We defined a buffer distance of 3002 m around each transect as outer limit of the area of integration, where foxes beyond this estimate were considered unlikely to be detected in the survey (Borchers and Efford 2008). This value was based on double the diameter of the average home range (95%

kernel) of GPS-collared foxes in the nearest similar climate zone (Carter *et al.* 2012). The buffer distance exceeded the minimum suggested buffer size of 2664 m, based on an estimate of dispersion, the root-pooled spatial variance (Calhoun and Casby 1958; Efford *et al.* 2009).

We modelled density with the ‘secr’ package (Efford 2020) by fitting the full-likelihood model assuming a half-normal detection function:

$$D \sim 1, g_0 \sim 1, \sigma \sim 1 \quad (2)$$

where D is density, g_0 is the probability of detecting an animal at a trap located at its activity centre (per occasion) and σ is a home range spatial scale parameter (Efford *et al.* 2009).

As for the activity analysis, we had a set of 12 density models. Further, we considered that fox detectability might vary with *period* due to seasonal differences in behaviour, or with *sex* if females and males had differences in movement behaviour (e.g. male foxes generally disperse further than females) (Trehwella *et al.* 1988). Therefore, for each density model, we varied the detectability parameter (g_0) and spatial scale parameter (σ) as constant, or as a function of *period* or *sex*. This resulted in 22 candidate models (Table S1).

Results

Fox activity

Across 11 and 12 survey months, we recorded 106 fox detections (27 unbaited, 79 baited) from 4503 trap nights in the Wimmera, and 483 detections (267 unbaited, 216 baited) from 6782 trap nights in the Mallee, respectively. This corresponded to a higher camera-trapping rate in the Mallee at 7.12 foxes per 100 trap nights, as compared with 2.35 foxes per 100 trap nights in the Wimmera. In the Wimmera region, foxes were detected at a total of 44% and 57% of the functioning camera traps at the non-treatment ($n = 9$) and treatment ($n = 7$) sites, respectively. In the Mallee region, foxes were detected at 100% of the functioning camera traps at the non-treatment ($n = 9$) and treatment ($n = 10$) sites.

A zero-inflated negative binomial distribution with a random effect for camera traps provided the best fit to the fox activity data. The top-ranked model was strongly supported over the other candidate models (Δ AICc to next model 8.16; model weight = 0.94; Table 1). The top model included a three-way interaction among *treatment*, *period* and *region*. However, this was primarily driven by a change in fox activity at the non-treatment site in the Mallee.

There was no evidence that fox baiting reduced fox activity in either region. In the Wimmera region, fox activity was low and similar at both sites across the unbaited and baited periods (Fig. 3a). In the Mallee region, fox activity was higher at the non-treatment than treatment site during the unbaited period; estimates overlapped during the baited period due to a

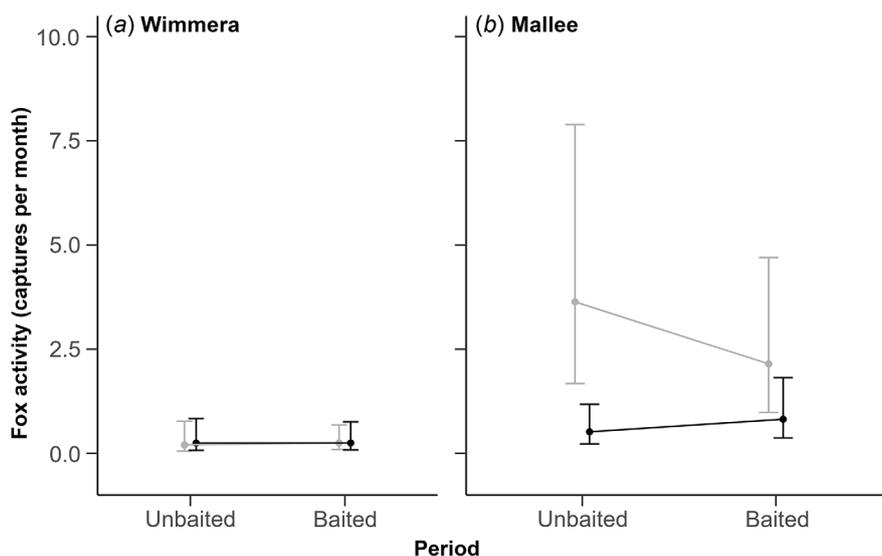


Fig. 3. Predicted fox activity estimates ($\pm 95\%$ CI) from the top-ranked model. Activity was derived from the monthly fox capture rate from 10 camera traps at each site. Estimates for non-treatment sites are light grey and treatment sites are black. Panels show estimates for the unbaited and baited periods in (a) the Wimmera and (b) the Mallee region.

large but uncertain decline in fox activity at the non-treatment site (Fig. 3b). The coefficient for the three-way interaction term was moderate and positive, but uncertain (1.19; 95% CI: -0.20 – 2.57). This uncertainty likely reflects the high level of variation in capture rates among camera traps within sites, which resulted in poor precision around most of the activity estimates (Table S2).

There was no support for the simpler model that could also indicate an effect of baiting on fox activity (*treatment* \times *period*); Table 1. The fixed effects for this model had little explanatory power ($R^2_m = 0.04$; Table 1).

Scat collection

In total, we collected 638 scats across the four sites. The number of scats collected per survey session varied between regions and periods. The greatest number of scats was detected from the Mallee region (Table 2). Scat numbers were lower in the baited periods, with approximately 60–70% fewer scats detected at all sites (Table 2), likely reflecting a seasonal difference.

Table 2. Total number of fox scats collected and genotyped, and percentage of genotyping success across the Wimmera and Mallee regions, Victoria, Australia.

Region	Wimmera				Mallee				Total
	Non-treatment		Treatment		Non-treatment		Treatment		
	Unbaited	Baited	Unbaited	Baited	Unbaited	Baited	Unbaited	Baited	
Scats collected	98	36	69	26	156	44	159	50	638
Scats genotyped	52	26	34	8	41	26	51	28	266
Genotyping success (%)	53	72	49	31	26	59	32	56	42

Genotyping success and recapture rates

Of the 638 scats, 285 (45%) yielded fox DNA and 266 (42%) amplified enough microsatellite loci ($n \geq 6$) to assign individual identities; 372 samples were unable to be genotyped (Table 2). We identified 108 individual foxes (Table 3). No individual foxes were detected moving between sites or regions, suggesting spatial independence of study sites.

In the Wimmera region, we detected 51 unique individuals; 67% of these were at the non-treatment site (Table 3). At the non-treatment site, no fox contributed to more than 8% of all detections ($n = 78$ scats; Table 3). However, at the treatment site, one fox accounted for 26% of all detections ($n = 42$ scats; Table 3). In the Mallee, we detected a similar number of individuals to the Wimmera (57), but the number of individuals was approximately evenly split between the non-treatment and treatment sites (Table 3). No fox accounted for more than 13% of detections at the non-treatment site ($n = 67$ scats; Table 3). At the treatment site, recaptures were heavily skewed towards a few individuals, with three foxes contributing 41% of all detections ($n = 79$ scats; Table 3).

Table 3. Number of fox individuals, detections and recaptures across the Wimmera and Mallee region, Victoria, Australia.

