

Supplementary Material

Hydrogen-rich water-alleviated ultraviolet-B-triggered oxidative damage is partially associated with the manipulation of the metabolism of (iso)flavonoids and antioxidant defence in *Medicago sativa*

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Table S1. The sequences of primers for real-time RT-PCR

Primer name	<i>M. truncatula</i> tentative consensus or accession number	Sequences
<i>PAL</i>	X58180	Forward: CTTGATGAGGTGAAGCGTAT Reverse: ACCGTAAGTGTCCGTGCC
<i>CHS</i>	AW776018	Forward: TGTTTGTGAATACATGGCACCTT Reverse: TGACTTTGGTTGACCCATTCT
<i>CHI</i>	KF765782	Forward: TACTTGAGACCCTTGACTT Reverse: GGTGATTGCCTGTAGAAA
<i>FLS</i>	XM_003601032	Forward: CTTGATGAGGTGAAGCGTAT Reverse: ACCGTAAGTGTCCGTGCC
<i>IFS</i>	AY167424	Forward: AATGGAGAAATCATAGAGGGCGAGCAG Reverse: GTTGATGAGCTCTGCCAAAGTCCATTC
<i>6IOMT</i>	DQ419913	Forward: ACATGGAAAGCCTATGACTGTTC Reverse: ACACAACTCCAGTCCCACCTG
<i>Cu/Zn-SOD</i>	AF056621	Forward: TAATTGCTGATGCCAACG Reverse: ACCACAGGCTAATC TTCCAC
<i>Mn-SOD</i>	AY145894	Forward: TGTCATCAGCG GCGTA ATCAT Reverse: GGGCTTCCTTTGGTGGTTCA
<i>POD 1A</i>	X90692	Forward: TCAATCGTACGTGGTGTGCT Reverse: TGCACTTTGCTCGCTCACTA
<i>POD 1B</i>	X90693	Forward: AGCTGCATTTGCTGCTCAAG Reverse: TTGGTAAGGTTCTGTGCCAGG
<i>POD 1C</i>	X90694	Forward: CCTCGCATGCTTGCTAGTCT Reverse: GGACCTTGTGCCAGAACAGA
<i>POD 2</i>	X90695	Forward: TCCTGCTACCC TTCGTCTCT Reverse: TTCTGCACTGTGGAACAGCA
<i>CAT</i>	TC100988	Forward: TTCTTCTTCTCCACCGTCCTCA Reverse: TCCAAGAGAATTGGACCTCTGG
<i>Actin 2</i>	JQ028730	Forward: AAAAGGATGCCTATGTTGGTG Reverse: AAGTGGAGCCTCAGTTAGAAGTA
<i>GAPDH</i>	GQ398120	Forward: TCATTCCGTGTCCCAACCG Reverse: CCACATCATCTTCAGTGTAACCCA

Table S2. Effects of HRW pretreatment on the relative contents of (iso)flavonoids in the leaves of *Medicago sativa* upon UVB irradiation

14-day-old seedlings were pretreated with or without 50% HRW for 12 h, and then exposed to 0 or 10.8 kJ.m⁻² UVB. After irradiation, seedlings were transferred to the normal growth conditions for another 3 days then sampled. Seedlings without irradiation were set as a control (Con). Relative values of (iso)flavonoids in seedling leaves were determined by UPLC-LTQ-Orbitrap-MS, which were expressed as relative peak areas normalised to the amount of 4-methyumbelliferone added as an internal standard. Data are means ± s.e. from three independent experiments. Different letters within each compound are significantly different at $P < 0.05$ according to Duncan's multiple range test

