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Supplementary Material

What can echolocation recordings reveal about the foraging ecology of

Saccolaimus saccolaimus (Emballonuridae) in north-western Australia?

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Appendix 1. The relationship between Q_{SM2} to Q_{Anabat} was derived empirically for each species. The 11 species illustrated are ordered according to average Q_{SM2} (the fineness-of-tuning of their search-mode pulses 6 dB below peak), with 95% confidence intervals shown. Solid boxes are values measured from Anabat2 recordings (see Methods); hollow boxes are values measured from Wildlife Acoustics SM2BAT recordings at a sampling rate of 384 kHz. Species codes are provided in Appendix 2. Sample-sizes, and sample-averages are tabulated below.



	Na	Ма	Aa	Cj	Sf	Sc	Sg	Тд	Ос	Vc	Pw
SM2											
avF _{peak} (kHz)	51.8	45.3	11.9	17.7	18.6	19.7	37.9	24.7	30.8	61.4	48.6
avQ _{SM2}	7.7	9.3	19.7	24.6	30.3	37.1	39.1	49.7	55.0	85.3	89.5
s.d.	2.0	2.0	3.1	4.5	5.0	3.2	13.1	7.0	9.2	22.1	14.3
#pulses	73	64	108	147	81	64	65	90	124	99	80
#sequences	15	11	17	19	20	14	16	23	14	20	15
#localities	9	6	7	11	14	10	8	12	6	8	8

Anabat											
avF _{peakC} (kHz)	50.7	43.4	12.0	18.4	18.1	19.6	38.2	24.7	32.0	61.0	49.0
avQ _{Anabat}	2.3	2.5	6.7	5.4	10.3	12.5	10.9	15.3	11.8	16.1	18.1
s.d.	0.7	0.6	2.3	1.7	2.6	1.7	2.7	2.8	3.3	3.5	3.8
#pulses	136	44	38	190	161	66	281	225	275	95	94
#sequences	11	16	17	27	20	12	59	41	43	30	6
#localities	6	6	13	18	16	10	40	39	5	28	5
Q SM2 /Q Anabat	3.4	3.7	2.9	4.6	2.9	3.0	3.6	3.2	4.7	5.3	4.9

Appendix 2. Species foraging strategy, foraging microhabitat, climatic range, phylogeny, muscle mass class and flight speed.

Population (Western Australian bioregion) from which study animals were recorded: K (Kimberley bioregions), P (Pilbara), C (Coolgardie), M (Murchison), SW (temperate forest regions). Usual foraging strategy categories: A_H = air superiority involving highly agile turns, A_o = air superiority in open situations, I = interceptor, S_{2D} = two dimensional surface gleaning, S_{3D} = 3D-surface gleaning (includes 'P', trawling for tiny fish and aquatic invertebrates). Usual foraging microhabitat categories: OC = over canopy, AC = above canopy, BS/O = beside stand in open, BS/A beside stand, against clutter, IS = inside stand. Geographical range abbreviations: Tr = tropical, Te = temperate, a =arid, m = mesic. Family name abbreviations: E = Emballonuridae, M = Molossidae, Mi = Miniopteridae, V = Vespertilionidae (includes N = Nyctophilinae). Flight muscle mass classes: LE = low energy, GE = general energy and HE = high energy (Bullen and McKenzie 2004). V_{ae} = maximum aerobic level-flight speed (Bullen *et al.* 2016), av F_{peak} = peak frequency of pulses comprising search mode echolocation sequences; av f_w = average wing-beat frequency; 'av #seq/locn' = average (s.d., N) number of sequences analysed per location. The average f_w and V_{mode} values for molossids excludes flight with un-cambered wings (Bullen *et al.* 2014), and for Nyctophilinae exclude 'departure' calls (McKenzie and Bullen 2012).

