

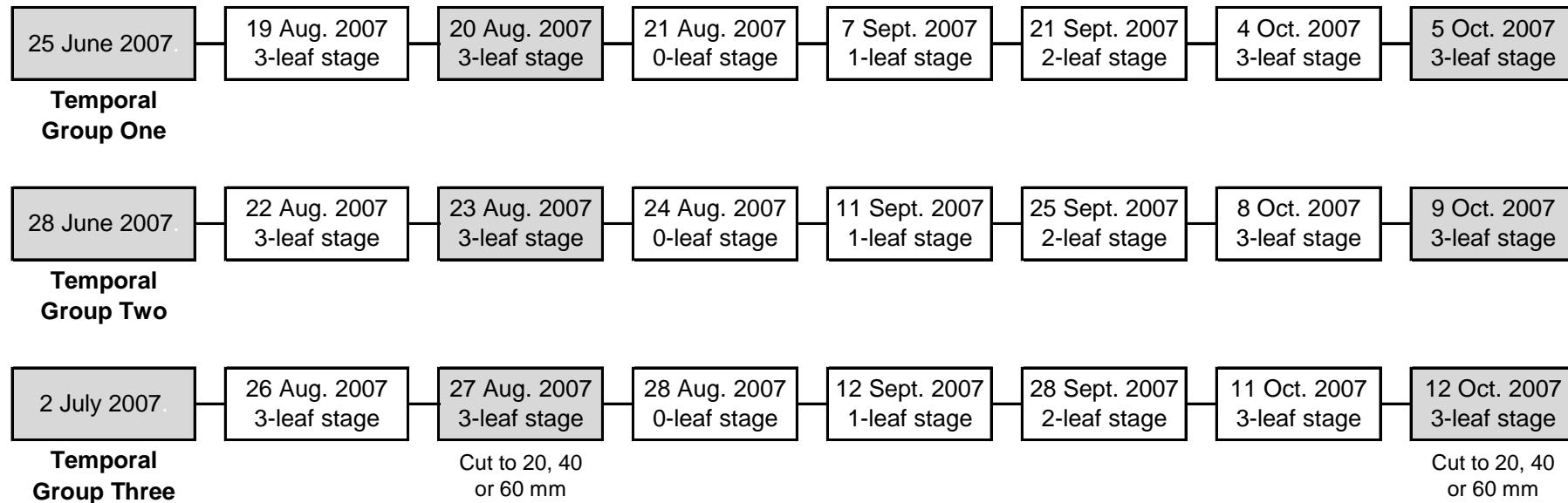
Accessory Publication

Fig. S1. Diagrammatical representation of defoliation events. Perennial ryegrass-dominant plots were defoliated to 20, 40 or 60 mm residual height (grey boxes) and samples of perennial ryegrass lamina and stubble tissue were harvested from plants for reverse transcription quantitative polymerase chain reaction (RT-qPCR) analyses (white boxes). The lamina and stubble samples for RT-qPCR collected at the 3-leaf stage on the 19, 22 and 26 August were used as calibrator/control samples in the LightCycler software.

Table S1. Meteorological data during the current study and averages from the past 30 years

	Current study					Monthly averages (1976–2006)				
	Jun.	Jul.	Aug.	Sept.	Oct.	Jun.	Jul.	Aug.	Sept.	Oct.
Maximum air temperature (°C)	14.6	14.7	15.1	17.0	17.6	14.3	13.7	14.6	16.2	17.9
Minimum air temperature (°C)	4.7	5.7	6.1	6.4	8.2	5.0	4.0	4.8	6.6	8.4
Radiation (MJ m ⁻²)	6.5	6.3	9.3	12.9	17.5	6.1	6.8	9.2	12.8	16.2
Total rainfall (mm)	94	174	113	59	95	114	124	106	93	95

Table S2. PCR primer sequences, amplicon sizes and amplification efficiency

bp, base pairs; 3'UTR, three prime untranslated region; *rbcS1* and *rbcS2*, ribulose 1,5-bisphosphate carboxylase/oxygenase (Rubisco) small subunits; *rca*, Rubisco activase; *lhcb1* and *lhcb2*, Light harvesting chlorophyll *a/b* binding proteins; *fba*, fructose bisphosphate aldolase; *fbp*, fructose 1,6-bisphosphatase; *fnr*, ferredoxin-nicotinamide adenine dinucleotide phosphate oxidoreductase; *ugp*, uridine diphosphate glucose pyrophosphorylase; *sps*, sucrose phosphate synthase; *sus*, sucrose synthase; *ninv*, neutral/alkaline invertase; *hxk*, hexokinase; *hxt*, hexose transporter; *sut*, sucrose transporter; *I-sst*, sucrose:sucrose 1-fructosyltransferase; *feh*, fructan exohydrolase