	No.	Tentative identifications	Relative content in different treatments			
			Con	Con→UVB	HRW→UVB	HRW→Con
Isoflavone	1	Afromosin	0.0065±0.0090c	16.4835±1.9186b	29.9851±2.4090a	0.0056±0.0005c
	2	Afromosin 7-O-β-D-glucoside-malonate	0.0148±0.0016c	4.8729±0.9170b	6.9882±0.7093a	0.0106±0.0017c
	3	Biochanin A	ND	5.8330±0.9532a	3.8790±0.6862b	0.0030±0.0004c
	4	Daidzein	0.0044±0.0006c	5.6354±1.0341b	11.3043±1.5436a	0.0467±0.0063c
	5	Daidzin	0.0012±0.0002c	0.3483±0.0406b	0.4140±0.0674a	0.0020±0.0004c
	6	Formononetin	ND	0.5423±0.0906a	0.2206±0.0887b	0.0028±0.0004c
	7	Formononetin 7-O-β-D-glucoside-6"-O-malonate	0.0303±0.0039c	10.2994±0.9317b	16.0789±2.0679a	0.0455±0.0065c
	8	Genistein	0.4943±0.0707c	0.8920±0.1753a	0.7027±0.0961b	0.4216±0.0623c
	9	Genistein 7-glucoside	ND	0.6855±0.0906b	1.2425±0.1250a	ND
	10	Irisolidone	ND	9.1137±1.4670a	6.5574±1.1056b	ND
	11	Isoformononetin	ND	5.3424±0.9023a	3.5890±0.4944b	0.0024±0.0002c
	12	Prunetin	ND	0.5995±0.0962b	0.7214±0.1052a	ND
Flavone	13	Apigenin	0.8509±0.1071c	3.2371±0.6387b	4.4512±0.6675a	0.5992±0.0661c
	14	Apigenin 7-galacturonide	11.1845±1.807b	16.4379±2.8824a	16.6542±2.8819a	9.1033±1.0966b
	15	Chrysoeriol	0.0008±0.0001c	4.7487±0.7331a	2.0388±0.2914b	ND
	16	Luteolin	ND	0.8046±0.1324a	0.0695±0.0078b	ND
	17	Millettocalyxin A	ND	1.0408±0.1694b	2.1291±0.3123a	0.0014±0.0001c
	18	Milleyanaflavone	ND	5.2941±0.9279b	6.6593±0.9540a	ND
	19	Ptaeroxylol	0.0018±0.0004c	0.7367±0.1019a	0.3010±0.0273b	0.0133±0.0017c
	20	Syzaltein	0.0026±0.0004c	0.8605±0.1089b	1.0263±0.1382a	ND
	21	2',5,6'-Trihydroxy-7-methoxyflavone	0.0008±0.0001c	5.0220±0.6505a	1.8592±0.1502b	ND
	22	3',4'-Dihydroxyflavone	ND	5.8434±0.9469a	3.7740±0.4849b	0.0026±0.004c
Flavanone	23	Betagarin	ND	0.4590±0.1854a	0.4661±0.1882a	ND
	24	Citronetin	ND	0.0888±0.0142a	0.0562±0.0086b	ND
	25	Garbanzol	ND	4.4301±0.3472b	6.9202±0.8852a	ND
	26	Isosakuranetin	0.0011±0.0002c	0.0207±0.0042b	0.0650±0.0101a	ND

	27	Liquiritigenin	ND	1.5731±0.1540a	1.5468±0.1443a	0.0048±0.0011b
	28	Matteucin	ND	3.1936±0.4958b	6.9565±0.5889a	ND
	29	Naringenin	ND	2.5531±0.4687b	4.4638±0.2971a	0.0065±0.0006c
	30	Pinostrobin	ND	2.2765±0.3445a	1.6271±0.2155b	ND
	31	Sakuranetin	0.0016±0.0002c	0.2468±0.0274b	0.4875±0.0660a	ND
	32	4'-Hydroxy-7-methoxy flavanone	0.0122±0.0023c	0.7838±0.0929a	0.5604±0.0632b	0.0216±0.0044c
Flavonol	33	Kaempferol	ND	0.6437±0.0625a	0.0987±0.0115b	ND
	34	Kaempferol-3-O-β-D-rutinoside	0.0012±0.0002c	0.1777±0.0304b	0.4163±0.0606a	0.0077±0.0012c
	35	3,5-Dihydroxy-4',7-dimethoxyflavone	ND	1.1314±0.1865b	2.1968±0.2467a	ND
	36	3-Hydroxy-3',4'-dimethoxyflavone	ND	0.3163±0.0517b	0.4032±0.0543a	ND
	37	4'-Hydroxy-3,5,7-trimethoxyflavone	ND	0.5851±0.0726b	1.0831±0.1368a	ND
Chalcone	38	Isoliquiritigenin	0.0102±0.0015c	8.3867±1.3805b	10.5248±1.3170a	0.0369±0.0048c
Coumestan	39	Coumestrol	ND	4.1194±1.2824a	2.8179±0.6393b	ND
Pterocarpan	40	Maackian	ND	2.3782±0.4363b	3.9002±0.6101a	ND