Region Site Period	Wimmera				Mallee			
	Non-treatment		Treatment		Non-treatment		Treatment	
	Unbaited	Baited	Unbaited	Baited	Unbaited	Baited	Unbaited	Baited
Females (n)	15	9	6	4	10	10	13	6
Males (n)	12	7	8	3	12	6	12	5
Total (n)	27	16	14	7	22	16	25	11
New individuals (n)	27	7	14	3	22	8	25	2
Recaptures within period (n)	25	10	20	1	19	10	26	17
Recaptures between periods (propr.)	–	0.56	–	0.57	–	0.50	–	0.82

'New individuals' is the number of distinct individuals detected for the first time in each period. 'Recaptures within period' are repeat detections of distinct individuals within a survey period. 'Recaptures between periods' is the proportion of unique individuals detected that had already been detected in the previous period.

Across all sites, many individuals detected during the baited periods had previously been detected in the unbaited periods. Recapture rates between the periods ranged between 50% and 82% (Table 3) and were similar or higher in the treatment sites than the non-treatment sites, suggesting that these individuals survived baiting, and that baiting did not increase population turnover (Table 3).

Spatially explicit capture–recapture density modelling

Nine models of fox density received substantial support (Table 4). Model uncertainty was high and individual model weight was low, partly due to uncertainty around the detectability parameters: four density models with an identical density model but with or without an effect of *sex* on the detectability parameters received very similar support (Table 4).

The top-ranked model contained an effect of *sex* on the detectability parameters, an interaction between *treatment* and *period* and an additive effect of *region*, indicating some evidence for an effect of baiting on fox density (Table 4; Fig. 4). However, uncertainty around the interaction term was wide (0.55; 95% CI: –0.17–1.26), and two other models

without this interaction term received similar support (Table 4). The top-ranked model suggested that g_0 was lower for female foxes (0.05; 95% CI: 0.04–0.08) than male foxes (0.08; 95% CI: 0.06–0.12). However, the σ value was larger for female foxes (726.78 m; 95% CI: 606.52–870.88), as compared with male foxes (569.01 m; 95% CI: 481.51–672.41).

The top-ranked model suggested that fox densities were similar between regions (Fig. 4). Density estimates in the unbaited period ranged from 0.69 (95% CI: 0.47–1.0) to 1.06 (95% CI: 0.74–1.51) foxes km⁻² across regions. Precision was highest at each site and region in the unbaited period, with RSE \leq 20% (Table S3). Fox densities in both regions declined more at treatment sites (35–40% decline) than non-treatment sites (12–14% decline) between the survey periods, although confidence intervals still overlapped pre-baiting estimates (Fig. 4).

Discussion

Measuring the effectiveness of lethal predator control is crucial for determining appropriate management actions

Table 4. Full likelihood SECR models of fox density in the Wimmera and Mallee regions, Victoria, Australia, fitted across eight sampling sessions.

Rank	Model	npar	LogLk	AICc	Δ AICc	AICcwt
1	$D \sim (t \times p + r) g_0 \sim (s) \sigma \sim (s)$	9	–1104.30	2228.00	0.00	0.13
2	$D \sim (t + p + r) g_0 \sim (s) \sigma \sim (s)$	8	–1105.44	2228.00	0.00	0.13
3	$D \sim (t \times p + r) g_0 \sim (.) \sigma \sim (.)$	7	–1106.60	2228.07	0.07	0.13
4	$D \sim (t + p + r) g_0 \sim (.) \sigma \sim (.)$	6	–1107.75	2228.14	0.14	0.12
5	$D \sim (t) g_0 \sim (s) \sigma \sim (s)$	6	–1108.11	2228.87	0.87	0.08
6	$D \sim (t) g_0 \sim (.) \sigma \sim (.)$	4	–1110.42	2229.14	1.14	0.07
7	$D \sim (t) g_0 \sim (p) \sigma \sim (p)$	6	–1108.28	2229.21	1.21	0.07
8	$D \sim (t + p \times r) g_0 \sim (s) \sigma \sim (s)$	9	–1105.21	2229.82	1.82	0.05
9	$D \sim (t + p \times r) g_0 \sim (.) \sigma \sim (.)$	7	–1107.51	2229.89	1.89	0.05

Models are defined with fox density (D) as constant ($.$), or varying according to treatment (t), period (p) and region (r). Parameters g_0 (likelihood of detection) and σ (spatial scale) are defined as constant ($.$), or varying according to period (p) or sex (s). Number of parameters (npar), LogLk, Δ AICc and AICcwt follow Burnham and Anderson (2002). Only models within two AICc of the top-ranked model are provided. See Table S1 of the Supplementary material for models ranked 10–22.

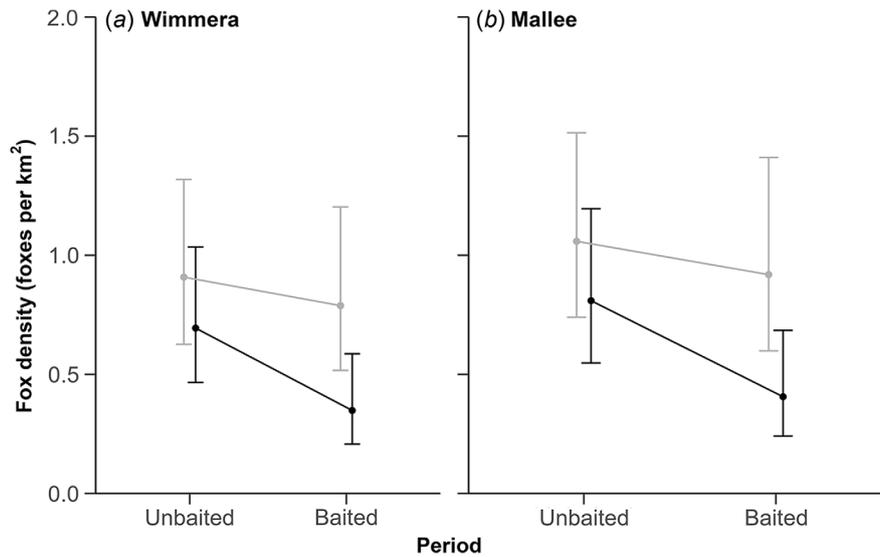


Fig. 4. Predicted fox density estimates ($\pm 95\%$ CI). Density was derived from scat genetic analysis and spatially explicit capture–recapture analysis. Estimates for non-treatment sites are light grey and treatment sites are black. Panels show estimates for the top-ranked model in the (a) Wimmera and (b) Mallee regions.

(Reddiex *et al.* 2006; Doherty and Ritchie 2017). Fox activity derived from camera traps showed no evidence of an effect of baiting but a high degree of variation among study sites and regions. Fox density estimates from scat genotyping were more consistent across regions and survey periods. There was weak evidence of a post-baiting decline in fox density, but a substantial number of individuals survived the baiting events in both regions. Our estimate of 0.69–1.06 foxes km^{-2} is the first empirical baseline fox density for semi-arid Victoria. Uncertainty around density estimates was much lower than uncertainty around activity estimates. Further increases in precision could be obtained by increasing the number of replicate surveys and genotyping success. This would likely increase statistical power to detect an effect of baiting on fox density if present. Our findings demonstrate the importance of high-quality information on predator density in the design and monitoring of predator management, as well as the need for cautious interpretation of relative abundance indices as standalone metrics.

Fox activity

Our top-ranked model of fox activity, derived from the AMPE camera trap data, provided no evidence that baiting decreased fox activity. Instead, fox activity differed greatly between the two regions. This variation was likely due to camera error and low sample size rather than a reflection of true difference. Three of the 20 camera traps in the Wimmera failed and did not record any photos. Others produced false zeros or were poorly aligned, leading to consistently low and imprecise fox capture rates.