Gene	GenBank accession no.	Primer sequences (5'→3')	Region primer in	Amplicon size (bp)	Amplicon in 3'UTR (%)	Efficiency
<i>rbcS1</i>	DX922407	(F) ATA TCA CCT GGG TCG AGG AA	3'UTR	134	100	1.904 (±0.013)
		(R) CCA ACG GCG AAT AAA GAA AC				
<i>rbcS2</i>	EC778436	(F) GAG GAG TCC GGC AAG GCA TAA	Coding	74	72	1.950 (±0.039)
		(R) TAT GCT TTT ACA TGT AGC CGG TTC				
<i>rca</i>	GO924784	(F) CCA AGA ACT TCG ACC CAA CTG	Coding	142	58	1.937 (±0.026)
		(R) AAC TTT CAT GCC CAG CCA TC				
<i>lhcb1</i>	GO924759, GO924785,	(F) GCC GAC AAC TTC ATT TCT	3'UTR	129	100	1.933

	GO924789	GA				
		(R) TGA GAA ATA ACC ACA AAC AGC A	3'UTR			(±0.013)
<i>lhcb2</i>	GO924771, GO924800	GO924781, AA	Coding	138	57	1.955
		(R) GTG CAG CTA CAC GTT GAT CC	3'UTR			(±0.005)
<i>fba</i>	GO924786	(F) CAC GTG TCG GGG TAC AAG T	Coding	127	81	1.886
		(R) AGT ACA CCG CAG GAA GGA AG	3'UTR			(±0.008)
<i>fbp</i>	GS923007	(F) TGA TGC AAA TCC ATT TTG AAG	3'UTR	130	100	1.763
		(R) TGC TCC TCT TGG TGT CTT TG	3'UTR			(±0.005)
<i>fnr</i>	GO924778, GO924792,	(F) GCA AAT GTT TCT TCC CAC CT	3'UTR	122	100	1.998
		(R) AAC AGC TTA TCG CGC TCT TT	3'UTR			(±0.073)
<i>ugp</i>	GO924802, GO924803	(F) TCC AGT TCT CCT TCC CTG AG	3'UTR	122	100	1.955

		(R) GAA GTG CAG ATG GAC AGA GG	3'UTR		(±0.035)
<i>sps</i>	GR510765	(F) AGA CGT TAT CGC GCT TGA TT	Coding	110	6 1.907
		(R) ACT GGT GTC ATT TTG TCG GTA	Coding/3'UTR		(±0.036)
<i>sus</i>	GR516033	(F) CAT GAA GAG GTG CCT TCA GA	3'UTR	126	100 1.834
		(R) CCA AAA CCT CAC ACA AGT GC	3'UTR		(±0.049)
<i>ninv</i>	AM489692	(F) TGA GTT CTC GGC ACT GTT AAG	3'UTR	146	100 1.827
		(R) AGG AAT GCT CAA GCA AGT GA	3'UTR		(±0.007)
<i>hxk</i>	GO924754	(F) ACC TTC CTT CGG CTA TTC TG	3'UTR	129	100 1.877
		(R) CCT CCA TCT CCA TTG TTC CT	3'UTR		(±0.033)
<i>hxt</i>	GS923006	(F) TCG CTG GTG GTA ATT TTG TG	3'UTR	134	100 1.953
		(R) CCA GGA TTA CAG CCT CAC	3'UTR		(±0.077)

		G				
<i>sut</i>	GO924755	(F) GCC TCA AGA CTC CGT AGA GC	3'UTR	123	100	1.977
		(R) CAG ACC ACC TAG CCG GTA AT	3'UTR			(±0.028)
<i>feh</i>	DQ016297, DQ073968	(F) CCT CTG ACA ACA TGG AGG AG	Coding	139	0	1.982
		(R) AAG CCT CCA TAK GCT GGT GT	Coding			(±0.004)
<i>I-sst</i>	AB288056	(F) CCA AGT TTA GCT CGT GTT GC	3'UTR	116	100	1.923
		(R) TAT TGT ACG ATC CCC ATC CA	3'UTR			(±0.040)