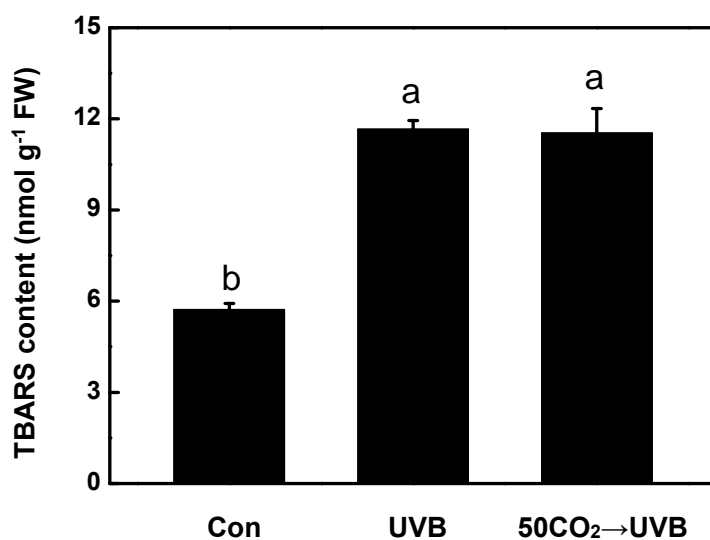
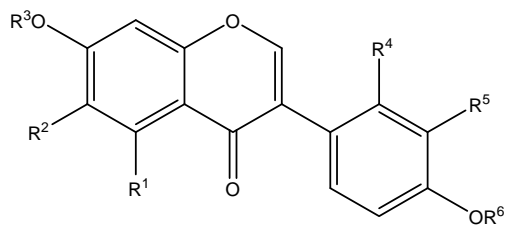
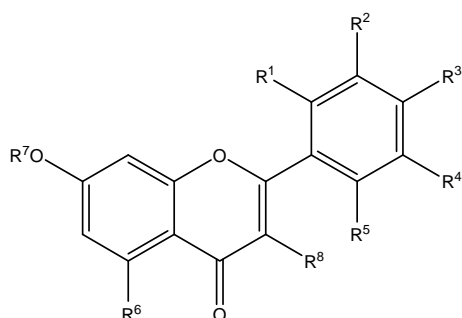


Fig. S1. Effect of CO₂ and UVB irradiation on TBARS content of alfalfa seedlings. 14-day-old seedlings were pretreated with or without 50%-saturated CO₂-rich water (50CO₂) for 12 h, and then exposed to 10.8 kJ.m⁻² UVB. After irritation, seedlings were transferred to the normal growth conditions for another 5 days. Seedlings without irradiation were set as a control (Con). Data are means ± s.e. from three independent experiments. Bars with different letters are significantly different at $P < 0.05$ according to Duncan's multiple range test.



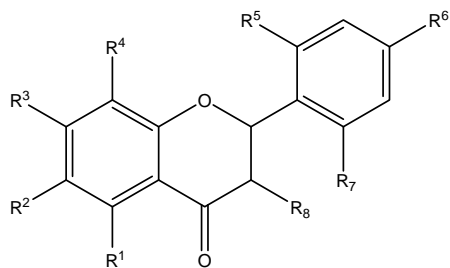
Isoflavone	R1	R2	R3	R4	R5	R6
Afromosin	H	OCH ₃	H	H	H	CH ₃
Afromosin 7-O-β-D-glucoside-malonate	H	OCH ₃	GM	H	H	CH ₃
Biochanin A	OH	H	H	H	H	H
Daidzein	H	H	H	H	H	H
Daidzin	G	H	H	H	H	H
Formononetin	H	H	H	H	H	CH ₃
Formononetin 7-O-β-D-glucoside-4"-O-malonate	H	H	GM	H	H	CH ₃
Genistein	OH	H	H	H	H	H
Genistein 5-glucoside	G	H	H	H	H	H
Irisolidone	OH	OCH ₃	H	H	H	CH ₃
Isoformononetin	H	H	CH ₃	H	H	H
Prunetin	OH	H	CH ₃	H	H	H

Fig. S2. Structures of identified isoflavone aglycones from seedling leaves of *Medicago sativa* upon UVB irradiation.