Environmental factors may also have influenced fox behaviour and therefore detectability and activity. Fox activity was higher at the Mallee non-treatment site, perhaps because cameras at this smaller site tended to be closer to habitat edges and roads, where fox activity is known to concentrate (Šálek *et al.* 2010; Hradsky *et al.* 2017). Furthermore, fox breeding (winter) and cub births (spring) coincided with unbaited survey periods, and the baited survey periods included the dispersal season (autumn) (Pech *et al.* 1992). Seasonal changes in movement behaviour (such as females provisioning for cubs) may have also contributed to the higher initial fox activity at the Mallee non-treatment site. However, the expected increase in fox activity from cub independence and dispersal is not seen in the baited period. Although activity counts from camera traps are commonly employed to evaluate the outcomes of fox baiting (Towerton *et al.* 2011; van Hespén *et al.* 2019), these metrics may have limited value when population densities are low, or when environmental factors – or baiting – alter fox behaviour (Sollmann *et al.* 2013; Stephens *et al.* 2015). Our findings reiterate that under such circumstances, camera traps may not receive sufficient capture rates to detect population change.

Trapping rates on cameras in the Wimmera were similar to rates from two baited studies in similar bioregions: nearby in the Grampians, approximately 100 km to the south-east (2.35 foxes per 100 trap nights; Ramsey *et al.* 2015), and from semi-arid Western Australia (2.28 foxes per 100 trap nights; Thompson *et al.* 2019). However, the Mallee camera trapping rate was substantially smaller than the rate of approximately 24.67 foxes per 100 trap nights recorded at

the same non-treatment site 4 years prior to our study (van Hespén *et al.* 2019). The discrepancy between these trapping rates is likely caused by differences in how consecutive captures on cameras were handled: we used a 30-min interval to class detections as ‘independent’, whereas van Hespén *et al.* (2019) used 5 min.

Spatially explicit capture–recapture

Our top model of fox density, derived from the scat genotyping data, provided weak evidence of an effect of baiting on fox density. Fox densities were similar across both regions prior to baiting, and there was some evidence that densities at the treatment sites declined more than at the non-treatment sites among the survey periods. It is important to note that fox densities at the treatment sites may have been impacted by prior baiting events. This may explain the tendency toward lower densities in treatment than non-treatment sites prior to baiting. The BACI design allowed us to quantify the effects of the next management pulse on the fox populations, while controlling for seasonal changes by monitoring changes in the non-impact landscapes. However, it was likely to be conservative in terms of quantifying an overall impact of baiting on unmanaged fox populations.

The identification of individual foxes using microsatellite genetic markers provided powerful information on fox survival rates in the baited period, revealing that more than half the foxes at each treatment site were unaffected by management actions. These foxes may not be encountering baits, not consuming baits or not consuming baits while the baits contain toxic levels of 1080. This suggests that managers need to explore options for improving population knockdown rates (rather than focus on reducing immigration). Such options could include modifying the baiting regime or method of bait delivery. For example, to increase the number of baits available to each fox, managers could increase bait density or the frequency of baiting (Saunders and McLeod 2007; Towerton *et al.* 2016). The likelihood of foxes consuming a toxic dose will decrease with bait avoidance and caching (Allsop *et al.* 2017). Trialling other bait matrices to improve palatability (Van Polanen Petel *et al.* 2001), trialling other poisons with greater efficacy (Southwell *et al.* 2011; Gentle *et al.* 2017) or combining baits with canid pest ejectors (Marks *et al.* 2002) may result in more rapid consumption of baits. Scat genetic analysis is evidently a valuable technique to refine baiting and monitoring regimes and for estimating fox densities.

Our study was able to establish the first empirical density estimates of fox populations for the semi-arid regions of Victoria. These estimates are broadly comparable with those from other semi-arid and arid grazing habitat in Australia (e.g. Marlow 1992; Marlow *et al.* 2000; Berry *et al.* 2012). They are higher than estimated densities from the Grampians, approximately 100 km south-east, which estimated 0.22 foxes km⁻²

using presence–absence camera trap data of foxes without individual identification (Ramsey *et al.* 2015). Our density estimates are also substantially greater than those from the closest analogous bioregion (approximately 200 km north-west). Here, concurrent GPS collaring and camera trap spatially explicit mark–resight analysis provided estimates of 0.05–0.14 foxes km⁻² (Carter *et al.* 2019). In contrast, however, densities were substantially lower than those observed in Australian urban and temperate agricultural habitats (Coman *et al.* 1991; Thompson and Fleming 1994; Marks and Bloomfield 1999; Berghout 2001; Gentle 2005); this was expected because foxes have larger home ranges and occur at lower densities when productivity is low.

Comparison of metrics

Despite considerable uncertainty associated with all metrics, density estimates produced consistently lower RSEs than the activity indices, indicating that scat genetic analysis had a greater capacity to detect a change in fox populations, if one occurred. One potential reason for this is that our scat survey transects covered a larger area than the camera arrays and were therefore likely to produce less variation in sample size and more-precise estimates (Güthlin *et al.* 2014b).

Further, operating error and seasonal variation in fox behaviour led to low and variable fox capture rates on the camera traps. Activity indices can be skewed if detections are sparse or variable (Sollmann *et al.* 2013). Where capture rates are low, as exemplified in the Wimmera region, an activity index will be biased by detectability and thus have limited capacity to infer trends in density (Sollmann *et al.* 2013; Stephens *et al.* 2015). Nonetheless, our activity models indicated some promise for inferring population density: Mallee fox activity approximately corresponded with the suggested decline in density produced by SECR analysis. This suggests that camera trap activity indices may have the potential to parallel trends in density, given appropriate capture rates.

Survey design – camera activity index

Simple changes that optimised survey effort would likely improve the statistical power of the camera survey design. Although increasing the survey period would reduce uncertainty in our spatially replicated study, it is irrelevant for long-term monitoring programs such as the AMPE. Instead, survey effort would best be increased by ensuring that all cameras are functioning and not obscured. The detection rate of foxes at each camera could also be increased by placing cameras at areas of higher fox activity, such as on tracks (Mann *et al.* 2015; Carter *et al.* 2019; Geyle *et al.* 2020). However, camera placement on tracks has several limitations that need to be considered, including spatial variation in track networks and capture rates (e.g. Carter *et al.* 2019; Kolowski *et al.* 2021), and temporal variation in fox behaviour, such as dispersal

patterns (Soulsbury *et al.* 2011). On-track cameras may also be logistically difficult for long-term monitoring programs due to concerns of theft and vandalism (e.g. Foster 2008; A. Watkins, pers. comm., May 2020). Lures and attractants may be an alternative to increase fox visitation rate and therefore capture rate (Meek *et al.* 2014) but again, may raise logistical concerns with maintenance.

Survey design – scat genotyping for density estimates

With spatially explicit capture–recapture analyses, the precision of the density estimate increases as the number of unique individuals and recaptures increases (albeit, non-linearly). That is, the relative standard error of the density estimate is inversely related to the square root of the minimum of either the number of unique individuals genotyped or the number of recapture events (Efford and Boulanger 2019). In our study, the number of individuals and recaptures were acceptably high (>20) for most of the unbaited surveys but were generally low (<10) during the baited surveys at both treatment and non-treatment sites. This likely reduced the statistical power of our study to detect an effect of fox baiting on fox densities, if one occurred (Efford and Boulanger 2019). Sample sizes could be boosted by improving genotyping success. Of the collected scat samples, only 42% of scats with extracted DNA were assigned individual identities, a rate comparable with similar nearby studies (Piggott *et al.* 2008; A. Weeks, unpubl. data). Degradation of scat DNA from environmental exposure can significantly increase microsatellite genotyping error (Murphy *et al.* 2007; Brinkman *et al.* 2010). Our survey periods overlapped extreme weather events, which exposed scat DNA to thunderstorms, dust storms and long intervals between collections. These conditions also prevented us conducting several surveys in the post-baiting sessions; more repeat surveys would have resulted in the collection of more scats for these sessions.