Code			Usual	Usual			Flight				
	Species	Population	Foraging	Foraging	Range	Family	muscle	Vae	avF _{peak} (sd, n)	avƒw (sd, n)	av#seq/locn
			Strategy	Microhabitat			mass				(sd, N)
							class				
Cj	Chaerephon jobensis (Miller)	к	2	OC-AC ²	Tr, a, m	М	LE	7.4	18.0 (1.9, 44)	8.4 (0.4, 44)	2.5 (1.4, 18)

Cj	Chaerephon jobensis (Miller)	Р	¹	OC-AC ¹	Tr, a, m	Μ	LE	8.4	17.7 (1.2, 34)	8.3 (1.2, 34)	1.6 (0.9, 16)
Cg	Chalinolobus gouldii (Grey)	К	A _H ²	BS/O ²	Tr, m	v	HE	6.9	31.5 (1.3, 109)	9.3 (0.8, 109)	2.3 (1.4, 7)
Cg	Chalinolobus gouldii (Grey)	Р	A _H ¹	BS/O ¹	Tr, a	v	HE	6.5	32.1 (1.3, 128)	9.4 (1.3, 128)	2.4 (1.6, 22)
Ст	Chalinolobus morio (Grey)	Р	A _H ¹³	BS/A 13	Te, Tr, a, m	v	HE	5.7	51 (0.5, 35)	10.6 (1.6, 35)	6 (, 1)
Cn	Chalinolobus nigrogriseus (Gould)	К	A _H ²	BS/O ²	Tf, m	v	HE	5.6	39.5 (1.8, 74)	10.7 (0.5, 74)	2.4 (1.4, 5)
Fm	Falsistrellus mackenziei Kitchener	SW	A 4	BS/O ⁴	Te, m	v	GE	7.6	35.1 (1.5, 43)	8.1 (0.4, 43)	1.3 (0.5, 7)
Ms	Miniopterus schreibersii orianae Thomas	К	A _H ²	BS/A-BS/O ²	Tr, m	Mi	HE	5.8	49.4 (1.2, 163)	9.5 (1, 163)	5.4 (6.7, 5)
Ма	Myotis macropus (Gould)	К	P (A-S _{3D} ²)	BS/O ²	Tr, m	v	GE	6.3	41.5 (2.6, 123)	9.5 (1.3, 123)	3.2 (4.4, 5)
Na	Nyctophilus arnhemensis Johnson	К	S_{3D} 2	BS/A-IS ²	Tr, m	Ν	GE	5.3	50.3 (3.7, 89)	10.4 (1.3, 89)	2.2 (1.8, 5)
Nd	Nyctophilus daedalus Thomas	K&P	S _{3D} ¹²	BS/A-IS ¹²	Tr, a, m	Ν	GE	5.7	54.7 (3.4, 23)	9.8 (1.1, 23)	2.4 (0.6, 4)
Ng	Nyctophilus geoffroyi pallescens (Thomas)	К	S_{3D} 2	BS/A-IS ²	Tr, a, m (mangal)	Ν	GE	5	47.9 (1.3, 65)	10.4 (1.3, 65)	2.4 (1.6, 3)
Ng	Nyctophilus geoffroyi pallescens (Thomas)	Ρ	S_{3D} ¹	BS/A-IS ¹	Tr, a, m (mangal)	Ν	GE	5.3	47.3 (3.1, 114)	10.8 (1, 114)	2.3 (2.1, 14)
Nw	Nyctophilus walkeri Thomas	К	S _{3D} -A ²	BS/A ²	Tr, m	Ν	HE	5.3	60.8 (3.6, 85)	11.2 (0.8, 85)	1.6 (1.4, 5)
Oc	Ozimops cobourgianus (Johnson) (= Mormopterus	<i>K 9</i> . D	Ao ¹	OC-BS/A ¹	Tr, a, m (mangal)	М	16	6.7	32.1 (1.8, 386)	10.1 (0.7, 177)	6.3 (7.3, 10)
00	loriae cobourgiana)	KQF									
01	Ozimops lumsdenae Reardon et al. (= Mormopterus	V	1.2	OC-BS/O ²	Tr, m	М	16	7.6	27.6 (1.3, 84)	9.4 (0.9, 84)	6 (, 1)
01	beccarii)	ĸ	·				LL				
01	Ozimops lumsdenae Reardon et al. (= Mormopterus	D	11		Tra	М	IF	76	27 2 (1 9 22)	96(0933)	16(0715)
01	beccarii)	ļ		00-03/0	11, a	IVI	LL	7.0	27.3 (1.5, 55)	5.0 (0.5, 55)	1.0 (0.7, 15)
Pw	Pipistrellus westralis Koopman	К	A _H ²	BS/A ²	Tr, m	v	GE	5.8	49.1 (1.6, 89)	11.2 (1.1, 89)	4.0 (5&3, 2)
Sf	Saccolaimus flaviventris (Peters)	Р	A ₀ ¹²	OC 12	Tr, a, m	E	GE	8	17.1 (1.4, 72)	7.9 (0.8, 72)	1.9 (1, 21)
Sg	Scotorepens greyii (Grey)	К	A _H ²	BS/O ²	Tr, a, m	v	HE	6.1	39.7 (1.4, 166)	10.6 (0.5, 166)	1.9 (1.9, 8)
Sg	Scotorepens greyii (Grey)	Р	A _H ¹	BS/O ¹	Tr, a, m	V	HE	6.4	37.5 (1.5, 111)	10.6 (0.9, 111)	2 (1.6, 23)