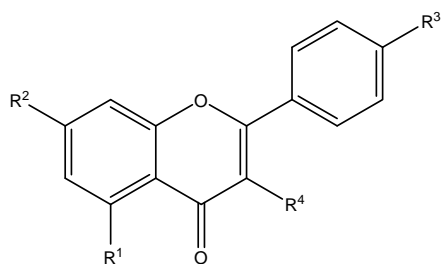
Flavone	R1	R2	R3	R4	R5	R6	R7	R8
Apigenin	H	H	H	H	H	OH	H	H
Apigenin 7-galacturonide	H	H	H	H	H	OH	G	H
Chrysoeriol	H	OCH ₃	H	H	H	OH	H	H
Luteolin	H	OH	H	H	H	OH	H	H
Millettocalyxin A	H	-OCH ₂ O-	H	H	OCH ₃	H	CH ₃	H
Milleyanaflavone	H	-OCH ₂ O-	H	H	H	H	CH ₃	H
Ptaeroxylol	OCH ₃	H	H	H	H	OH	H	H
2',5,5'-Trihydroxy-7-methoxyflavone	OH	H	H	OH	H	OH	H	H
3',4'-Dihydroxyflavone	H	OH	OH	H	H	H	H	H
3-Hydroxy-3',4'-dimethoxyflavone	H	H	OCH ₃	OCH ₃	H	H	H	OH

Fig. S3. Structures of identified flavone aglycones from seedling leaves of *Medicago sativa* upon UVB irradiation.



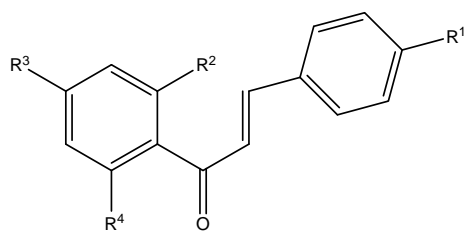
Flavanone	R1	R2	R3	R4	R5	R6	R7	R8
Betagarin	OCH ₃	-OCH ₂ O-	H	H	OCH ₃	H	H	H
Citronetin	OH	H	OH	H	H	H	OCH ₃	H
Garbanzol	H	H	OH	H	H	OH	H	OH
Isosakuranetin	OH	H	OH	H	H	OCH ₃	H	H
Liquiritigenin	H	H	OH	H	H	OH	H	H
Matteucin	OH	CH ₃	OH	CH ₃	H	H	H	OH
Naringenin	OH	H	OH	H	H	OH	H	H
Pinostrobin	OH	H	OCH ₃	H	H	H	H	H
Sakuranetin	OH	H	OCH ₃	H	H	OH	H	H
4'-Hydroxy-7-methoxy flavanone	H	H	OCH ₃	H	H	OH	H	H

Fig. S4. Structures of identified flavanone aglycones from seedling leaves of *Medicago sativa* upon UVB irradiation.



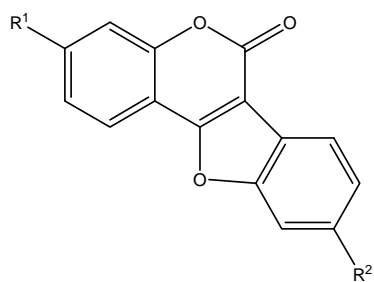
Flavonol	R1	R2	R3	R4
Kaempferol	OH	OH	OH	OH
Kaempferol-3-O-β-D-rutinoside	OH	OH	OH	OR
3,5-Dihydroxy-4',7-dimethoxyflavone	OH	OCH ₃	OCH ₃	OH
4'-Hydroxy-3,5,7-trimethoxyflavone	OCH ₃	OCH ₃	OH	OCH ₃

Fig. S5. Structures of identified flavonol aglycones from seedling leaves of *Medicago sativa* upon UVB irradiation.