Three key assumptions may also have impacted the accuracy of our density estimates, primarily through bias in fox detectability. First, we assumed equal scat collection survey effort among observers. If observer experience affected detection of individual foxes, the detectability component of the SECR model may have underestimated fox density (Borchers and Efford 2008). Second, we assumed that fox detectability followed a half-normal function, and therefore that large detection distances were improbable. Given that a transect survey design only informs fox detection within two directions of its activity centre, a half-normal detection function is a conservative estimate of detection probability (Efford 2011; Efford *et al.* 2016; Efford 2019). Third, our primary survey sessions were paired and relatively short, so we assumed demographic closure within a survey session. The ‘secr’ closure test function confirmed all sites and periods were closed to gains and losses, except the Mallee non-treatment site in the baited period where data were insufficient to test for

violation detect (Efford *et al.* 2009). However, baited surveys may have captured transient foxes due to timing of juvenile independence and subadult dispersal (Pech *et al.* 1992). Where possible, we advise future studies to time scat collection surveys during periods of stable fox population dynamics to minimise any confounding effects of season, or to repeat surveying to lower this uncertainty.

Management implications

Our study highlights the many challenges that face land managers when monitoring and managing a cryptic, wide-ranging invasive predator. We reiterate the importance of rigorous study design and setup in detecting population change. Camera traps can be a long-term and cost-effective option for detecting elusive, low-density species (Cutler and Swann 1999; Moruzzi *et al.* 2002). However, without an informed power analysis or pilot study, poor detection rates and limited numbers of cameras can reduce their power to detect changes in predator activity, so careful consideration must be given to the make and number of cameras, their set-up, positioning and maintenance (Meek *et al.* 2015; Newey *et al.* 2015; Findlay *et al.* 2020; Kolowski *et al.* 2021). Additionally, activity indices are prone to bias if behavioural changes from baiting, season or habitat affect detectability, obscuring changes in density (Sollmann *et al.* 2013; Gütthlin *et al.* 2014b; Stephens *et al.* 2015).

Non-invasive scat genetic analysis is a powerful alternative for detecting predators and monitoring population change (e.g. Gopalaswamy *et al.* 2012; Anile *et al.* 2014; Rodgers *et al.* 2014; Wegge *et al.* 2019). This methodology can be used to inform management actions, providing useful information on relatedness, survival and population turnover, as well as direct density estimates to refine baiting and monitoring programs, or as an initial reference value for individual-based population simulations (e.g. Hradsky *et al.* 2019). Technological advances are expected to reduce costs and increase genotyping success rates, improving precision in density estimates and making scat collection more cost-efficient (Morin and McCarthy 2007; A. Weeks, unpubl. data). However, scat genetic analysis, paired with SECR, provides only ‘snapshots’ of density in time, and is unlikely to be financially suitable for continuous monitoring of invasive predator populations.

Camera trap surveys and scat genetic analysis are potentially most useful in tandem, because they can provide multiple lines of evidence about the effectiveness of invasive predator management. Where density estimates achieve a desirable level of precision, they could also be used to calibrate activity indices (e.g. Hopkins and Kennedy 2004; Siddig *et al.* 2015). Exploring these two metrics collectively has the potential to improve overall monitoring; for example, density estimates from SECR could be used to design appropriate camera trap arrays, and camera traps could locate hotspot areas of fox activity to target scat collection.

Our study is one of the few existing comparisons of relative and absolute predator population density metrics under lethal management actions. Our results suggested some evidence of an effect of baiting on fox density but also high survival rates, a finding supported by a similar study in the wet forests of southern Victoria, Australia (Le Pla *et al.* 2022). We therefore add to the growing body of literature emphasising the importance of evidence-based management actions. Above all, we demonstrate the many challenges associated with the monitoring and management of invasive mammalian predators, and the need for cautious interpretation of relative abundance indices as standalone metrics.

Supplementary material

Supplementary material is available [online](#).