Ss	Scotorepens sanborni (Troughton)	К	A _H ²	BS/A-BS/O ²	Tr, m	V	HE	6.1	41.1 (2.0, 99)	10.8 (0.5, 99)	5.4 (2.6, 3)
Та	Austronomus australis (Gray) (= Tadarida australis)	COOL	¹³	OC ¹³	Tr, Te, a, m	М	GE	8.7	12.5 (1.4, 39)	7.7 (0.7, 39)	1.6 (1.1, 13)
Тg	Taphozous georgianus Thomas	К	Ao ¹²	OC 12	Tr, m	E	GE	7.1	24.7 (0.6, 98)	8.2 (0.3, 98)	1.8 (1.2, 8)
Тg	Taphozous georgianus Thomas	Ρ	Ao ¹²	OC 12	Tr, a	E	GE	7.5	24.6 (1.0, 69)	8.5 (1.1, 69)	2 (1.5, 18)
Th	Taphozous hilli Kitchener	MUR	A ₀ ¹	OC ¹	Tr, Te, a	E	GE	7.1	25.7 (0.8, 140)	8.2 (1, 140)	9 (6.6, 4)
Vc	Vespadelus caurinus (Thomas)	К	A _H ²	BS/A ²	Tr, m	V	HE	4.2	61 (1.6, 40)	11.8 (0.6, 40)	1.5 (0.8, 8)
Vd	Vespadelus douglasorum (Kitchener)	К	A _H ²	BS/A ²	Tr, m	V	GE	5.8	51.1 (1.4, 53)	10.4 (0.6, 53)	1.8 (1.5, 4)
Vf	<i>Vespadelus finlaysoni</i> Kitchener <i>et al.</i>	Р	A _H ¹	BS/A ¹	Tr, Te, a	V	HE	4.6	56.3 (1.6, 74)	11.4 (1.5, 74)	2.4 (2.8, 19)
Sf K	Saccolaimus flaviventris (Peters)	К			Tr, a, m	E			18.3 (1.6, 46)	7.2 (0.4, 46)	2.0 (2.2, 23)
Sc K&NT	Saccolaimus saccolaimus Temminck	K&NT			Tr, m	E			20.3 (0.8, 51)	7.3 (0.3, 51)	2.0 (1.8, 25)

¹ McKenzie and Bullen (2009); ² McKenzie and Bullen (2012); ³ Bullen and McKenzie (2001); ⁴ Van Dyck and Strahan (2008).

References

- <jrn>Bullen, R. D., and McKenzie, N. L. (2001). Bat airframe design flight performance, stability and control in relation to foraging ecology. *Australian Journal of Zoology* 49, 235–261. doi:10.1071/Z000037
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 <jrn>Bullen, R. D., and McKenzie, N. L. (2004). Bat flight-muscle mass: implications for foraging strategy. Australian Journal of Zoology 52, 605–622.
 doi:10.1071/Z004036
- <jrn>Bullen, R. D., McKenzie, N. L., and Cruz-Neto, A. P. (2014). Aerodynamic power and mechanical efficiency of bat airframes using a quasi-steady model. *CEAS Aeronautical Journal* 5, 253–264. doi:10.1007/s13272-014-0104-5
- <jrn>Bullen, R. D., McKenzie, N. L., and Cruz-Neto, A. P. (2016). Characteristic flight speeds in bats. CEAS Aeronautical Journal 7, 621–643. doi:10.1007/s13272-016-0212-5
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- <jrn>McKenzie, N. L., and Bullen, R. D. (2012). An acoustic survey of zoophagic bats on islands in the Kimberley, Western Australia, including data on the echolocation ecology, organisation and habitat relationships of regional communities. *Records of the Western Australian Museum* 81(Supplement), 67–108. doi:10.18195/issn.0313-122x.81.2012.067-108
-
bok>Van Dyck, S., and Strahan, R. (2008). 'Mammals of Australia.' (New Holland Publishers: Australia.)</bok>