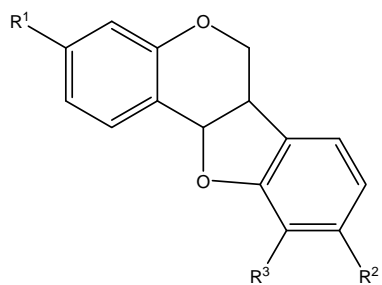
Chalcone	R1	R2	R3	R4
Isoliquiritigenin	OH	OH	OH	H

Fig. S6. Structures of identified chalcone from seedling leaves of *Medicago sativa* upon UVB irradiation.



Coumestan	R1	R2
Coumestrol	OH	OH

Fig. S7. Structures of identified coumestan from seedling leaves of *Medicago sativa* upon UVB irradiation.



Pterocarpan	R1	R2	R3
Maackian	OH	-OCH ₂ O-	

Fig. S8. Structures of identified pterocarpan from seedling leaves of *Medicago sativa* upon UVB irradiation.

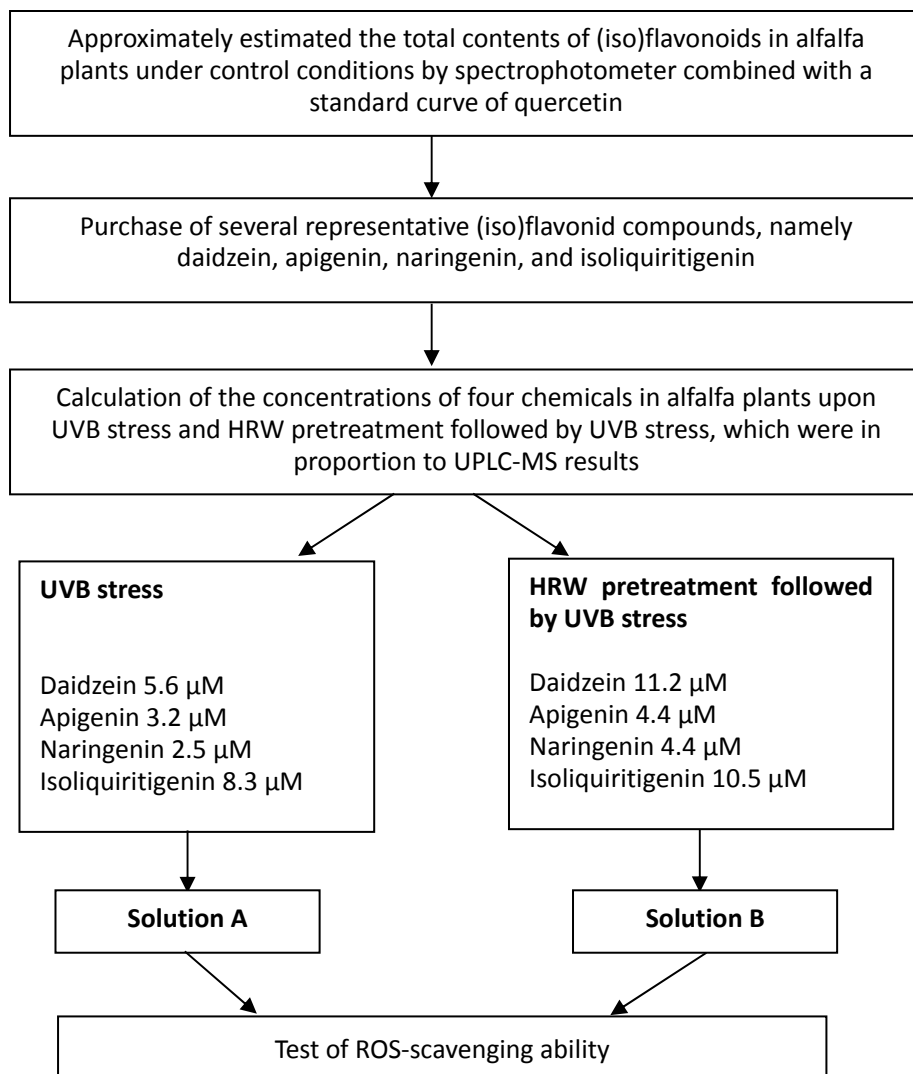


Fig. S9. Workflow for the *in vitro* test of ROS-scavenging ability.