References

- Allsop SE, Dundas SJ, Adams PJ, Kreplins TL, Bateman PW, Fleming PA (2017) Reduced efficacy of baiting programs for invasive species: some mechanisms and management implications. *Pacific Conservation Biology* **23**, 240–257. doi:10.1071/PC17006
- Anile S, Ragni B, Randi E, Mattucci F, Rovero F (2014) Wildcat population density on the Etna volcano, Italy: a comparison of density estimation methods. *Journal of Zoology* **293**, 252–261. doi:10.1111/jzo.12141
- Benshemesh J, Stokie P, Thompson D, Irvin J, Macfarlane N, Willis K, Willis C, Cattanach P, Bannerman M, Davies S (2014) Motion-sensitive cameras for monitoring a range of animals in malleefowl monitoring sites. In 'Proceedings of the 5th National Malleefowl Forum'. pp. 151–161. (Printak Pty Ltd: Adelaide, South Australia)
- Benshemesh J, Southwell DM, Lahoz-Monfort JJ, Hauser CE, Rumpff L, Bode M, Burnard T, Wintle BA (2018) The national malleefowl monitoring effort: citizen scientists, databases and adaptive management. In 'Monitoring threatened species and ecological communities'. (Eds S Legge, DB Lindenmayer, NM Robinson, BC Scheele, DM Southwell, BA Wintle) pp. 389–398. (CSIRO Publishing: Australia)
- Berghout M (2001) The ecology of the red fox (*Vulpes vulpes*) in the central tablelands of New South Wales. PhD Thesis, University of Canberra, Canberra, Australia.
- Berry O, Sarre SD, Farrington L, Aitken N (2007) Faecal DNA detection of invasive species: the case of feral foxes in Tasmania. *Wildlife Research* **34**, 1–7. doi:10.1071/WR06082
- Berry O, Algar D, Angus J, Hamilton N, Hilmer S, Sutherland D (2012) Genetic tagging reveals a significant impact of poison baiting on an invasive species. *The Journal of Wildlife Management* **76**, 729–739. doi:10.1002/jwmg.295
- Blacket MJ, Robin C, Good RT, Lee SF, Miller AD (2012) Universal primers for fluorescent labelling of PCR fragments – an efficient and cost-effective approach to genotyping by fluorescence. *Molecular Ecology Resources* **12**, 456–463. doi:10.1111/j.1755-0998.2011.03104.x
- Blasco-Moreno A, Pérez-Casany M, Puig P, Morante M, Castells E (2019) What does a zero mean? Understanding false, random and structural zeros in ecology. *Methods in Ecology and Evolution* **10**, 949–959. doi:10.1111/2041-210x.13185
- Borchers DL, Efford MG (2008) Spatially explicit maximum likelihood methods for capture–recapture studies. *Biometrics* **64**, 377–385. doi:10.1111/j.1541-0420.2007.00927.x
- Brinkman TJ, Schwartz MK, Person DK, Pilgrim KL, Hundertmark KJ (2010) Effects of time and rainfall on PCR success using DNA extracted from deer fecal pellets. *Conservation Genetics* **11**, 1547–1552. doi:10.1007/s10592-009-9928-7
- Bureau of Meteorology (2020a) Climate statistics for Australian locations: Nhill. Available at http://www.bom.gov.au/climate/averages/tables/cw_078086.shtml [accessed 21 July 2020]
- Bureau of Meteorology (2020b) Climate statistics for Australian locations: Ouyen. Available at http://www.bom.gov.au/climate/averages/tables/cw_076047.shtml [accessed 21 July 2020]
- Burnham KP, Anderson DR (2002) 'Model selection and multimodel inference: a practical information-theoretic approach.' 2nd edn. (Springer-Verlag: New York) doi:10.1007/b97636
- Calhoun JB, Casby JU (1958) Calculation of home range and density of small mammals. *Public Health Monograph* **55**, 1–24.
- Carter A, Luck GW, McDonald SP (2012) Ecology of the red fox (*Vulpes vulpes*) in an agricultural landscape. 2. Home range and movements. *Australian Mammalogy* **34**, 175–187. doi:10.1071/AM11041
- Carter A, Potts JM, Roshier DA (2019) Toward reliable population density estimates of partially marked populations using spatially explicit mark–resight methods. *Ecology and Evolution* **9**, 2131–2141. doi:10.1002/ece3.4907
- Clayton R, Cowan P (2010) Management of animal and plant pests in New Zealand – patterns of control and monitoring by regional agencies. *Wildlife Research* **37**, 360–371. doi:10.1071/WR09072
- Coman BJ, Robinson J, Beaumont C (1991) Home range, dispersal and density of red foxes (*Vulpes vulpes* L.) in central Victoria. *Wildlife Research* **18**, 215–223. doi:10.1071/WR9910215
- Cutler TL, Swann DE (1999) Using remote photography in wildlife ecology: a review. *Wildlife Society Bulletin (1973-2006)* **27**, 571–581.
- Daniels MJ, Beaumont MA, Johnson PJ, Balharry D, Macdonald DW, Barratt E (2001) Ecology and genetics of wild-living cats in the north-east of Scotland and the implications for the conservation of the wildcat. *Journal of Applied Ecology* **38**, 146–161. doi:10.1046/j.1365-2664.2001.00580.x
- Doherty TS, Ritchie EG (2017) Stop jumping the gun: a call for evidence-based invasive predator management. *Conservation Letters* **10**, 15–22. doi:10.1111/conl.12251
- Doherty TS, Glen AS, Nimmo DG, Ritchie EG, Dickman CR (2016) Invasive predators and global biodiversity loss. *Proceedings of the National Academy of Sciences* **113**, 11261–11265. doi:10.1073/pnas.1602480113
- Efford MG (2011) Estimation of population density by spatially explicit capture–recapture analysis of data from area searches. *Ecology* **92**, 2202–2207. doi:10.1890/11-0332.1
- Efford MG (2019) Non-circular home ranges and the estimation of population density. *Ecology* **100**, e02580. doi:10.1002/ecy.2580
- Efford MG (2020) secr: spatially explicit capture–recapture models. R package version 4.3. Available at <https://CRAN.R-project.org/package=secr>
- Efford MG, Boulanger J (2019) Fast evaluation of study designs for spatially explicit capture–recapture. *Methods in Ecology and Evolution* **10**, 1529–1535. doi:10.1111/2041-210X.13239
- Efford MG, Borchers DL, Byrom AE (2009) Density estimation by spatially explicit capture–recapture: likelihood-based methods. In 'Modeling demographic processes in marked populations'. (Eds DL Thomson, EG Cooch, MJ Conroy) pp. 255–269. (Springer US: Boston, MA)
- Efford MG, Dawson DK, Jhala YV, Qureshi Q (2016) Density-dependent home-range size revealed by spatially explicit capture–recapture. *Ecography* **39**, 676–688. doi:10.1111/ecog.01511
- FastStone (2020) FastStone Image Viewer. Available at <https://www.faststone.org/>. [accessed 24 October 2020]
- Findlay MA, Briers RA, White PJC (2020) Component processes of detection probability in camera-trap studies: understanding the occurrence of false-negatives. *Mammal Research* **65**, 167–180. doi:10.1007/s13364-020-00478-y
- Fleming PJS, Ballard G, Reid NCH, Tracey JP (2017) Invasive species and their impacts on agri-ecosystems: issues and solutions for restoring ecosystem processes. *The Rangeland Journal* **39**, 523–545. doi:10.1071/RJ17046
- Foster RJ (2008) The ecology of jaguars in a human-influenced landscape. PhD Thesis, University of Southampton, England, United Kingdom.
- Francisco LV, Langsten AA, Mellersh CS, Neal CL, Ostrander EA (1996) A class of highly polymorphic tetranucleotide repeats for canine genetic mapping. *Mammalian Genome* **7**, 359–362. doi:10.1007/s00335990104
- Gentle M (2005) Factors affecting the efficiency of fox baiting practices on the central tablelands of New South Wales. PhD Thesis, University of Sydney, Sydney, Australia.
- Gentle M, Speed J, Allen BL, Harris S, Haapakoski H, Bell K (2017) The longevity of para-aminopropiophenone (PAPP) wild dog baits and the implications for effective and safe baiting campaigns. *Environmental*

- Science and Pollution Research International* **24**, 12338–12346. doi:10.1007/s11356-017-8668-3
- Geyle HM, Stevens M, Duffy R, Greenwood L, Nimmo DG, Sandow D, Thomas B, White J, Ritchie EG (2020) Evaluation of camera placement for detection of free-ranging carnivores; implications for assessing population changes. *Ecological Solutions and Evidence* **1**, e12018. doi:10.1002/2688-8319.12018
- Gopalaswamy AM, Royle JA, Delampady M, Nichols JD, Karanth KU, Macdonald DW (2012) Density estimation in tiger populations: combining information for strong inference. *Ecology* **93**, 1741–1751. doi:10.1890/11-2110.1
- Greentree C, Saunders G, Mcleod L, Hone J (2000) Lamb predation and fox control in south-eastern Australia. *Journal of Applied Ecology* **37**, 935–943. doi:10.1046/j.1365-2664.2000.00530.x
- Güthlin D, Storch I, Küchenhoff H (2014a) Is it possible to individually identify red foxes from photographs? *Wildlife Society Bulletin* **38**, 205–210. doi:10.1002/wsb.377
- Güthlin D, Storch I, Küchenhoff H (2014b) Toward reliable estimates of abundance: comparing index methods to assess the abundance of a mammalian predator. *PLoS ONE* **9**, e94537. doi:10.1371/journal.pone.0094537
- Harris DB, Macdonald DW (2007) Interference competition between introduced black rats and endemic galápagos rice rats. *Ecology* **88**, 2330–2344. doi:10.1890/06-1701.1
- Hartig F (2020) DHARMA: residual diagnostics for hierarchical (multi-level/mixed) regression models. R package 0.3.2.0. Available at <https://cran.r-project.org/web/packages/DHARMA/index.html>
- Hauser CE, Southwell D, Lahoz-Monfort JJ, Rumpff L, Benshemesh J, Burnard T, van Hespren R, Wright J, Wintle B, Bode M (2019) Adaptive management informs conservation and monitoring of Australia's threatened malleefowl. *Biological Conservation* **233**, 31–40. doi:10.1016/j.biocon.2019.02.015
- Holmes NG, Dickens HF, Parker HL, Binns MM, Mellersh CS, Sampson J (1995) Eighteen canine microsatellites. *Animal Genetics* **26**, 132–133. doi:10.1111/j.1365-2052.1995.tb02659.x
- Hopkins HL, Kennedy ML (2004) An assessment of indices of relative and absolute abundance for monitoring populations of small mammals. *Wildlife Society Bulletin* **32**, 1289–1296. doi:10.2193/0091-7648(2004)032[1289:AAOIOR]2.0.CO;2
- Hradsky BA, Robley A, Alexander R, Ritchie EG, York A, Di Stefano J (2017) Human-modified habitats facilitate forest-dwelling populations of an invasive predator, *Vulpes vulpes*. *Scientific Reports* **7**, 12291. doi:10.1038/s41598-017-12464-7
- Hradsky BA, Kelly LT, Robley A, Wintle BA (2019) FoxNet: an individual-based model framework to support management of an invasive predator, the red fox. *Journal of Applied Ecology* **56**, 1460–1470. doi:10.1111/1365-2664.13374
- Hunter DO, Lagisz M, Leo V, Nakagawa S, Letnic M (2018) Not all predators are equal: a continent-scale analysis of the effects of predator control on Australian mammals. *Mammal Review* **48**, 108–122. doi:10.1111/mam.12115
- Immell D, Anthony RG (2008) Estimation of black bear abundance using a discrete DNA sampling device. *Journal of Wildlife Management* **72**, 324–330. doi:10.2193/2006-297
- Keem JL, Hradsky B, Benshemesh J, Le Pla M, Watkins A, Weeks AR, van Rooyen A, Black J, Southwell D (2022) Data from: Evaluating predator control using two non-invasive population metrics: a camera trap activity index and density estimation from scat genotyping. [Dataset]. Zenodo. Available at <https://doi.org/10.5281/zenodo.5883417>
- Kolowski JM, Oley J, McShea WJ (2021) High-density camera trap grid reveals lack of consistency in detection and capture rates across space and time. *Ecosphere* **12**, e03350. doi:10.1002/ecs2.3350
- Lazenby BT, Mooney NJ, Dickman CR (2014) Effects of low-level culling of feral cats in open populations: a case study from the forests of southern Tasmania. *Wildlife Research* **41**, 407–420. doi:10.1071/WR14030
- Le Pla MN, Birnbaum EK, Rees MW, Hradsky BA, Weeks AR, Van Rooyen A, Pascoe JH (2022) Genetic sampling and an activity index indicate contrasting outcomes of lethal control for an invasive predator. *Austral Ecology* **47**, 1062–1076. doi:10.1111/aec.13182
- Lonsinger RC, Lukacs PM, Gese EM, Knight RN, Waits LP (2018) Estimating densities for sympatric kit foxes (*Vulpes macrotis*) and coyotes (*Canis latrans*) using noninvasive genetic sampling. *Canadian Journal of Zoology* **96**, 1080–1089. doi:10.1139/cjz-2017-0332
- Lowe S, Browne M, Boudjelas S, Poorter M (2000) '100 of the world's worst invasive alien species: a selection from the global invasive species database.' (Invasive Species Specialist Group: Auckland, New Zealand) doi:10.1525/9780520948433-159
- Magnusson A, Skaug H, Nielsen A, Berg C, Kristensen K, Maechler M, Bentham K van, Bolker B, Lüdecke D, Lenth R, O'Brien J, Geyer CJ, McGillicuddy M, Brooks M (2021) glmmTMB: generalized linear mixed models using template model builder. Available at <https://CRAN.R-project.org/package=glmmTMB> [accessed 22 August 2021]
- Mann GKH, O'Riain MJ, Parker DM (2015) The road less travelled: assessing variation in mammal detection probabilities with camera traps in a semi-arid biodiversity hotspot. *Biodiversity and Conservation* **24**, 531–545. doi:10.1007/s10531-014-0834-z
- Manning AD, Andrewartha TA, Blencowe A, Brewer K, Gordon IJ, Evans MJ (2021) Bettering the devil you know: can we drive predator adaptation to restore native fauna? *Conservation Science and Practice* **3**, e447. doi:10.1111/csp.2.447
- Marks CA, Bloomfield TE (1999) Distribution and density estimates for urban foxes (*Vulpes vulpes*) in Melbourne: implications for rabies control. *Wildlife Research* **26**, 763–775. doi:10.1071/WR98059
- Marks CA, Gigliotti F, Busana F (2002) Estimated 1080 dose rate for the M-44 ejector for the control of red foxes (*Vulpes vulpes*). *Wildlife Research* **29**, 291–294. doi:10.1071/WR00115
- Marks CA, Gigliotti F, McPhee S, Piggott MP, Taylor A, Glen AS (2009) DNA genotypes reveal red fox (*Vulpes vulpes*) abundance, response to lethal control and limitations of contemporary survey techniques. *Wildlife Research* **36**, 647–658. doi:10.1071/WR08109
- Marlow NJ (1992) Ecology of the introduced red fox (*Vulpes vulpes*) in the arid zone. PhD Thesis, University of New South Wales.
- Marlow NJ, Thomson PC, Algar D, Rose K, Kok NE, Sinagra JA (2000) Demographic characteristics and social organisation of a population of red foxes in a rangeland area in Western Australia. *Wildlife Research* **27**, 457–464. doi:10.1071/WR99035
- McLeod R (2004) 'Counting the cost: impact of invasive animals in Australia.' (Pest Animal Control Co-operative Research Centre: Canberra, Australia)
- Meek PD, Ballard G, Claridge A, Kays R, Moseby K, O'Brien T, O'Connell A, Sanderson J, Swann DE, Tobler M, Townsend S (2014) Recommended guiding principles for reporting on camera trapping research. *Biodiversity and Conservation* **23**, 2321–2343. doi:10.1007/s10531-014-0712-8
- Meek PD, Ballard G-A, Fleming PJS, Meek PD, Ballard G-A, Fleming PJS (2015) The pitfalls of wildlife camera trapping as a survey tool in Australia. *Australian Mammalogy* **37**, 13–22. doi:10.1071/AM14023
- Morin PA, McCarthy M (2007) Highly accurate SNP genotyping from historical and low-quality samples. *Molecular Ecology Notes* **7**, 937–946. doi:10.1111/j.1471-8286.2007.01804.x
- Morin DJ, Waits LP, McNitt DC, Kelly MJ (2018) Efficient single-survey estimation of carnivore density using fecal DNA and spatial capture-recapture: a bobcat case study. *Population Ecology* **60**, 197–209. doi:10.1007/s10144-018-0606-9
- Moruzzi TL, Fuller TK, DeGraaf RM, Brooks RT, Li W (2002) Assessing remotely triggered cameras for surveying carnivore distribution. *Wildlife Society Bulletin (1973-2006)* **30**, 380–386.
- Moseby K, Nano T, Southgate R (2012) 'Tales in the sand: a guide to identifying Australian arid zone fauna using spoor and other signs.' 3rd edn. (Ecological Horizons: South Australia)
- Murphy MA, Kendall KC, Robinson A, Waits LP (2007) The impact of time and field conditions on brown bear (*Ursus arctos*) faecal DNA amplification. *Conservation Genetics* **8**, 1219–1224. doi:10.1007/s10592-006-9264-0
- Nakagawa S, Schielzeth H (2013) A general and simple method for obtaining R^2 from generalized linear mixed-effects models. *Methods in Ecology and Evolution* **4**, 133–142. doi:10.1111/j.2041-210x.2012.00261.x
- Nakashima Y, Fukasawa K, Samejima H (2018) Estimating animal density without individual recognition using information derivable exclusively from camera traps. *Journal of Applied Ecology* **55**, 735–744. doi:10.1111/1365-2664.13059
- Newey S, Davidson P, Nazir S, Fairhurst G, Verdicchio F, Irvine RJ, van der Wal R (2015) Limitations of recreational camera traps for wildlife