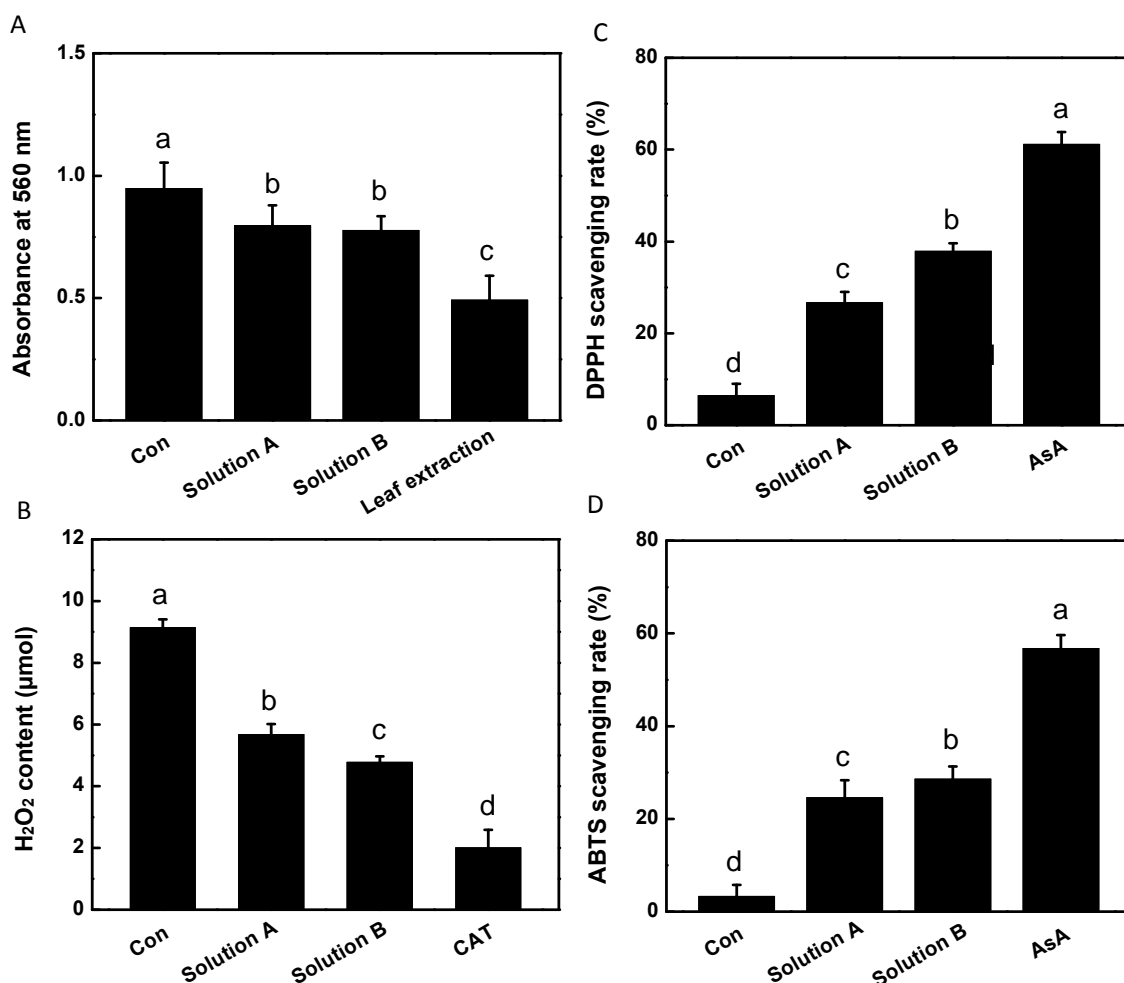


Fig. S10. *In vitro* analysis of ROS-quenching abilities (A and B) and antioxidant activities (C and D). For O₂⁻-quenching (A), sterilized water was regarded as the control sample (Con). O₂⁻ was generated by the riboflavin system under illumination, and the photochemical reduction of NBT was monitored (Absorbance at 560 nm) after 5 min of incubation. Crude enzyme extract from alfalfa leaves (leaf extraction from 5-day-old seedlings) was added as a positive control. For H₂O₂-quenching (B), the absorbance of the Fe³⁺-xylenol orange complex was determined after 45 min of incubation. The specificity of H₂O₂ was tested by eliminating H₂O₂ in the reaction mixture containing catalase alone (CAT; 100 U). *In vitro* antioxidant activities were determined by DPPH free radical-scavenging assay (C) and TEAC assay (D). Sterilized water was regarded as the control sample (Con). The AsA (50 μM) were used as a positive control. Additionally, solution A contains 5.6 μM daidzein, 3.2 μM

apigenin, 2.5 μM naringenin, and 8.3 μM isoliquiritigenin, which could partially mimic (iso)flavonoid profiles of UVB-treated samples. Solution B contains 11.2 μM daidzein, 4.4 μM apigenin, 4.4 μM naringenin, and 10.5 μM isoliquiritigenin, which could partially mimic (iso)flavonoid profiles of samples of HRW pretreatment followed by UVB stress. Data are means \pm s.e. from at least three independent experiments. Bars with different letters are significantly different at the $P < 0.05$ level according to Duncan's multiple range test.

Methods and materials

In vitro ROS-quenching abilities determination

O_2^- was generated by the riboflavin system under illumination, and the photochemical reduction of NBT was monitored (Absorbance at 560 nm) after 5 min of incubation. Crude enzyme extract from leaves (leaf extraction from 5-day-old *Medicago sativa* seedlings) were added as a positive control. H_2O_2 content was determined by detecting the absorbance of the Fe^{3+} -xylenol orange complex (Jiang *et al.* 1990). Additionally, the specificity of H_2O_2 was tested by eliminating H_2O_2 in the reaction mixture containing catalase (CAT; 100 U) alone. Measurements were performed at least in triplicate.