- management and conservation research: a practitioner's perspective. *Ambio* **44**, 624–635. doi:10.1007/s13280-015-0713-1
- Niedballa J, Sollmann R, Courtiol A, Wilting A (2016) camtrapR: an R package for efficient camera trap data management. *Methods in Ecology and Evolution* **7**, 1457–1462. doi:10.1111/2041-210X.12600
- Norbury G (2001) Conserving dryland lizards by reducing predator-mediated apparent competition and direct competition with introduced rabbits. *Journal of Applied Ecology* **38**, 1350–1361. doi:10.1046/j.0021-8901.2001.00685.x
- Palmer MS, Swanson A, Kosmala M, Arnold T, Packer C (2018) Evaluating relative abundance indices for terrestrial herbivores from large-scale camera trap surveys. *African Journal of Ecology* **56**, 791–803. doi:10.1111/aje.12566
- Peakall R, Smouse PE (2012) GenAlEx 6.5: genetic analysis in Excel. Population genetic software for teaching and research – an update. *Bioinformatics* **28**, 2537–2539. doi:10.1093/bioinformatics/bts460
- Pech RP, Sinclair ARE, Newsome AE, Catling PC (1992) Limits to predator regulation of rabbits in Australia: evidence from predator-removal experiments. *Oecologia* **89**, 102–112. doi:10.1007/BF00319021
- Piggott MP (2004) Effect of sample age and season of collection on the reliability of microsatellite genotyping of faecal DNA. *Wildlife Research* **31**, 485–493. doi:10.1071/WR03096
- Piggott MP, Taylor AC (2003) Extensive evaluation of faecal preservation and DNA extraction methods in Australian native and introduced species. *Australian Journal of Zoology* **51**, 341–355. doi:10.1071/ZO03012
- Piggott MP, Wilson R, Banks SC, Marks CA, Gigliotti F, Taylor AC (2008) Evaluating exotic predator control programs using non-invasive genetic tagging. *Wildlife Research* **35**, 617–624. doi:10.1071/WR08040
- Proffitt KM, Garrott R, Gude JA, Hebblewhite M, Jimenez B, Paterson JT, Rotella J (2020) Integrated carnivore-ungulate management: a case study in West-Central Montana. *Wildlife Monographs* **206**, 1–28. doi:10.1002/wmon.1056
- Ramsey DSL, Caley PA, Robley A (2015) Estimating population density from presence-absence data using a spatially explicit model: estimating density from presence-absence data. *The Journal of Wildlife Management* **79**, 491–499. doi:10.1002/jwmg.851
- R Core Team (2022) 'R: a language and environment for statistical computing.' R Core Team.
- Reddiex B, Forsyth DM, McDonald-Madden E, Einoder LD, Griffioen PA, Chick RR, Robley AJ (2006) Control of pest mammals for biodiversity protection in Australia. I. Patterns of control and monitoring. *Wildlife Research* **33**, 691–709. doi:10.1071/WR05102
- Robinson OJ, Fefferman NH, Lockwood JL (2013) How to effectively manage invasive predators to protect their native prey. *Biological Conservation* **165**, 146–153. doi:10.1016/j.biocon.2013.06.001
- Rodgers TW, Giacalone J, Heske EJ, Janečka JE, Phillips CA, Schooley RL (2014) Comparison of noninvasive genetics and camera trapping for estimating population density of ocelots (*Leopardus pardalis*) on Barro Colorado Island, Panama. *Tropical Conservation Science* **7**, 690–705. doi:10.1177/194008291400700408
- Rowcliffe JM, Field J, Turvey ST, Carbone C (2008) Estimating animal density using camera traps without the need for individual recognition. *Journal of Applied Ecology* **45**, 1228–1236. doi:10.1111/j.1365-2664.2008.01473.x
- Royle JA, Young KV (2008) A hierarchical model for spatial capture–recapture data. *Ecology* **89**, 2281–2289. doi:10.1890/07-0601.1
- Ruell EW, Riley SPD, Douglas MR, Pollinger JP, Crooks KR (2009) Estimating bobcat population sizes and densities in a fragmented urban landscape using noninvasive capture–recapture sampling. *Journal of Mammalogy* **90**, 129–135. doi:10.1644/07-MAMM-A-249.1
- Sadler LMJ, Webbon CC, Baker PJ, Harris S (2004) Methods of monitoring red foxes *Vulpes vulpes* and badgers *Meles meles*: are field signs the answer? *Mammal Review* **34**, 75–98. doi:10.1046/j.0305-1838.2003.00029.x
- Šálek M, Kreisinger J, Sedláček F, Albrecht T (2010) Do prey densities determine preferences of mammalian predators for habitat edges in an agricultural landscape? *Landscape and Urban Planning* **98**, 86–91. doi:10.1016/j.landurbplan.2010.07.013
- Saunders G, McLeod L (2007) 'Improving fox management strategies in Australia.' (Bureau of Rural Sciences: Canberra, Australia)
- Saunders GR, Gentle MN, Dickman CR (2010) The impacts and management of foxes *Vulpes vulpes* in Australia: impact and management of foxes in Australia. *Mammal Review* **40**, 181–211. doi:10.1111/j.1365-2907.2010.00159.x
- Schaus J, Uzal A, Gentle LK, Baker PJ, Bearman-Brown L, Bullion S, Gazzard A, Lockwood H, North A, Reader T, Scott DM, Sutherland CS, Yarnell RW (2020) Application of the Random Encounter Model in citizen science projects to monitor animal densities. *Remote Sensing in Ecology and Conservation* **6**, 514–528. doi:10.1002/rse2.153
- Siddig AA, Ellison AM, Jackson S (2015) Calibrating abundance indices with population size estimators of red back salamanders (*Plethodon cinereus*) in a New England forest. *PeerJ* **3**, e952. doi:10.7717/peerj.952
- Sollmann R, Mohamed A, Samejima H, Wilting A (2013) Risky business or simple solution – relative abundance indices from camera-trapping. *Biological Conservation* **159**, 405–412. doi:10.1016/j.biocon.2012.12.025
- Soulsbury CD, Iossa G, Baker PJ, White PCL, Harris S (2011) Behavioral and spatial analysis of extraterritorial movements in red foxes (*Vulpes vulpes*). *Journal of Mammalogy* **92**, 190–199. doi:10.1644/09-MAMM-A-187.1
- Southwell D, McCowen S, Mewett O, Hennecke B (2011) Understanding the drivers and barriers towards adoption of innovative canid control technologies: a review. Australian Bureau of Agricultural and Resource Economics and Sciences: report prepared for the Invasive Animals Cooperative Research Centre, Canberra.
- Spencer EE, Newsome TM, Dickman CR (2017) Prey selection and dietary flexibility of three species of mammalian predator during an irruption of non-cyclic prey. *Royal Society Open Science* **4**, 170317. doi:10.1098/rsos.170317
- Stephens PA, Pettorelli N, Barlow J, Whittingham MJ, Cadotte MW (2015) Management by proxy? The use of indices in applied ecology. *Journal of Applied Ecology* **52**, 1–6. doi:10.1111/1365-2664.12383
- Thompson JA, Fleming PJS (1994) Evaluation of the efficacy of 1080 poisoning of red foxes using visitation to non-toxic baits as an index of fox abundance. *Wildlife Research* **21**, 27–39. doi:10.1071/WR9940027
- Thompson GG, Thompson SA, Bengsen A (2019) The value of camera traps in monitoring a feral-cat and fox reduction program. *Wildlife Research* **46**, 599–609. doi:10.1071/WR18087
- Towerton AL, Penman TD, Kavanagh RP, Dickman CR (2011) Detecting pest and prey responses to fox control across the landscape using remote cameras. *Wildlife Research* **38**, 208–220. doi:10.1071/WR10213
- Towerton AL, Dickman CR, Kavanagh RP, Penman TD (2016) Control of the red fox in remnant forest habitats. *Wildlife Research* **43**, 169–177. doi:10.1071/WR15133
- Trewhella WJ, Harris S, McAllister FE (1988) Dispersal distance, home-range size and population density in the red fox (*Vulpes vulpes*): a quantitative analysis. *Journal of Applied Ecology* **25**, 423–434. doi:10.2307/2403834
- Triggs B (2004) 'Tracks, scats and other traces: a field guide to Australian mammals.' (Oxford University Press: South Melbourne, Australia)
- van Bommel L, Johnson CN (2012) Good dog! Using livestock guardian dogs to protect livestock from predators in Australia's extensive grazing systems. *Wildlife Research* **39**, 220–229. doi:10.1071/WR11135
- van Hesper R (2015) Designing a camera trap monitoring program to detect changes in fox abundance. Masters Thesis, School of BioSciences: The University of Melbourne.
- van Hesper R, Hauser CE, Benshemesh J, Rumpff L, Monfort JJJ (2019) Designing a camera trap monitoring program to measure efficacy of invasive predator management. *Wildlife Research* **46**, 154–164. doi:10.1071/WR17139
- van Polanen Petel AM, Marks CA, Morgan DG (2001) Bait palatability influences the caching behaviour of the red fox (*Vulpes vulpes*). *Wildlife Research* **28**, 395–401. doi:10.1071/WR00046
- Victorian Government Department of Sustainability and Environment (2005a) Ecological vegetation classes: Wimmera bioregion. The State of Victoria Department of Sustainability and Environment. Available at https://www.environment.vic.gov.au/_data/assets/pdf_file/0033/48759/Wim_EVCs_combined.pdf
- Victorian Government Department of Sustainability and Environment (2005b) Ecological vegetation classes: Murray Mallee bioregion. The State of Victoria Department of Sustainability and Environment. Available at https://www.environment.vic.gov.au/_data/assets/pdf_file/0026/48734/MuM_EVCs_combined.pdf

- Vine SJ, Crowther MS, Lapidge SJ, Dickman CR, Mooney N, Piggott MP, English AW (2009) Comparison of methods to detect rare and cryptic species: a case study using the red fox (*Vulpes vulpes*). *Wildlife Research* **36**, 436–446. doi:10.1071/WR08069
- Waits LP, Luikart G, Taberlet P (2001) Estimating the probability of identity among genotypes in natural populations: cautions and guidelines. *Molecular Ecology* **10**, 249–256. doi:10.1046/j.1365-294X.2001.01185.x
- Walsh JC, Wilson KA, Benshemesh J, Possingham HP (2012) Unexpected outcomes of invasive predator control: the importance of evaluating conservation management actions. *Animal Conservation* **15**, 319–328. doi:10.1111/j.1469-1795.2012.00537.x
- Wandeler P, Funk SM (2006) Short microsatellite DNA markers for the red fox (*Vulpes vulpes*). *Molecular Ecology Notes* **6**, 98–100. doi:10.1111/j.1471-8286.2005.01152.x
- Wearn OR, Glover-Kapfer P (2019) Snap happy: camera traps are an effective sampling tool when compared with alternative methods. *Royal Society Open Science* **6**, 181748. doi:10.1098/rsos.181748
- Wegge P, Bakke BB, Odden M, Rolstad J (2019) DNA from scats combined with capture–recapture modeling: a promising tool for estimating the density of red foxes – a pilot study in a boreal forest in southeast Norway. *Mammal Research* **64**, 147–154. doi:10.1007/s13364-018-0408-7
- Williams BK, Nichols JD, Conroy MJ (2002) Analysis and management of animal populations: modeling, estimation and decision making. Available at <https://pubs.er.usgs.gov/publication/5200256> [accessed 24 January 2022]
- Wilson DJ, Mulvey RL, Clarke DA, Reardon JT (2017) Assessing and comparing population densities and indices of skinks under three predator management regimes. *New Zealand Journal of Ecology* **41**, 84–97. doi:10.20417/nzjecol.41.7
- Woinarski JCZ, Burbidge AA, Harrison PL (2015) Ongoing unraveling of a continental fauna: decline and extinction of Australian mammals since European settlement. *Proceedings of the National Academy of Sciences of the United States of America* **112**, 4531–4540. doi:10.1073/pnas.1417301112
- Wyatt KB, Campos PF, Gilbert MTP, Kolokotronis S-O, Hynes WH, DeSalle R, Daszak P, MacPhee RDE, Greenwood AD (2008) Historical mammal extinction on Christmas Island (Indian Ocean) correlates with introduced infectious disease. *PLoS ONE* **3**, e3602. doi:10.1371/journal.pone.0003602

Data availability. Analyses reported in this study can be reproduced using the data available via Zenodo: <https://doi.org/10.5281/zenodo.5883417> (Keem et al. 2022).

Conflicts of interest. The authors declare that they have no conflicts of interest.

Declaration of funding. This study was funded by the State of Victoria through the Department of Environment, Land, Water and Planning TP829577: *Optimising non-invasive genetic sampling strategies to evaluate predator control efficacy and impacts on native fauna*.

Acknowledgements. We pay our respects to the Traditional Owners of the land on which research was conducted: Wotjobaluk, Jaadwa, Jadawadjali, Wergaia and Jupagulk Nations of the Wimmera region, and Kureinji, Ngintait, Nyeri Nyeri, Latji Latji, Tatti Tatti, Wadi Wadi and Maraura Nations of the Mallee land and waterways. This project was approved by the Science Animal Ethics Committee at the University of Melbourne (project ID 1914959.1). The Department of Environment, Land, Water and Planning (DELWP) granted a wildlife permit (no. 10009227) for this study, which adhered to the Wildlife Act 1975 and the National Parks Act 1975. We thank Parks Victoria and the Mallee Catchment Management Authority, especially Derrick Boord. Thank you to the National Malleefowl Recovery Team and the Victorian Malleefowl Recovery Group for ongoing support, in particular Iestyn Hosking, Mirinda Thorpe, Ronni O'Donnell, Tom O'Donnell, Stuart Thorpe and Kerrie Thorpe. Thank you to fieldwork volunteers Natalie Bates, Stefanie Jones, Laura Kors, Stella Johnston, Hannah Glasson, Annabel Cadzon, Kathy Keem, Hayden Keem, Erin Thomas and Eliza Thompson. We thank Matthew Rees, Brendan Wintle, José Lahoz-Monfort and two anonymous reviewers for useful comments and research guidance.

Author affiliations

^AQuantitative and Applied Ecology Group, School of Ecosystem and Forest Sciences, University of Melbourne, Parkville, VIC 3010, Australia.

^BNational Malleefowl Recovery Group, VIC, Australia.

^CConservation Ecology Centre, Cape Otway, VIC 3233, Australia.

^DParks Victoria Wimmera, Wail, VIC 3414, Australia.

^EBio21 Institute, School of Biosciences, University of Melbourne, Parkville, VIC 3010, Australia.

^FCesar Australia, Brunswick, VIC 3056, Australia.

^GConservation Science Research Group, School of Environmental and Life Science, University of Newcastle, Callaghan, NSW 2308, Australia.