In vitro antioxidant activities determination

The methods of DPPH and TEAC free radical-scavenging assay are based on reaction with electron-donating or hydrogen radicals producing compounds. The DPPH free radical-scavenging activity of samples was determined according to the method described by Liu *et al.* (2009). A 0.1 mM solution of ethanolic DPPH solution was prepared. The initial absorbance of the DPPH in ethanol was measured at 517 nm and did not change throughout the period of assay. Discolorations were measured at 517 nm after the incubation for 30 min at 30 °C in the dark. In TEAC assay, the ABTS free radical-scavenging activity of each sample was determined according to the method described by Stratil *et al.* (2006). A working solution was diluted to absorbance values between 1.0 and 1.5 AU at 734 nm with phosphate buffer solution. Standards or samples (from 5 to 25 μ L according to reaction intensity) were mixed with the working solution (975 μ L) and diluted up to 1000 μ L with deionized water. A decrease of absorbance was measured at 734 nm after 20 min. Aqueous phosphate buffer solution (1 mL, without ABTS^{•+} solution) was used as a control. Measurements were performed at least in triplicate.

References

- Jiang ZY, Woollard ACS, Wolff SP (1990) Hydrogen peroxide production during experimental protein glycation. *FEBS Letters* **268**, 69–71.
doi:10.1016/0014-5793(90)80974-N
- Liu L, Sun Y, Laura T, Liang XF, Ye H, Zeng XX (2009) Determination of polyphenolic content and antioxidant activity of kudingcha made from Ilex kudingcha CJ Tseng. *Food Chemistry* **112**, 35–41.
doi:10.1016/j.foodchem.2008.05.038
- Stratil P, Klejdus B, Kuban V (2006) Determination of total content of phenolic compounds and their antioxidant activity in vegetables evaluation of spectrophotometric methods. *Journal of Agricultural and Food Chemistry* **54**, 607–616. doi:10.1021/jf052334j